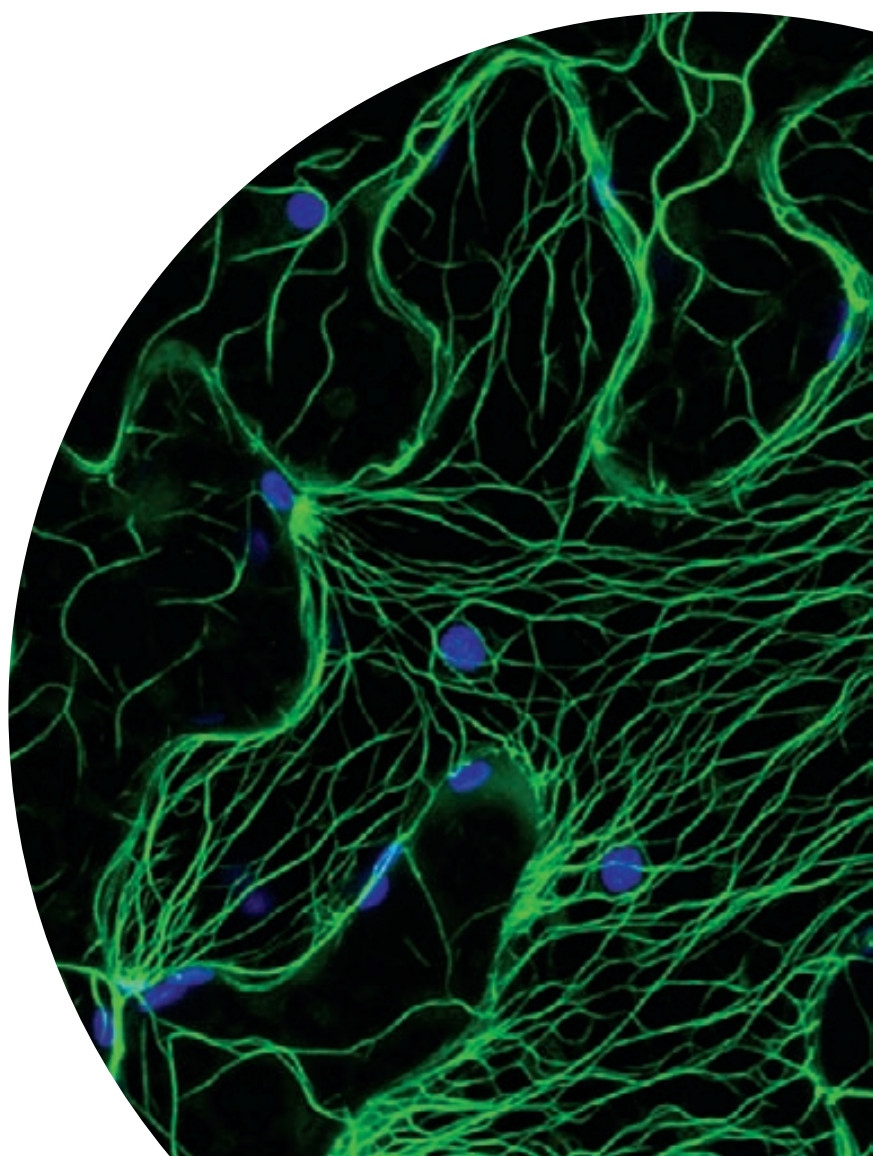


## Annual Report 2009



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**Cover Illustration:** The Cell Biology & Imaging Group at SCRI has built up a library of transgenic plants that express stable, fluorescent markers in different cellular locations. This valuable resource lets us identify proteins of interest with organelle markers and work out likely functions based on their location. In the cover illustration, green fluorescent protein decorates actin filaments in a leaf epidermal cell while chloroplasts are shown in blue.



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# Introduction

**Peter J. Gregory**

Although food prices in the shops and cereal prices on the world commodity markets have fallen from their highs in mid 2008, the wake up call to governments around the world has continued to foster interest in the issue of food security and associated concerns around water and energy supplies and use. During the year,

we have been actively engaged in numerous workshops and discussions organised by the UK and Scottish governments, the UK research councils and internationally in meetings in Australia, Austria and India, and as part of the Copenhagen climate change talks on the subject of food security. The renewed importance of the topic is evident in the new strategy

of the Biotechnology and Biological Sciences Research Council (BBSRC) in which it is one of three major themes along with bioenergy and healthy food. More

locally, the Scottish Government, too, is engaged with interested parties in determining the research required to give substance to its Food and Drink Policy published in mid-2009 which aims to promote sustainable economic growth through work with Scotland's food and drink industry to address quality, health, wellbeing,

environmental sustainability and the need for access and affordability. All of this debate highlights the importance of the research undertaken by us, and it has been very pleasing to see this recognised in both the various surveys of UK national capability in agricultural research and the reopening of BBSRC funding to us facilitated by the Scottish Government.

These small

individual steps will allow us to play a significant role in the emerging UK research programmes on issues related to food security.



Peter Gregory

In this context, the 'in principle' decision of our Governing Board and that of the Macaulay Land Use Research Institute (MLURI) to merge the two institutes from April 2011 is of considerable significance. The merger will bring together about 200 senior research scientists plus their support staff into a single institution capable of undertaking research across multiple disciplines and scales. The New Institute (the name has yet to be decided) will have a vision that encompasses 'excellence with delivery', a remit that has global reach, and a research focus on the environment/plant/land continuum and the uses that society makes of it. It will combine scientific and research excellence with delivery of knowledge, products and services to international, national and local customers. As with both SCRI and MLURI, the New Institute will be mission oriented and its research will feed directly into long term societal concerns relating to sustainable economic development, food/energy/water securities, environmental change, biodiversity, waste reduction and community development and wellbeing. Both institutions believe that by coming together at this time we can create a world leading institution that will be able to address these challenging issues in a way that, individually, would be impossible. As the economic recession starts to be felt in the public sector, we think that this timely initiative will place the New Institute in a position to attract scientists of the highest international quality and to broker access to the best minds and research in other organisations by providing global leadership in a broad range of land based disciplines.



SCRI Director Professor Peter Gregory interviewed by Radio Tay as plans for the 'New Institute' are announced.

Obviously substantial work remains to be done to make the New Institute a reality, and much of 2010 will be focused to this end. Already, though, agents have been appointed to recruit a Chairman and Chief Executive for the new company and we have held a joint meeting of the science programme leaders to familiarise ourselves with each other's research interests and to explore potential areas of synergy. Richard Aspinall (the Chief Executive of MLURI) and I have been working together on the science vision and on the business plan with the Scottish Government and consultants from Arthur D Little who assisted the Boards in making their decisions. We anticipate that the New Institute will play a leading role in the food security and multifunctional land use debates currently exercising governments and policy bodies.



The 2009 SCRI Director's summer soirée.

2009 marked the 20<sup>th</sup> birthday of Mylnfield Research Services Ltd (MRS Ltd) which was celebrated as part of the Director's summer soirée in July. MRS Ltd has continued to develop profitable business despite the recession, and we were very pleased to be able to appoint two new field geneticists who will undertake research in modern molecular genetics while receiving hands-on training in breeding of potatoes and barley. We hope that these 'trainees' will form the next generation of breeders, and so keep alive a skill that is in danger of disappearing from public sector research institutions. An allied activity has been the launch in October by our colleagues in the Division of Plant Sciences, University of Dundee of a Master of Research (MRes) course in Crops for the Future. Staff from the Institute are contributing to the course modules, but we hope that the project element of the course will also contribute to our research





activities and to our breeding programmes. MRS Ltd has successfully renegotiated the raspberry breeding contract with a consortium of growers, the Horticultural Development Company and the Scottish Government and we look forward to new varieties flowing from that work.

Our research continues to produce new knowledge and insights into the workings of plants, their pests and pathogens and interactions with the environment. In our Genetics programme a highlight of the year has been the public release of a draft genome sequence for potato together with international colleagues from around the world. In barley, a gene controlling fertility of the lateral florets in barley spikes has been identified, and with colleagues from the Division of Plant Sciences we have shown that the flowering regulator, FPA, functions in messenger RNA 3' end formation and that the nucleolus has a novel role in recognition and turnover of aberrant mRNAs. Software (Flapjack and Tablet) for visualising dense molecular genotype information and second generation sequence data respectively, has been developed by our informatics team and has been widely adopted by the international genetics community. Meanwhile staff in Plant Products and Food Quality have used a combined metabolomic, transcriptomic and sensory analytical approach to identify the key volatile components driving desirable potato aroma and texture, and the genes underpinning biosynthesis of the associated components. Collaborative research with partners in the UK and Europe has shown that berries such as strawberries and raspberries can inhibit pancreatic lipase. This enzyme controls fat digestion and associated calorie intake in humans and opens avenues for the dietary management of obesity and related degenerative diseases.

In Environment Plant Interactions, there has been a particular push to develop novel methods for studying root system development in natural substrates using x-ray tomography (with the University of Abertay, Dundee), rhizotron image capture and structured mesocosm techniques, combined with image tracking approaches. Results from these techniques are being used to inform mathematical models predicting how root architecture affects the acquisition of water and

mineral elements from the soil, and how this might be manipulated for improved resource capture and ecosystem sustainability. As ever, our Plant Pathology programme has been very active and we were part of an international consortium that published the full genome sequence of *Phytophthora infestans*, the causal agent of the damaging late blight disease of potato. Availability of the genome sequence has had immediate effect in enabling a large number of pathogen effector molecules (pathogen encoded factors important in the disease process) such as transcription factors and proteins involved in secretory, signalling and ubiquitination pathways to be identified. These factors are being used to develop 'smart screens' to identify durable resistance in potato germplasm. In related work, we have used molecular markers to analyse isolates of *P. infestans* from >1000 late blight outbreaks in the UK and Europe. In the UK, data from >5000 isolates analysed in recent years show that populations of *P. infestans* are currently dominated by one genotype (designated 13\_A2). This aggressive genotype is causing concern as it has overcome the resistance of some previously resistant commercial cultivars such as our own variety Lady Balfour. Work is ongoing to understand the biological and genetic properties that make isolates of genotype 13\_A2 so successful.

January saw the launch of the BBSRC Sustainable Bioenergy Centre in which the Division of Plant Sciences of the University of Dundee, along with SCRI, plays a major part. The Dundee work will concentrate on altering lignin production in barley to make it easier to produce bioenergy from straw without reducing the quality of the grain crop. This research is an essential step in the development of second generation biofuels.

The high quality of our research has continued to be attested to by the external reviews undertaken by our Governing Board. In May it was the turn of BioSS to be reviewed. The team concluded that "BioSS is a first-rate organisation which is delivering international level research in fundamental science as well as statistics and bioinformatics methodology." This ringing endorsement of their work was much appreciated. The turn of Environment Plant Interactions came in November and again the programme received praise for its quality and





Dr Keith Dawson (left) receives a farewell gift from SCRI Chairman Peter Berry as he retires from the Governing Board.

the enthusiasm of the staff. Changes on the Governing Board this year were limited to the departure of Keith Dawson after eight years of substantial service during which he worked tirelessly for the promotion of the Institute, especially alongside the crop based skills of SAC (Scottish Agricultural College). Andrew Wilson joined the Board; his interests in communication and knowledge of politics and commerce in Scotland will be valuable to us.

We were very pleased at the award of the Jones-Bateman Cup of the Royal Horticultural Society to Julie Graham for her work on the genetics of soft fruit; this is vital underpinning research for our commercial breeding

of new raspberry cultivars. The Director's Award for 2009 was made to Ian Pitkethly for his very substantial contributions to our publications, scientific posters and general communications. He will always go the extra distance to deliver high quality products against very tight deadlines, and this is much appreciated by a wide range of staff.



Dr Julie Graham of the Genetics programme is presented with the prestigious Jones-Bateman Cup by Giles Coode-Adams, President of the Royal Horticultural Society (RHS). Julie was honoured for her work on raspberry breeding.

I hope that you will enjoy reading about the rest of our activities in 2009 and learn something of the exciting research that we are engaged in to underpin food security.



# Environmental Change

**Lesley Torrance**

*Our society faces serious threats posed by environmental change and increases in global population which will impact on food, energy and water security. The UK Climate Impacts Programme predicts that in the next 75 years Britain will become warmer, with drier summers and wetter winters and greater volatility resulting in unpredictable extreme rainfall and temperature events. Innovative solutions are required to adapt to, or mitigate, these effects and to promote environmentally and socially sustainable economic growth. Some of these effects are already apparent in Scotland; for example, this year we have experienced the wettest November in 50 years with other indicators such as earlier aphid migrations.*

At SCRI we are conducting research to identify and counter risks from environmental change and develop solutions that will sustain the competitiveness of our agricultural industry while maintaining a vibrant and economically successful rural environment. The articles

in this section illustrate the work we are doing on plant pathology, physiology and genetics to study the effects of temperature and water availability and increased pest and disease risks on Scottish crops.

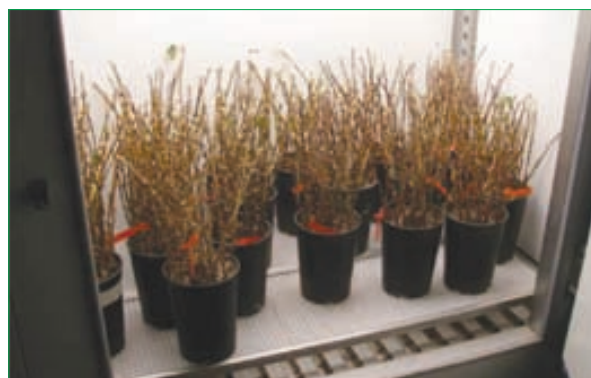


## Dormancy in blackcurrant and the potential effects of future climatic conditions

**Rex M. Brennan, Hamlyn G. Jones\*, Joanne R. Russell, Peter E. Hedley, Linzi Jorgensen, Chris A. Hackett & Sandra Gordon**

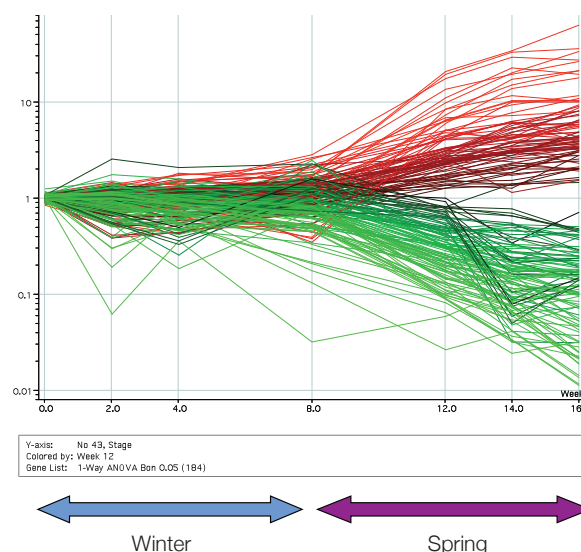
One of the key environmental factors affecting cropping of woody fruit species is winter temperature, through its effect on the dormancy cycle. The degree of chilling received by the plants during the dormant period, to fulfill a chilling requirement that varies between species and cultivars, has profound effects on budbreak, flowering and ultimately fruit quality. In the UK, a study of historical data using a range of models for assessment of winter temperatures indicated a decline in levels of winter chilling over the past 50 years. Projected future increases in temperature suggest that in northern latitudes these will be proportionately greater in winter.

Blackcurrant is grown widely across northern temperate regions of Europe and also in New Zealand for commercial processing. UK production is based entirely on the SCRI 'Ben' series, from the earliest release 'Ben Lomond' in 1972 to the more recent 'Ben Starav' and 'Ben Klibreck' (2008). Blackcurrant has a relatively high chilling requirement, ranging from <1300 h below 7.2°C for some New Zealand cultivars to over 2000 h for some late-flowering Scottish types such as 'Ben Lomond'. Plantations of high-chill cultivars in southern locations, such as Kent and Herefordshire which have received insufficient chilling during recent UK winters, have suffered erratic budbreak and a subsequent reduction in harvested quality.



**Figure 1** Phenotyping of blackcurrant cuttings for chilling requirement.

The response of blackcurrant genotypes to chilling temperatures was examined using controlled environment facilities at SCRI (Fig. 1). Hardwood cuttings of diverse genotypes were treated at chilling temperatures for periods between 5 and 21 weeks, after which the cuttings were moved to forcing environments so that budbreak and flowering could be assessed. The data were analysed using novel curve-fitting approaches, from which it was established that the various genotypes had very diverse chilling responses, with some cultivars, for example 'Ben Avon', responding only slowly to chilling temperatures while others, for example the French 'Andega', responding much more rapidly. Additionally, the calculation of an effectiveness coefficient for the different temperatures showed that the most effective chilling temperature varied between genotypes. Using this information, a high throughput method of phenotyping blackcurrant breeding progenies and advanced selections is under development, to assess their chilling requirement and enable the selection of environmentally resilient 'Ben' cultivars into the future. Collaborations with Plant and Food NZ are facilitating the use of low chill germplasm in the development of new progenies, and further investigation of climate effects on blackcurrant dormancy and development is planned across Scandinavia and northern Europe as part of the 'Climafruit' project in the EU Interreg North Sea programme.



**Figure 2** Microarray analysis of blackcurrant buds at dormancy break showing differential expression of genes.

\* University of Dundee





At the molecular level, putative Quantitative Trait Loci (QTL) linked to budbreak and flowering traits have been identified on the genetic linkage map for blackcurrant, and further analysis is in progress using an extended reference population. Dormancy break and subsequent development is controlled through the coordinated action of large numbers of genes in woody plants. Differential gene expression during dormancy and budbreak in blackcurrant has been investigated (Fig. 2), and key genes associated with budbreak have been identified and mapped on the blackcurrant linkage map. Initial steps have been taken to develop associated markers that will then be deployed in the characterisation of the diverse *Ribes* germplasm in downstream breeding and mapping populations. The information on candidate genes will also have relevance to other woody species apart from blackcurrant.

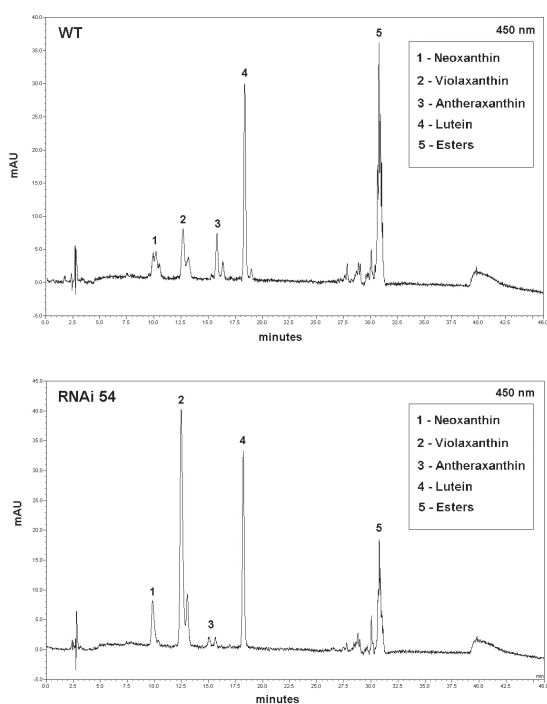
### The role of carotenoid cleavage products in heat responses of potato

**Raymond Campbell, Laurence Ducreux, Wayne L. Morris, Peter E. Hedley, Glenn J. Bryan, Gavin Ramsay & Mark A. Taylor**

The cultivated potato exists in a variety of forms originating from diverse environments such as coastal areas in Chile and dry Andean valleys at high altitude. The types finding favour across the world, including tropical and sub-tropical areas, are often those with high yield potential, bred in Europe and North America from a stock with its ultimate origins primarily in Chilean material and poorly adapted to high day and night temperatures. In fact most potato cultivars bred in temperate climates respond to high day and night temperatures with a reduced number of stolons (underground stems that form tubers) and tubers, and altered carbon partitioning to the tuber,



**Figure 3** The *CCD4* silenced phenotype was characterised by early sprouting during tuber development or by the formation of chain tubers. These effects were observed in 20 out of 33 independent RNAi lines.



**Figure 4** HPLC analysis of tuber carotenoid content in a *CCD4* silenced tuber compared with wild type (WT). Note the large increase in the violaxanthin peak in the *CCD4* silenced tuber.

resulting in a significant loss of tuber yield and quality. Climate changes due to global warming are expected to similarly affect the potato crop with warmer winters and summers predicted to increase abiotic stresses. Temperature fluctuation also causes economically significant damage to the potato crop. Growth under cool temperature interrupted by periods of high temperature leads to heat sprouting, chain tubers and secondary growth of tubers. This has serious effects on economic yield and tuber quality. These effects are due to a fluctuation in the level of stimulus that causes tuber formation to alternate with more stolon-like growth. This physiological problem can be a serious problem for some cultivars exported to or trialled in Mediterranean climates, and in the longer term may also affect tuber production in the UK.

The mechanism of heat sensing in plants is not fully understood, however it is known that the potato circadian clock can integrate environmental signals and thus modulate the level of tuberisation stimulus. It is also becoming apparent that signals derived from carotenoid pigments, by the action of carotenoid cleavage dioxygenases (CCDs), are potent regulators of plant growth and development. The classic example is abscisic acid, a well characterised carotenoid derived plant growth regulator. More recently it has been demonstrated that other members of the CCD family are involved in the biosynthesis of strigolactones, a newly discovered class of plant growth regulator that control plant branch structure. At SCRI we silenced the expression of the potato *CCD4* gene and this resulted in a dramatic phenotypic change. Tubers from silenced plants exhibited a heat sprouting phenotype even though they were not exposed to heat stress (Fig. 3). The phenotype was even apparent in tubers grown under *in vitro* conditions. Analysis of carotenoid content in the silenced tubers revealed that the carotenoid violaxanthin was the likely substrate for the *CCD4* as its level was elevated several fold in the silenced lines (Fig. 4). There is no evidence for changes in abscisic acid or strigolactones in these plants. We are currently investigating the nature of the carotenoid derived signal that is implicated in temperature perception by these observations. Additionally we are investigating





whether elevated levels of *CCD4* expression can result in protection from heat stress and increased tuber yields under fluctuating temperature conditions, thereby identifying a clear gene target for future potato breeding strategies.

### Root traits for a changing physical environment

**Tracy A. Valentine, A. Glyn Bengough, Paul D. Hallett & Blair M. McKenzie**

With increasing constraints on the use of fertilisers and irrigation water worldwide, crops need root systems better able to grow in soils containing limited water and nutrient resources. Plants must be able to place their roots close to where these resources are located in the soil profile. We have been studying how plant genotype influences the growth and distribution of its roots, at

scales from the laboratory to the field, with a view to developing better ways of identifying, and ultimately screening for, desirable root traits. Another equally important aspect is evaluating the physical restrictions to root growth that occur in a range of soil types, so that factors limiting root growth can be quantified and used to identify the most important physical restrictions throughout a field season. This information will be used to develop improved soil management practices and the breeding of plant varieties suited to particular soil conditions.

In a recent survey of more than 50 farms in the east of Scotland we found that, even when soil water availability was close to optimal, nearly 40% of soils had strength sufficient to mechanically impede root growth. Seedling screens confirmed that soil physical conditions were limiting root elongation to less than half of the unimpeded rate in a similar percentage of sites examined. The impact of changes in climate and

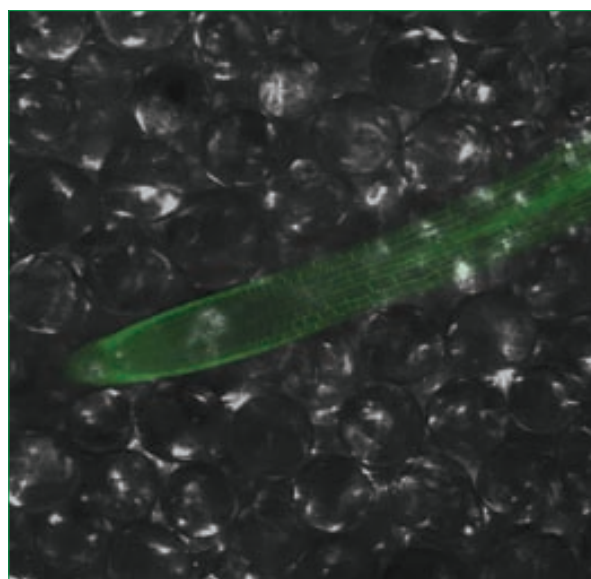




**Figure 5** The influence of genotype and interaction with the environment in the ability of barley plants to access subsoil water, leading to differences in plant height and potentially crop productivity.

farming on plant and crop performance will depend not only on the severity of these physical limitations to roots, but also on the frequency, duration, and timing of these limitations. Rainfall patterns are predicted to become more erratic due to climate change and may result in more frequently waterlogged soil while at other times periods of drought may result in increased soil strength. Changes in farming such as decreasing tillage to save fuel may adversely affect soil conditions. Soil compaction, particularly of the subsoil, could be exacerbating the limitations to root growth.

We have developed techniques to study the spatial distribution of the roots of both seedlings and more mature plants. The angular spread of seedling cereal roots, measured in two dimensions using a gel plate screen developed at SCRI has been found to relate



**Figure 6** Root tip behaviour under physically inhibiting environment: Live imaging techniques of plant–environment interactions in a model system using fluorescently labelled roots and glass beads.

closely to root spread and water extraction by mature drought tolerant cereal plants. The angular spread of roots in two dimensions from a range of contrasting genotypes was also found, by X-ray tomography, to relate closely to root spread in three dimensions. We have also developed a field method for screening the ability of crop genotypes to access subsoil water, and this is demonstrating both the influence of genotype on this root trait and the importance of deep root growth even relatively early in the growth of spring sown barley (Fig. 5). The importance of root tip traits in the ability of roots to overcome soil physical restrictions is being explored at the individual plant and individual root tip scales using live imaging techniques (Fig. 6) and by linking to root growth models of soil exploration. Mechanisms used by roots to overcome stress, such as producing a lubricating layer or changing the properties of soil, are a major component of our research in this area.

## Risks of new or emerging pests and pathogens posed by environmental change

**Lesley Torrance, Ian K. Toth, David E.L. Cooke, Brian Fenton, Vivian C. Blok & Alison K. Lees**

Environmental factors greatly influence the incidence and geographical distribution of pests and diseases. The availability of a susceptible host together with optimum conditions of temperature and humidity are major factors in determining whether a pathogen can establish in a new location. Adaptation to the predicted environmental changes will affect Scottish agronomic practices as well as the spectrum of non-crop species grown. Some examples of the likely changes in pests and disease threats to potato production in Scotland and the research being done to counter these threats are discussed.

Virus diseases are the major cause of seed potato degeneration causing down-grading of seed crops. Aphids transmit viruses from plant to plant (Fig. 7) and transmission early in the season has a major effect on virus incidence in the harvested tubers. Scotland has an effective aphid monitoring programme and this



has shown that some of the main virus vector aphids are appearing earlier, when they could be spreading viruses. If this trend continues it may change the areas of Scotland with a natural advantage for production of healthy seed potatoes and require earlier applications of insecticides. In collaboration with industry we are conducting research to establish effective aphid-



Figure 7 Aphid virus vector.

borne virus control strategies. However, the most effective long term disease control is achieved through resistant crops and we have found a new source of extreme resistance which is effective against three important aphid transmitted Potyviruses (PVY, PVA, PVV). Current research is focused on developing molecular markers which can be used to deploy the resistance in breeding programmes. *Phytophthora infestans*, the cause of late blight, has been shown to be a highly adaptable pathogen that can respond rapidly to changing environmental conditions and overcome major host resistance genes. Late blight will pose a continuing threat to Scottish crops. Aided by the recently published whole genome sequence, we are making major advances in understanding the function and evolution of *P. infestans* pathogenicity factors. We are also generating valuable data on the response of the current pathogen populations to moisture and temperature criteria which are relevant to current and future prediction and control of late blight in Scottish crops. Adaptation of the potato cyst nematode (PCN)





to environmental changes and the potential for higher multiplication rates are also being examined. The risks of hatching and completion of a second generation

during the growing season and the potential for selection of populations with faster generation times are being assessed in relation to integrated management of PCN.

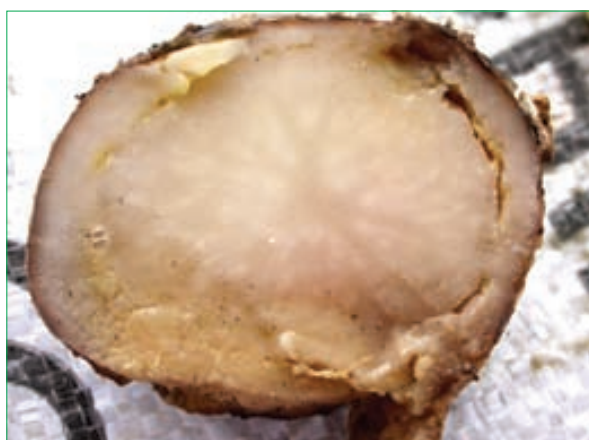


Figure 8 *Dickeya solani* on ware potatoes cv Agria imported to UK from Spain. Tubers show Cheesy Rot and breakdown of the vascular ring (very similar to that of Ring Rot). Picture supplied by Eric Anderson (Scottish Agronomy)



Figure 9 *P. kernoviae* infection of rhododendron.

Threats from non-indigenous diseases include the introduction by contamination of infected planting material, e.g., the bacteria *Dickeya dianthicola* and *D. solani* (formerly *Erwinia chrysanthemi*) on potato tubers (Fig. 8) and the nematode *Meloidogyne chitwoodi* in adherent soil particles. These pests and pathogens have been intercepted on potato seed tubers and are currently responsible for major disease losses in Europe. We have rapid and sensitive diagnostics to detect *M. chitwoodi* and are working with SASA (Science and Advice for Scottish Agriculture) and FERA (The Food and Environment Research Agency) to develop similar diagnostics for *Dickeya*. These diagnostics are important for monitoring the movement of these pests and pathogens, and complement our work on the biology of the diseases they cause.

Environmental change will also alter the spectrum of plants growing in natural, semi-natural ecosystems and gardens in Scotland. Associated threats from invasive pathogens are thus also being studied. In a collaborative project on *P. ramorum* and *P. kernoviae* (Fig. 9) knowledge and molecular markers derived from research on *P. infestans* are being used to assess the risks posed to Scottish heathland ecosystems.

These examples are just a few of the many threats that will face agricultural production systems now and in the future. Our work provides robust evidence and analysis to advise policy makers in Scottish Government and the private sector to help combat these threats. An outcome of this work suggests that a re-evaluation of the current operation of the 'Safe Haven' certification scheme to protect Scottish seed potatoes is worthwhile.



# Biodiversity

**Robbie Waugh**

*Biodiversity describes the variation of life forms that exist within a population, a species, an ecosystem or a geographical region. Measures of biodiversity are frequently used to assess the health of a 'biological system' and although commonly associated with conservation biology, biodiversity research has broad application in plant and environmental sciences. It provides a framework for understanding and quantifying the molecular variation in individual genes and the richness of species in complex ecosystems, and may guide management interventions to maintain or enhance the viability of both species and communities. Biodiversity is the driving force of evolution, and its efficient exploitation is the basis of crop plant improvement. Not surprisingly then, across SCRI, considerable energy is being expended on the quantification and interpretation of biodiversity to address a wide range of biological questions.*

Here we provide four examples of biodiversity research being conducted across the research programmes. In the first, Joanne Russell and colleagues show how a clear relationship exists between genetic distances and geographic distances among populations of barley landraces and discuss how these observations shape our understanding of the domestication and evolution of the species, with potential implications for plant breeding. By walking through an example project, in the second, Iain Milne and colleagues illustrate how the

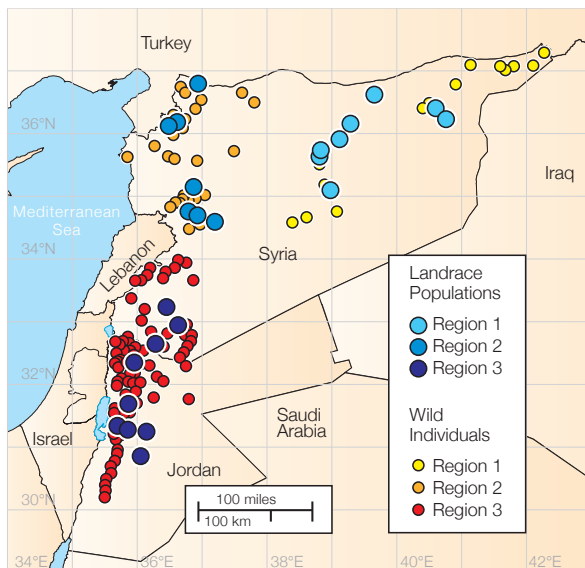
visualisation tools they have developed have become essential components in the analysis of highly complex molecular diversity datasets. Ali Karley and colleagues then describe how interactions between insects and microbes can shape interactions between herbivores and other components of an agroecosystem while, in the final contribution, the importance of understanding pest and pathogen diversity is described by Alison Lees and colleagues. These serve to illustrate the breadth of activity being conducted under the biodiversity banner.



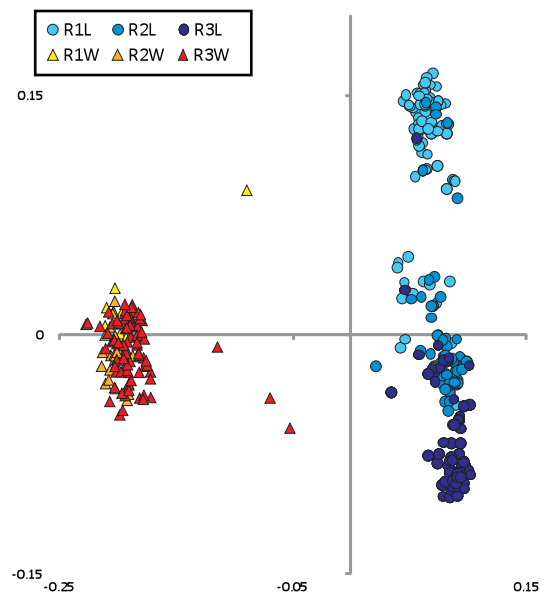
## Relationship between genetic distances and geographic distances among populations of barley landraces

Joanne R. Russell, Ian K. Dawson, Allan Booth, Andy J. Flavell<sup>1</sup>, Brian J. Steffenson<sup>2</sup>, Eva Weltzien<sup>3</sup>, Salvatore Ceccarelli<sup>4</sup> & Robbie Waugh

Within barley an enormous amount of natural diversity exists, which has been shaped by the processes of domestication, cultivation and breeding. Different views have been proposed about the domestication of barley, with some authors stressing a single origin in the Fertile Crescent and others suggesting more complex domestication events. One of the most effective means of understanding the origins of domestication and cultivation is by comparative sampling of wild and cultivated accessions from regions where their ranges overlap. We have assembled and studied a collection of approximately 500 stratified, geographically referenced and matched samples of landrace (cultivated) and wild barley from across Jordan and Syria (Fig. 1). We used a high resolution genotyping platform that provides accurate measures of molecular biodiversity across the entire barley genome to explore the origins of barley cultivation. We observed clear



**Figure 1** Geographic locations of sites in Jordan and Syria from which landrace and wild barley accessions were sampled for an analysis of SNP variation. Twenty-four landrace populations, comprising 317 accessions in total and 131 individual wild plants differentiated into three geographical regions: Region 1, North-eastern Syria; Region 2, North-western Syria; Region 3, Jordan and southern Syria.



**Figure 2** Principal coordinate analysis for 1135 SNP loci in landrace ( $N = 317$ ) and wild ( $N = 131$ ) barley accessions sampled from Jordan and Syria. Landrace and wild accessions were clearly differentiated, except for four 'wild' individuals that placed unusually in ordination. The first principal axis (horizontal) accounted for 35% of all variation, the second principal axis (vertical) for 13%. For information on sample designations see Fig. 1.

evidence for geographic structuring of genetic variation in both wild and landrace barley (Fig. 2). Differentiation was however less pronounced in wild barley. Three geographical regions were clearly identified and cross comparisons indicated that landrace populations from all three regions were most similar to wild material from Jordan and south Syria. An important observation is that landrace and wild accessions were 'not-matched' in north-western and north-eastern Syria, as would have been anticipated if cultivated material was of local origin. At the same time, a high degree of structuring in landrace accessions suggests that these cultivated stands have been present at their current locations for a considerable period of time. This distinctness has been maintained in spite of human exchange of germplasm that could potentially swamp any pre-existing landraces and suggests strong pressure towards local adaptation to varying environmental conditions in the two countries.

Gene flow between cultivated and wild populations after domestication has been suggested as an important mechanism for adaptation in barley and has been

<sup>1</sup>University of Dundee, <sup>2</sup>University of Minnesota, <sup>3</sup>ICRISAT, <sup>4</sup>ICARDA



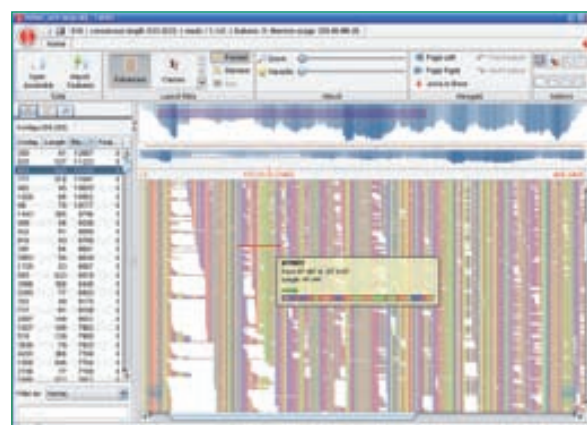
proposed to be relatively common. Our analysis highlighted four unusual 'wild' individuals consistent with minor directional interaction between wild and landrace barley (Fig. 2). However overall, landrace and wild material act essentially as independent entities despite geographic overlap. This may reflect variation in flowering time and maturity, with the brittle rachis of wild barley meaning that in mixed stands wild heads will shatter and seed be dispersed before the cultivated seed is collected. This research provides insight into the process of domestication of barley in the Fertile Crescent. Comparisons of the observed patterns of molecular diversity in this material with that in advanced cultivars will help identify chromosomal regions that have been subject to human and environmental selection during the process of domestication.

### Visualising genetic diversity

**Iain Milne, Paul Shaw, Linda Milne, Micha Bayer, Gordon Stephen & David F. Marshall**

The increasingly complex data sets generated through new DNA sequencing and genotyping technologies, such as described above for barley, require a new generation of software tools to aid analysis and interrogation. The sheer volume of data imposes limitations on our ability to inspect the results of analyses and look for patterns that reflect either quality control issues or biologically meaningful structure in the data. Therefore, to support and enable analyses of genetic and genomic data we have developed a suite of computational visualisation and analysis tools. To illustrate their value, in this report we follow a typical project that starts with the discovery of single nucleotide polymorphism (SNP) markers, is followed by their use for genotyping to assess patterns of diversity in germplasm collections, and ends with comparative genomics – an approach used to identify putative regional gene content.

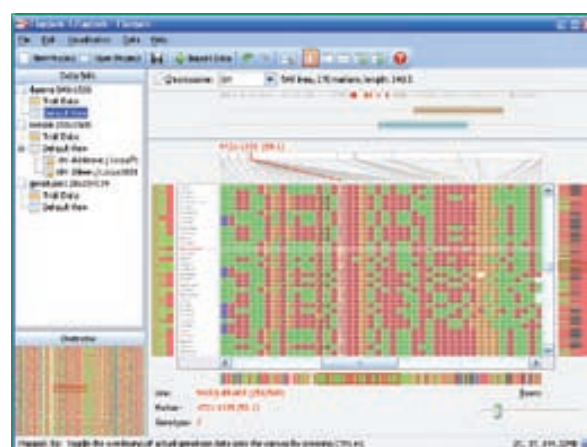
The process starts with using second generation sequencing technologies to sequence the transcriptome from two or more plant lines from any plant species. The output of this exercise, more than 1,000,000 individual sequences, are either aligned against an existing sequence (such as an Expressed Sequence



**Figure 3** Tablet displaying an assembly of singletons that are left over from a Mosaik mapping of barley read using Solexa technology from seven different barley lines.

Tag assembly) or assembled (i.e. joined together into longer contiguous sequences) *de novo*. The output may result in more than 50,000 assembled transcripts, which can be analysed for polymorphic nucleotide sites that are then 'marked up' as potential SNPs. These assemblies can be visualised with Tablet (Fig. 3), software we specially designed to cope with this scale of data, allowing us to view and navigate the millions of aligned DNA fragments and identify which SNPs are most suitable for translation into high-throughput SNP genotyping assays.

The resulting avalanche of high-quality genotypic data presents us with new challenges in data maintenance and analysis. In collaboration with colleagues at the University of Dundee we are developing a new implementation of our existing marker database



**Figure 4** 160 SNPs from chromosome 6H of barley are displayed within Flapjack, where multiple graphical views allow us to visualise the associated genotype, phenotype and QTL data simultaneously.

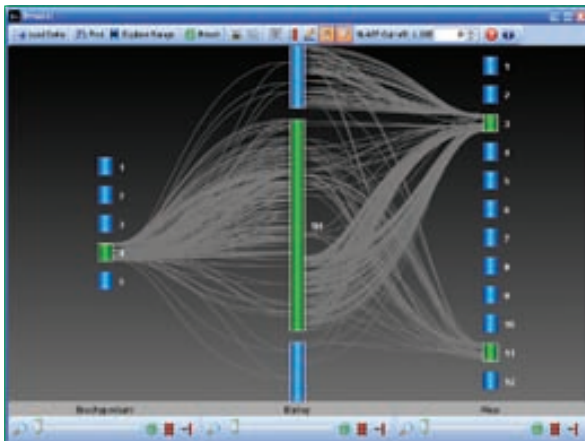


Figure 5 *Strudel* rendering homologous mappings between high-density linkage maps across multiple genomes of *Brachypodium*, barley and rice.

infrastructure, *Germinate 2*, to cope with these demands. *Germinate 2* allows us to store, query and visualise a broad range of associated information, including passport data, trait information, pedigrees and the results of analyses. We developed *Flapjack* (Fig. 4) – a revolutionary new graphical-genotyping and manipulation tool – to visualise the genotype data from many thousands of lines at many thousands of SNPs, and we are working with collaborators around the world

to ensure that *Flapjack* is compatible with major new genotypic data sets from maize, rice and *Arabidopsis* as well as genetic analysis and simulation packages.

High-throughput genotyping technology enables us to rapidly identify key genomic regions that control agriculturally important traits through either conventional Quantitative Trait Loci (QTL) analysis or association analysis. But we want to identify genes – not regions. Comparative mapping is facilitated by our visualisation tool *Strudel* (Fig. 5), which enables the rapid identification of the gene content of orthologous genomic regions (derived from a common ancestor) across multiple species. In addition, *Hordeum Relator* visualises the relationships between DNA sequences from various species related to barley, including rice, *Brachypodium*, oat and sorghum. Based on this set of integrated databases, web resources, and Java desktop applications, our software – although primarily developed to meet the needs of ourselves and our collaborators – is made freely available to all, and is used by researchers in over 40 countries around the world.





## Insect herbivore–microbe interactions: impacts on insect fitness and behaviour

Ali J. Karley, Scott N. Johnson, Emily Clark & Lindsay S. McMenemy

Insect herbivores are key components of agroecosystems, both as consumers of crop and non-crop plants and as a food source and host for a wide range of natural enemies. Interactions between insects and microbes often shape how insect herbivores interact with other parts of the system. In particular, microbes are frequently harboured or transmitted by insect herbivores and fulfil a range of roles as symbionts, parasites or pathogens. We study the biology of plant sap-feeding aphids to unravel the complex interactions between insect herbivores, their bacterial symbionts and the plant viruses that they transmit, to determine the role of microbes in regulating insect populations in agroecosystems.

Many herbivorous insects possess microbes known as symbionts that live within the insect and perform vital functions that are essential for insect survival. In aphids, most species harbour a ‘primary’ bacterial symbiont, *Buchnera aphidicola*, which lives in specialised organs within aphid tissues and produces amino acids that the aphid cannot produce. Aphids can also harbour one or more types of ‘secondary’ bacterial symbiont whose functions are less certain. There is increasing evidence that these secondary bacteria play a significant role in aphid interactions with other organisms, particularly by altering aphid susceptibility to natural enemy attack. In the Cabbage aphid (*Brevicoryne brassicae*), we have identified different bacterial types, split into two dominant groups (Group 1 and Group 2), in aphid lines collected from Fife, Tayside and North Yorkshire. We have developed quantitative molecular methods to determine the relative abundance of secondary bacteria associated with the Cabbage aphid and shown that frequency of Group 1 and Group 2 secondary bacteria compared to *Buchnera* is variable between aphid lines. Experimental work indicates that Group 2 bacteria are associated with reduced aphid fecundity (Fig. 6a) and with increased egg production in emerging *Diaeretiella rapae*, the hymenopteran wasp species that parasitises cabbage aphids (Fig. 6b). Changes in aphid fitness

associated with the presence of particular secondary bacteria types are likely to affect the ability of aphids to act as vectors of plant pathogens.



**Figure 6** a) Impact of secondary bacteria on reproduction by the Cabbage aphid, *Brevicoryne brassicae*. b) The hymenopteran wasp *Diaeretiella rapae* together with Cabbage aphid nymphs. c) Large raspberry aphid, *Amphorophora idaei*. d) Impact of RLSV infection on large raspberry aphid pre-reproductive period.



Moreover, plant pathogens themselves can change plant chemistry in a way that makes crops more attractive to insect vectors. We have shown that the large raspberry aphid, *Amphorophora idaei* (Fig. 6c), preferentially colonises plants infected with one or more of the viruses it vectors, such as *Raspberry leaf spot virus* (RLSV). However, the plant virus impairs aphid performance by slowing its development so that the time taken by the aphid to start reproducing, the pre-reproductive period, is extended (Fig. 6d). This developmental delay may cause the aphid to disperse from the plant soon after initial colonisation, thereby facilitating the spread of the plant pathogen. Thus, characterising how microbes influence insect herbivores in arable and horticultural crops is providing us with an insight into how we might exploit ecosystem processes to ensure productive systems and maintenance of biodiversity.

### Understanding pathogen biodiversity as a key to controlling crop diseases

**Alison K. Lees, David E.L. Cooke, Brian Fenton & Adrian C. Newton**

Crop pathogens and pests are an unwelcome but important component of the biodiversity found in managed ecosystems. They are diverse and adaptable, meaning that they are able to respond quickly to

disease management practices and changing climate and therefore to reduce agricultural sustainability. Evolution of pests and pathogens, and the migration of new invasive species, leads to a constantly changing threat to crop production. In particular, the ability of pests and pathogens to overcome genetic resistance that has been bred, or engineered, into crops and the development of resistance to agrochemicals is problematic. Our work seeks to understand these threats, and the factors that drive population changes, with the goal of effective disease management. We illustrate this approach by discussing examples of our work on three very different host–parasite interactions. In each case we have developed and used DNA fingerprinting methods (microsatellite markers) to allow us to monitor and understand diversity.

*Phytophthora infestans* (Fig. 7) is the cause of potato late blight which results in serious reductions in yield and quality of ware and seed potato crops on a global scale. By monitoring several thousand isolates of *P. infestans*, sampled predominantly from commercial crops, we identified a dramatic shift in the UK population with an increase in the frequency of a particularly aggressive A2 mating type strain that is able to overcome some sources of host resistance. The implications of the changes have been reported to the potato industry and management practices altered accordingly.

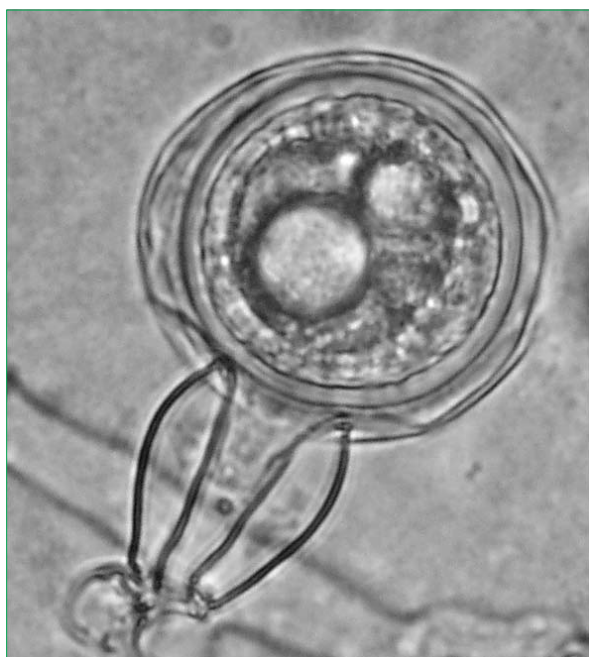


Figure 7 Image of an oospore of *Phytophthora infestans* – a long lived soil-borne survival structure which is a source of increased biodiversity in the population.

The fungal pathogen *Rhynchosporium secalis* causes ‘rhynchosporium’ or ‘scald’, the most problematic disease currently challenging sustainable barley production. Major genes for resistance to *R. secalis* in barley varieties are readily overcome, but partial resistance genes can provide effective resistance in the field. *R. secalis* is being monitored to understand how populations change in response to use of varieties with major resistance genes, partial resistance, and even non-symptomatic resistance and how barley variety mixtures affect population structure. Studies so far show that *R. secalis* is clearly a very variable pathogen with a very high capacity for adaptation.

In addition, natural variation in the *P. infestans* and *R. secalis* genes that govern the ability of the pathogens to infect and cause disease is being investigated so that the likely durability of corresponding resistance genes in the potato and barley gene-pools can be inferred, thus making efficient use of host biodiversity.

The peach–potato aphid is an important vector of potato viruses and can readily adapt to insecticides.

In a parallel situation to late blight we have followed the relative success of different genotypes each year and have shown that a single genotype, or clone, denoted type O, has come to dominate the UK population almost certainly due to the presence of a mutation causing resistance to dimethyl carbamate, a very commonly used insecticide. As the growth rate of clone O is no better than other genotypes (Fig. 8) it is also likely to have maintained good defences against attack by parasitoids in the field. Aphid genes are responsible for this phenotype as these *M. persicae* have no secondary symbionts. It appears that, like the potato plant itself its pests and pathogens effectively use asexual reproduction to multiply well adapted gene combinations.

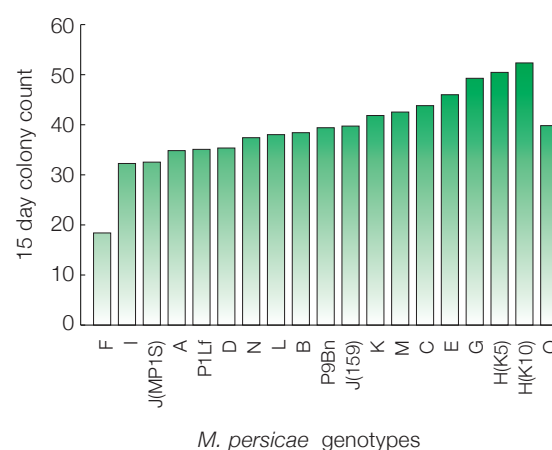


Figure 8 Variation in growth rate of different *M. persicae* genotypes (as characterised using SSR markers) measured as number of individual aphids per colony after culture on oilseed rape for 15 days.

## Perspectives

As illustrated above, it is clear that agriculture faces many challenges to sustain production as climatic zones shift and changes to agricultural systems accelerate. Consequently, new biotic and environmental stresses continually threaten crop production. A solid understanding, continued monitoring and appropriate use and/or management of biodiversity, from genes to agroecosystems, will remain a key component of agriculture's ability to adapt.



# Wealthier & Healthier

**Derek Stewart**

*A combined objective that is key to the development of a vibrant sustainable Scottish economy is the generation of food that creates wealth while being healthier and safer. The Scottish diet has long been held up as the worst in Europe, with Scots regularly topping the tables of degenerative disease such as cardiovascular disease, cancer etc. However, the national diet has improved with a commensurate reduction in disease levels, and through several national and international projects we aim to continue this trend. For example, as part of the EU-funded project, DEVELONUTRI, we are aiming to identify the points throughout the potato, tomato and wheat food chains at which nutritive value is lost.*

In addition, we are also trying to enhance the value of our staple foods such as bread by substituting a proportion of the wheat with barley. Barley is one of the few crops that has led to a product with an approved health claim due to the natural component  $\beta$ -glucan which – if present in the diet at ~4% – can lead to a reduction in the risk of cardiovascular disease. This project, BarleyBread, was funded by the EU and involved interaction with SMEs across Europe including the Scottish food ingredients company, Macphie of

Glenbervie Ltd, who generated trial breads which were hugely successful in consumer trials. Work on crop nutritional value and human health benefits is also being pursued in soft fruit such as blackcurrant, raspberry and other related crops.

The following articles show that our research into wealthier, healthier and safer food encompasses science from the fundamental through to the applied, and highlights our ability to translate this through to products.





## Understanding the genetics of raspberry fruit quality for improved consumer appeal

Julie Graham

Berry sales in the UK are now second only to those of apples, accounting for 19% of all fruit sold. Just under 33% of consumers now purchase raspberries (estimated retail value £125.7 million). Currently one week's raspberry sales are equivalent to what would have been purchased in a whole year six years ago. This success of the raspberry industry is due in part to commercial raspberry production having been revitalised by new horticultural strategies, specifically protected cultivation. Protected cultivation has been pivotal in the success of UK berry cultivation, and experiments at SCRI and in Blairgowrie have demonstrated significant increase in all quality parameters from tunnel grown raspberries. Predictions are for even greater increases given availability of appropriate high quality varieties that meet market requirements across a wider season.

Consumers are willing to pay a premium for quality raspberries and health would benefit from increased consumption. The most important factor for improvement in eating quality is flavour or specifically

taste, made up of sweetness and sourness and the balance of these two attributes and overall intensity of flavour with impacts from flavour volatiles. However, appearance is as important as taste for initial consumer selection, as only berries of high visual quality will be purchased. Berry size and colouration are key to consumer success here. Consumer disappointment, given a high price for raspberries, discourages repeat purchases.

Breeding new high quality raspberries which can be scheduled across a wider season is a long, difficult process hampered by several genetic problems. Concern over environmental impact and sustainability of agricultural and horticultural practices is leading to a greater emphasis on pest and disease resistance, as well as the ability of plants to withstand local environmental stresses. The changes in environmental, cultural and agronomic practices within the industry will impact strongly on the nature of the germplasm required for the future, with a greater interest in the conservation of genetic resources and utilisation of diverse locally adapted germplasm. Breeding methods used in raspberry have changed very little over the last 40 years and little novel germplasm has made its way into commercial cultivars. However, with the narrowing genetic base coupled with the increasing demands from consumers, new breeding methods are required to meet demands. The speed and precision of breeding can be improved by the understanding of the genetic control of key traits and then the deployment of molecular tools or markers (linked to good traits) for germplasm assessment and management. The correlation between traits and chromosomes is done by developing linkage maps which represent the plant chromosomes with locations of markers and traits placed along them. Implementation of marker assisted breeding which links easily scorable molecular markers to complex traits which require extensive field evaluations, can yield defined improvements in fruit quality across the cropping season in harmony with developments of disease resistances and production agronomic traits. In the raspberry this is now possible through the development of the SCRI raspberry genetic map that forms the basis for linking phenotype

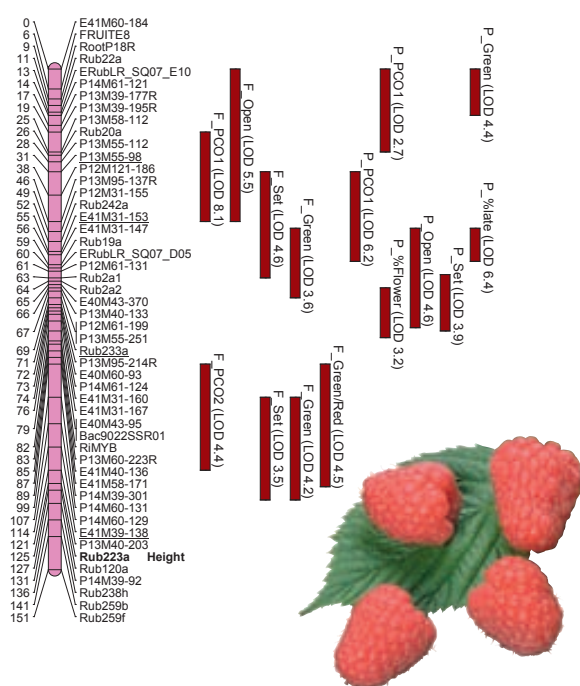


Figure 1 Chromosomal regions controlling fruit development and quality.



(plant traits) to genotype (plant genes). A number of chromosomal locations (quantitative trait loci [QTL]) for key quality traits in raspberry have been identified and in some cases the major genes responsible for trait variation have been identified (Fig. 1). These include steps towards understanding the ripening process which is essential to enable the development of high quality varieties fruiting across a wide season. Chromosomal locations responsible for ripening have been identified. A number of key genes including a MADS-box gene and Gene H were associated with the QTL and markers associated with plant height have also been identified. The major anthocyanin pigments in red raspberry have also been mapped to the same chromosome region on linkage groups (LG) 1 and 4 across years and from different environments. The most significant markers were genes including bHLH, and bZIP transcription factors which are thought to regulate the anthocyanin pathway. Colour has also been assessed with progress made in understanding the major genetic and environmental control. Sensory traits and the composition of fruit, both of which influence

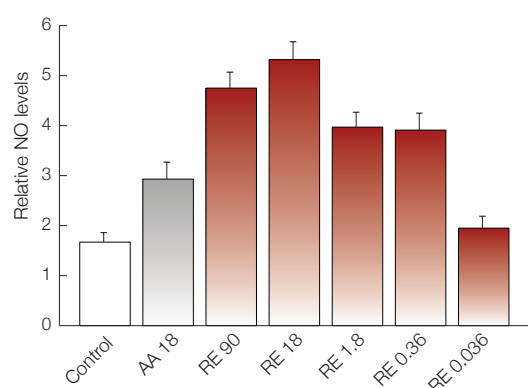
flavour, have been studied and again preliminary chromosomal regions for control identified. Methods have been developed that will allow us to measure and subsequently understand the genetic regulation of health components for improvements through marker assisted breeding.

Progress in the area of the genetic regulation of fruit quality is well underway, with marker assisted breeding now possible for a number of quality traits which can be transferred into conventional breeding programmes for targeted and timely improvements.

### Fruit components and their impact on fundamental disease mechanisms

#### Gordon J. McDougall

The health benefits associated with a diet rich in fruit and vegetables may be derived from the intake of natural phytochemicals. One theory suggests that polyphenol antioxidants (which are especially high in fruits and berries) protect against oxidative damage



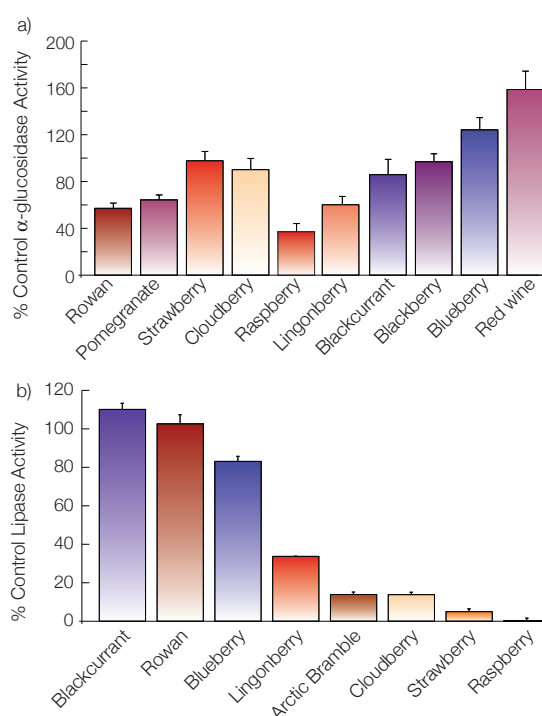
**Figure 2** Protection of nitric oxide levels in a rat model by berry extracts. AA = ascorbic acid, RE = raspberry extracts. All values are  $\mu\text{g/ml}$ .

involved in disease development. However, many polyphenols (e.g. anthocyanins) are poorly bioavailable and are unlikely to act as antioxidants at the cellular level. Indeed, large portions of berry polyphenols are not taken up into the blood and remain in the gut.

Nevertheless, evidence continues to accrue that berries or polyphenol-rich berry preparations can influence the progression of neurodegenerative diseases, cancers, cardiovascular disease and diabetes *in vivo* even if an overarching theory to explain their mechanism of action has not been formulated. We present evidence that berry polyphenols have bioactivities (often independent of their antioxidant capacity) relevant to cardiovascular health, obesity and glycaemic control.

Nitric oxide is a key signalling molecule which controls vasodilatory responses in blood vessels. Raspberry extracts (at 90 - 0.36  $\mu\text{g/ml}$ ) significantly increased nitric oxide levels in a rat model system and were more effective than ascorbic acid, a known cardio-protectant (Fig. 2). Protection of nitric oxide levels, presumably through an antioxidant mechanism by reducing free radical degradation, could influence cardiovascular fitness *in vivo*. Such model studies are backed by studies which highlight the cardiovascular benefits of berry intake.

Slowing the rate of digestion of starch rich foods could influence post meal blood glucose levels and glycaemic control, thereby benefiting type II diabetes. In model



**Figure 3** a) Inhibition of  $\alpha$ -glucosidase activity by berry extracts. b) Inhibition of lipase by berry extracts. All extracts assayed at 50  $\mu\text{g/ml}$ .

systems, polyphenol extracts from berries inhibit  $\alpha$ -glucosidase, the key enzyme in glucose release from starch, at doses easily achievable from normal dietary intake (Fig. 3). Inhibition of  $\alpha$ -glucosidase is the target for acarbose-type pharmaceuticals (e.g. Glucobay) which are currently prescribed to type II diabetics to reduce starch breakdown after meals. Identifying the active polyphenol ingredients is under further study.

Inhibition of fat digestion could reduce blood lipid levels and calorie intake from fat rich meals and benefit cardiovascular complications and directly influence obesity. In model systems, polyphenol extracts from berries inhibited pancreatic lipase, which is the key enzyme for fat digestion (Fig. 3). Inhibition of this enzyme by drugs, such as orlistat (Xenical also sold as Alli), is a current treatment for reducing obesity. Inhibition of lipase and reduction in blood lipid levels may explain the effects of berries on obesity in animals. The large differences in effectiveness of the berry extracts indicate that particular polyphenol components are more effective.





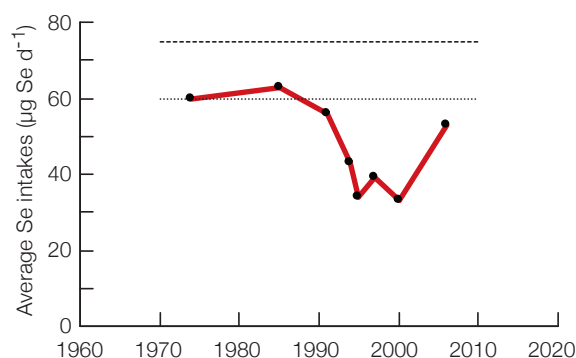
Derek Stewart and a tasting panel at work.

## Selenium biofortification of bread

**Philip J. White**

Selenium (Se) is an essential mineral element for human nutrition. It is incorporated into several important proteins as the amino acid selenocysteine. Among these proteins are iodothyronine deiodinase, which is responsible for converting the prohormone thyroxine to the active thyroid hormones; sperm-capsule selenoprotein, which is implicated in sperm motility; the antioxidant enzyme glutathione peroxidase; and selenoprotein P, the main Se-compound in plasma. According to the UK Food Standards Agency, “Selenium plays an important role in our immune system’s function, in thyroid hormone metabolism and in reproduction. It is also part of the body’s antioxidant defence system, preventing damage to cells and tissues”.

The UK reference nutrient intakes (RNI = the dietary intake that meets the needs of 97.5% of the population) for adult males and females are 60 and 75  $\mu\text{g Se d}^{-1}$ , respectively. However, the average dietary Se intakes in the UK have declined from over 60  $\mu\text{g Se d}^{-1}$  in 1985 to between 48 and 58  $\mu\text{g Se d}^{-1}$  today (Fig. 4). In Scotland, recent dietary surveys suggest that average dietary Se intakes may be lower than this. This downward trend is thought to be a consequence of replacing North American milling wheat, which is grown on high-Se soils and has a high Se concentration, with wheat grown on soils containing little Se in the UK. It has been suggested, therefore, that UK dietary Se intakes could be improved by increasing Se concentrations in homegrown milling wheat by applying Se-fertilisers to the crop. This practice is referred to as ‘agronomic biofortification’ and has been adopted



**Figure 4** Average selenium intakes ( $\mu\text{g Se d}^{-1}$ ) estimated from UK Total Diet Studies undertaken between 1974 and 2006. Horizontal lines indicate the UK reference nutrient intakes (RNI) for adult males ( $75 \mu\text{g Se d}^{-1}$ ) and females ( $60 \mu\text{g Se d}^{-1}$ ). Data sourced from the UK Food Standards Agency.

successfully in countries such as Finland. This year, the BAGELS consortium, which included Philip White from SCRI's Environment Plant Interactions Programme, have reported that the application of Se-fertilisers to UK wheat crops increases grain Se concentrations, and that Se concentrations in bread baked from this grain can be increased without affecting other quality attributes important for breadmaking.

BAGELS (Biofortification through Agronomy and Genotypes to Elevate Levels of Selenium; bagels.ukcrop.net) was a 'farm to fork' project sponsored by Defra through the Sustainable Arable LINK Programme. The partners spanned the entire food chain and included researchers, manufacturers and retailers. Partners included SCRI, University of Nottingham, University of East Anglia, Rothamsted Research, Institute of Food Research, Nickerson-Advanta, Velcourt, Carrs Fertiliser, Yara UK and Marks & Spencer. The main aim of the BAGELS project was to determine if the selenium levels of UK-grown wheat could be increased safely through the use of selenium-containing fertilisers, without causing harm to the environment. Field trials were conducted at the University of Nottingham, Rothamsted Research and Velcourt sites over two years on two widely-grown cultivars of Grade I bread-making wheat, Hereward and Solstice. These field trails demonstrated that the application of about  $10 \text{ g Se ha}^{-1}$  at the right time could increase grain selenium concentrations to levels similar to that of imported North

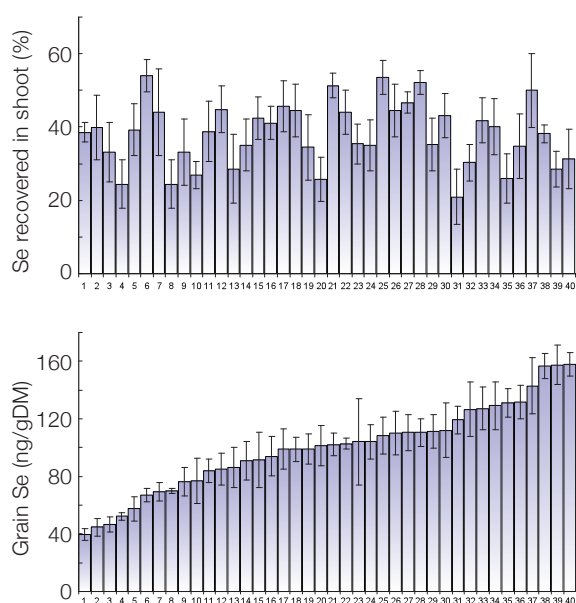


Harvesting a BAGELS field trial at the University of Nottingham. Photograph courtesy of Dr Martin Broadley.

American grain. The fate of the selenium applied in fertilisers was monitored in both the soil and the crop. Analysis of soils indicated that there was no residual build up of selenium in the soil that could pose an environmental risk. Analysis of the crop revealed that it acquired between 20 and 35% of the selenium applied in the fertiliser. About half of the selenium acquired by the crop remained in the straw, while half was recovered in the grain.



Glasshouse survey of genotypic variation in Se acquisition, Se redistribution within the plant, and grain Se concentration among wheat genotypes.



**Figure 5** Genotypic variation in Se accumulation in above ground tissues, expressed as a percentage of the Se-fertiliser applied, and grain Se concentration among 40 cultivars of winter wheat grown in the glasshouse at SCRI in 2007/2008. (1) Dover, (2) Batis, (3) Opus, (4) Ochre, (5) Brompton, (6) Riband, (7) Isengrain, (8) Monopol, (9) Petrus, (10) Hereward, (11) Alchemy, (12) Lynx, (13) Mascot, (14) Malacca, (15) Zebedee, (16) Einstein, (17) Solstice, (18) Gatsby, (19) Deben, (20) Gladiator, (21) Rialto, (22) Hyperion, (23) Caphorn, (24) Cordiale, (25) Avalon, (26) Enorm, (27) A50-03, (28) PBIS, (29) Claire, (30) Scorpion 25, (31) Soissons, (32) Sokrates, (33) Gulliver, (34) Maris Widgeon, (35) Arche, (36) Robigus, (37) Flanders, (38) Cappelle-Desprez, (39) Glasgow, (40) ELS.

At present, Se-fertiliser is a non-renewable resource, and it has been estimated that if all of the world's wheat was fertilised at  $10 \text{ g Se ha}^{-1}$  then commercially viable Se reserves could be exhausted in less than 80 years. We must, therefore, use Se-fertilisers wisely. The efficient use of soil-applied Se-fertiliser by the wheat crop is determined by two physiological processes: (i) the acquisition of Se by the root system and (ii) the translocation of Se to the shoot and its accumulation in grain. At SCRI, we have surveyed genotypic variation in Se acquisition, Se redistribution within the plant, and grain Se concentration among 10 spring wheat and 40 winter wheat cultivars grown in a controlled glasshouse environment. Individual plants were grown in tubular plumbing pipes filled with a gravel:grit:sand mixture (40:40:20 by volume) fertigated with a complete mineral nutrient solution. A single application of sodium selenate was made during early vegetative growth, delivering  $10 \text{ g Se ha}^{-1}$ . Significant differences were



The biofortification of bread with selenium.

observed between wheat cultivars in their acquisition of selenium, selenium distribution within the plant and grain Se concentration (Fig. 5). There was a strong negative relationship between grain Se concentrations and grain yield. Since previous studies have shown that Se accumulation in grain is generally linearly related to the rate of Se-fertiliser application, one strategy to achieve target Se-concentrations in wheat grain would be to match Se-fertiliser applications to expected grain yields.

To determine the fate of selenium during processing, the concentrations and chemical forms of selenium in grain, flour fractions and bread products were analysed at the Institute of Food Research and University of East Anglia. The dominant organic form of selenium in wheat grain, flour fractions and bread products was found to be selenomethionine, which is readily bioavailable to humans. It was also observed that over 90% of the selenium present in the grain was retained in wholemeal bread. The application of  $10 \text{ g Se ha}^{-1}$  to the wheat crop produced loaves of white and wholemeal bread containing  $6.4$  and  $7.1 \mu\text{g Se}$  per slice, respectively. Loaves baked from Se-biofortified UK wheat, in which one slice delivered approximately 10% of the RDI, were produced by Marks & Spencer for the Cereals 2008 Event.

In summary, the work of the BAGELS consortium has demonstrated that homegrown wheat can be biofortified with selenium through the agronomic application of Se-fertilisers, and that bread baked from Se-biofortified grain has the potential to increase dietary Se intakes in the UK.



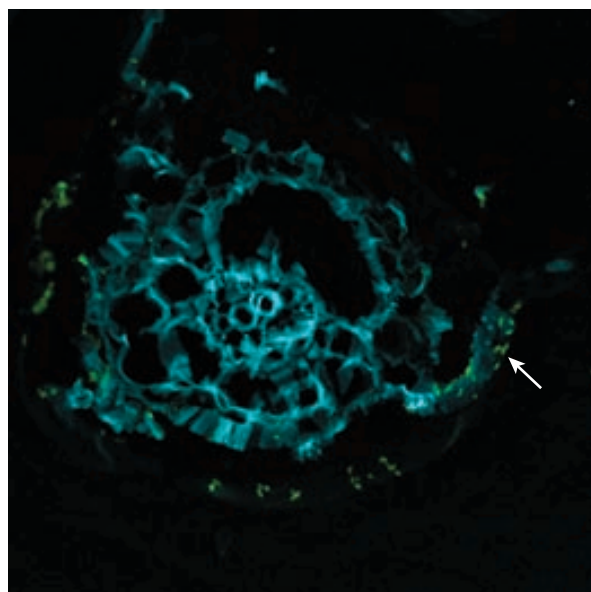


## Enterobacteria survival on plants: implications for food safety

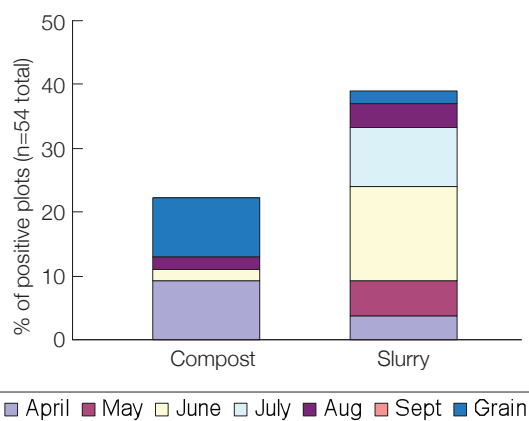
Nicola Holden

Food-borne bacteria that belong to the Enterobacteria family are able to persist in a wide range of environments and colonise hosts from every kingdom. Among these are the zoonotic pathogens that are passed from animals to humans, including toxigenic *Escherichia coli* and *Salmonella enterica*. Although traditionally associated with their animal hosts, an increasing number of food-borne outbreaks of these pathogens have occurred as a result of contaminated fresh produce, for example, ready-to-eat bagged salads. Contamination can occur at several stages during production and processing. However, we and others have shown that the bacteria actively interact with plants and can colonise them as alternative hosts. Our research at SCRI aims to understand the mechanisms behind bacterial colonisation of plants with the long term goal of reduction of food-borne bacteria from fresh produce.

At the present time, the number of outbreaks from fresh produce is relatively small. However, there has been a steady rise in outbreaks over the past two decades



**Figure 6** Microscopic image of a transverse section of a lettuce root that has been incubated with *Escherichia coli* O157:H7. The bacteria (in green) are located on the external tissue of the root (in blue) and can be clearly seen on a protruding root hair (arrowed).



**Figure 7** Presence of *Escherichia coli* isolated from the soil and barely grain of a barley trial site. The soil was amended with either municipal compost or animal slurry prior to sowing. Soil samples were collected on a monthly basis, barley grain post-harvest, and were analysed for the presence of *E. coli*-like bacteria. The number of plots, either treated with compost or slurry, that contained bacteria are expressed as a percentage of the total number of plots ( $n = 54$ ).

or so, while the incidence from farm animal products is declining. Several of the reasons for the increase have been recognised and appropriate measures adopted to counteract their effect. Nevertheless, many fundamental questions remain to be answered on the interactions between bacteria and host plants and on the mechanisms of transmission from animal to crop.

Research at SCRI covers both areas. Firstly, in the processes that underpin bacterial colonisation of plants: how the bacteria initially interact with plants (Fig. 6); how colonisation becomes established; how the plants respond to the bacteria. It appears that specific interactions occur between the bacterium and plant host during the initial stages of colonisation, in a similar fashion to colonisation of mammalian cells. Work is continuing to elucidate the nature of the interaction, which appears to involve multiple factors. Another aspect of colonisation is internalisation of the bacteria from the external tissue of the plant to the internal tissue. Again, many fundamental questions remain to be answered about this phenomenon, such as the extent of internalisation and the cues that drive it. This has important implications for food safety where current practices only decontaminate the external parts of fresh produce.



The second area is the transmission of zoonotic bacteria from animal hosts to crop plants. We have investigated the effect of adding farm waste to cereal plots and found that it is possible to detect the bacteria throughout the growing season and at harvest time (Fig. 7). This reinforces the finding that these bacteria are perfectly able to adapt to conditions quite different from their animal hosts and shows the potential for long-term persistence in the soil. Collaborative work between

Plant Pathology (Nicola Holden) and Environment Plant Interactions enables investigation of the bacterial populations on plants, which in turn, will aid in modelling the spread of the main food-borne bacterial pathogens in agriculture. This is particularly important in the context of climate change, which in addition to a direct influence on temperature and precipitation, has far reaching consequences for farming practices.

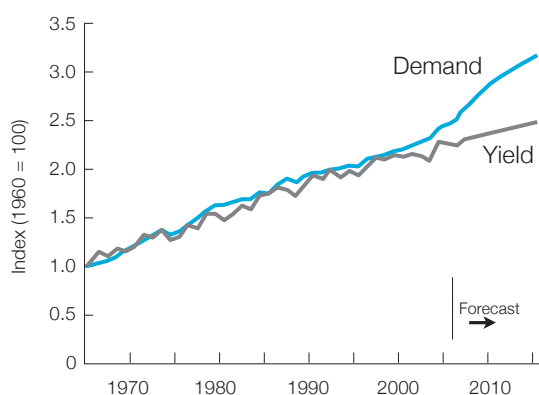


# Sustainability

**Philip J. White**

*The future brings unprecedented challenges to agriculture. The world must produce sufficient nutritious food for its burgeoning population, despite global climate change, the exhaustion of its natural resources, and environmental pressures to reduce pollution, greenhouse gas emissions and chemical inputs (Fig. 1). Great changes in agronomy are required to meet these challenges and scientific research is necessary to inform these changes.*

An increasing number of research projects at SCRI address agricultural sustainability, which is defined as the ability of a system to maintain stable levels of food production and quality in the long term, without



**Figure 1** Trends in global demand and yield for food and feed. Data are from "Food, Feed and Fuels: An outlook on the agriculture, livestock and biofuel markets" by Jeffrey Currie, Goldman Sachs International, March 2007.

compromising economic profitability or the environment. To achieve this, SCRI scientists are devising appropriate management strategies and developing novel crop varieties that will allow agriculture to preserve soil fertility, reduce chemical inputs, and optimise the use of water and fertilisers. The articles in this section illustrate aspects of SCRI's research towards agricultural sustainability.

It is estimated that over one third of potential crop yield is lost to pests and diseases. Historically, these losses were minimised using agrochemicals, but recent legislation has removed from use over half the active substances controlling agricultural pests and diseases. Alternatives to agrochemicals, such as the development of resistant varieties or management practices that reduce the incidence and severity of pest or pathogen attack, are urgently required. John Jones and colleagues describe how basic knowledge



of the molecular interactions between plants and their pathogens can be used to identify genes that can be introduced into crop varieties to confer disease resistance, while Tom Shepherd and Nick Birch describe the use of biochemical signals to lure pests into traps. This research complements other work at SCRI, such as the development of monitoring tools, predictive models and biological control measures, which underpin integrated strategies for management of pests and diseases in sustainable crop production.

Another major challenge for crop production will be restrictions on irrigation water. Ankush Prashar and colleagues describe how physiologists and geneticists at SCRI are identifying traits that improve water use efficiency by the potato crop. Genetic markers for these traits can then be identified and used in breeding improved varieties. This work is one of several projects investigating agronomic and genetic strategies for the efficient use of water and fertilisers by arable crops, whose objective is to maintain stable yields with reduced inputs in the face of global change.

SCRI is putting theory into practice in the Centre for Sustainable Cropping – Balruddery. Here, six fields will become a long term experiment to test whether our knowledge of agronomy, crop physiology, soil and environmental processes, and integrated pest and disease management strategies can be used to develop a sustainable agroecosystem that will optimise inputs, biodiversity and ecosystem processes in addition to crop yield. The experiment will comprise a six year rotation that will be maintained over at least four cycles. The design will incorporate crop traits and agronomic practices that enable the reduction of water and fertiliser inputs, soil and weed management strategies that allow high yields while maintaining soil fertility and arable biodiversity, and environmentally benign and cost effective integrated pest and disease management strategies. Further aspects of the rationale and design of this ambitious experiment are summarised by Cathy Hawes and David W. Hopkins.



## Functional analysis of pathogen effectors: how to go from genomics to durable resistance

Anna O. Avrova, Paul R. J. Birch\*, Ingo Hein, John T. Jones & Leighton Pritchard

Pests and diseases cause extensive damage to crops and represent a threat to sustainable production. In Scottish agrosystems pathogens such as late blight (*Phytophthora infestans* – Fig. 2), potato cyst nematode (*Globodera*) and *Pectobacterium* on potato and *Rhynchosporium* on barley cause yield losses and require application of a wide range of pesticides. Although these pathogens use different mechanisms to infect plants, all need to produce a number of protein molecules, known as effectors, that interact with the plant to manipulate host processes to the benefit of the pathogen. Effectors may adjust plant metabolism so that nutrients are transferred to the pathogen, or may suppress host defences. Recognition of these effector molecules by host resistance genes triggers a strong local defence response that leads to resistance. The identification of pathogen effectors is therefore of key importance for understanding how pathogens infect plants and for identifying resistance genes that can be introduced into crop varieties.

The increased accessibility of high throughput sequencing tools means that genome and transcriptome sequencing is now achievable for a wide range of plant pathogens. Researchers at SCRI are part of the consortia that have sequenced or are sequencing the genomes of *P. infestans*, *R. secalis* and *G. pallida*. We have also sequenced the genomes of a range of bacterial plant pathogens including *Pectobacterium atrosepticum* and several species in the



Figure 2 *P. infestans* infected potato leaf.

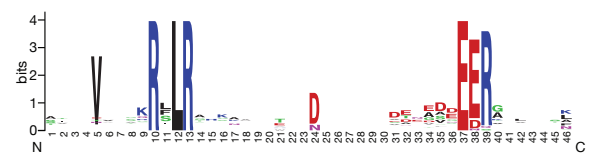


Figure 3 Sequence logo indicating the relative frequency with which particular amino acids occur in the RxLR translocation motifs of *P. infestans* effectors.

genus *Dickeya*. Some of these recently defined *Dickeya* species are emerging as severe potato pathogens in the UK, possibly as a consequence of climate change.

Bioinformatics can be used to aid identification of pathogen effectors. Effectors within a pathogen may have conserved sequence motifs that are important for their function. For example, we have found that *P. infestans* effectors contain a conserved RxLR motif (Fig. 3) that allows them to be translocated into the host cell, and this motif can be used to identify novel candidate effectors from a genome sequence. We can also link bioinformatics to experimental evidence, such as expression analysis, to identify genes that are up-regulated as the pathogen infects the plant, or that are expressed in pathogen tissues known to be a source of effector molecules, in order to predict which genes may be effectors.

These approaches have allowed identification of effector gene families from all of the key pathogens that we work with. Bacterial effectors that are secreted through the Type III Secretion System have been identified, and some have been shown to suppress host defences. Over 500 predicted proteins containing the RxLR motif which is required for some oomycete effectors to enter host cells have been identified in *P. infestans*, and a number of these have also been shown to suppress plant defences. Effectors identified from *G. pallida* include a very large family of proteins (SPRYSECs) that may suppress host defences and that have been shown to localise to a range of plant subcellular structures (Fig. 4). Finally, a project in which the transcriptome of germinated conidia of *R. secalis* was studied has led to the identification of a large number of secreted proteins, including candidate effectors. The roles of these effectors in suppressing host defences are now being investigated within the Plant Pathology programme.

\*University of Dundee

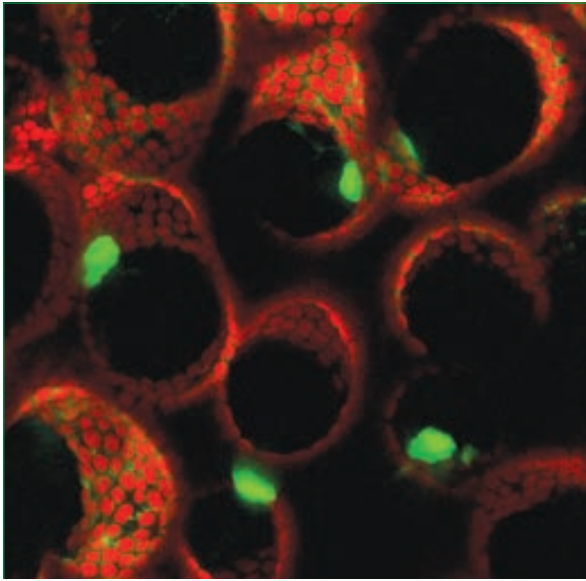


Figure 4 Subcellular localisation of a nematode effector to the nuclei of plant cells.

Although effectors play a role in suppressing the plant's innate immune system to allow infection, they may nevertheless be detected by surveillance systems in the plant. The constant evolution of pathogen effectors and host resistance proteins to, respectively, evade or gain molecular recognition, provides an intense



Figure 5 Recognition of AVR3a EM and KI in CPC accession *S. chacoense*.



arms race that may be exploited to seek durable forms of resistance for future potato breeding efforts. We have adopted a strategy to find effectors that are relatively conserved at the sequence level in pathogen populations; that are expressed in all pathogen isolates; and that provide an essential virulence function. Such effectors provide a point of vulnerability, an Achilles' heel, for the pathogen *if* resistance proteins can be found that detect and respond to them. To find such resistances, we are screening the Commonwealth Potato Collection for wild *Solanum* genotypes that respond to expression of these key effectors. A number of promising resistances have already been found in this way (Fig. 5), and their judicious combination in finished cultivars provides the potential for durable resistance to defeat our major crop diseases, and to reduce the levels of chemicals currently needed to control them.

### Fooling the bugs – using plant volatiles as chemo-attractants in integrated pest management schemes

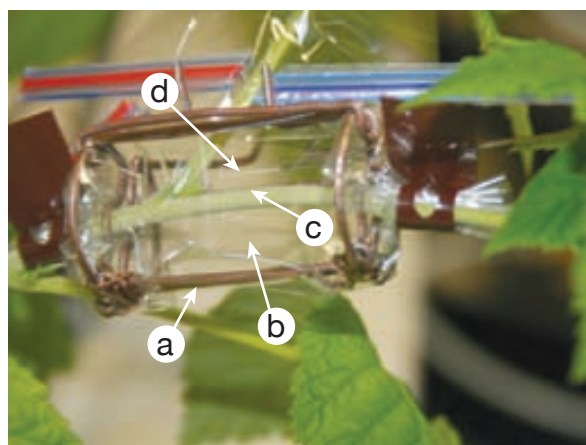
**Tom Shepherd & A. Nick E. Birch**

In northern and central Europe, raspberry production can be seriously affected by two major pests: raspberry beetle, *Byturus tomentosus* and raspberry cane midge, *Resseliella theobaldi*. The beetles lay eggs on opening buds and flower heads, and the resultant larvae burrow deeply into the berries to feed on the plug. This can result in damage to flowers and fruit with subsequent crop rejection. Splits in young canes caused by damage or natural splitting are the sites of egg laying by adult female raspberry midges. Emerging larvae feed on the pith, causing lesions which provide a means of entry for diseases such as cane blight fungus and midge blight. Currently there is an urgent need to reduce pesticide usage under revisions to EU pesticide policy 91/414/EEC, therefore development of environmentally friendly control measures forming part of an Integrated Pest Management (IPM) system are becoming vital. Two aspects of a successful IPM approach may include monitoring of insect numbers to improve the timing of insecticide treatments and use of control measures to



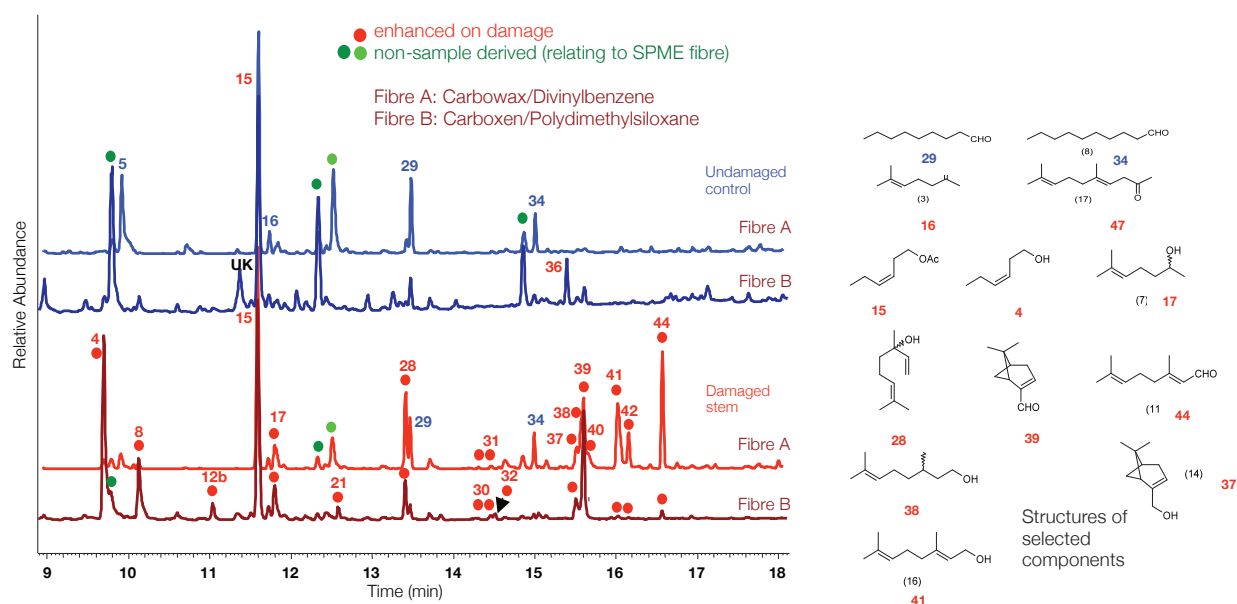
**Figure 6** Bucket trap for monitoring raspberry beetles. The trap mimics the exact colour and scent of a raspberry flower.

disrupt the normal life cycle. Methods for monitoring and biological control of pests involve development of monitoring and mass trapping systems using natural chemical attractants as lures ('biomimicry' of the host plant). In the case of raspberry beetle, analysis of floral volatiles at SCRI using gas chromatography-mass spectrometry (GC-MS) in conjunction with electrophysiology, resulted in identification of several beetle attractants which now serve as a lure used in a



**Figure 7** Enclosure for sampling volatile chemicals released from raspberry stems. Wire frames (a) supporting an inert plastic film (b) were positioned around the cane at sites having a 2–3 cm manually created split (c). SPME fibres (d) inserted into the enclosure adjacent to the split trap released volatiles which were then desorbed in the GC injector and analysed by GC-MS.





4: *cis*-3-hexen-1-ol; 5: camphene; 8: 2-heptanol; 12b: unknown terpene; 15: *cis*-3-hexen-1-ol acetate; 16: 6-methyl-5-hepten-2-one; 17: 6-methyl-5-hepten-2-ol; 21: 5-ethyl-2(5H)-furanone or 5-methyl-4-hexen-3-one; 28: linalool; 29: *n*-nonanal; 30: citronellal; 31: *trans*-pinocarveol or *trans*-verbenol; 32: unknown; 34: *n*-decanal; 36: methylsalicylate; 37: myrtenol; 38:  $\beta$ -citronellol; 39: myrtenal; 40: nerol; 41: geraniol; 42: neral; 44: geranial

**Figure 8** Volatile metabolites released from stems of raspberry cv Malling Promise analysed using SPME and GC-MS.

device (Fig. 6) marketed by AgriSense Ltd for precision monitoring of this key pest, so 'hot spots' can be treated as required.

For monitoring and control of raspberry midge two approaches are being taken. Traps developed and tested by colleagues at the Natural Resources Institute (NRI) and East Malling Research (EMR) use the female sex pheromone for local monitoring of males and, potentially, also for mating disruption. However, effective control requires identification of an attractant to lure emerging females in early spring (first generation) and during the second generation, coincident with fruit harvest and the main period of fungal colonisation. We have developed a novel sampling technique using solid phase microextraction (SPME) fibres to trap cane volatiles close to a split (Fig. 7).

A suite of volatiles has been identified, using several different raspberry varieties, which show a consistent pattern of enhancement following damage (Fig. 8). These compounds consist mainly of a family of structurally related terpenes, many of which are known to have behavioural effects on insects and plants (for example they attract natural enemies) and/or are produced in response to insect herbivory.

The physiological activity of the entrained volatiles is being investigated with collaborators at NRI by means of a GC-electroantennography (EAG) system which identifies the metabolites that elicit a response from the antenna of female midges. Compounds showing enhanced production when canes are damaged, and those shown to elicit a response from the midge are being used to test and develop lures for biological control and monitoring using laboratory and field based behavioural bioassays developed by SCRI, EMR and NRI. This work is funded by a five-year HortLINK project (SF 74): 'Integrated pest and disease management for high quality raspberry production'.

## Water use efficiency in potato

**Ankush Prashar, Timothy S. George, Gavin Ramsay, Paul D. Hallett, Hamlyn G. Jones\*, Peter Hedley, Jim W. McNicol, M. Finlay B. Dale, Philip J. White & Glenn J. Bryan**

Water is one of the key resources challenging the sustainability of modern agriculture. In developing countries, potato production is increasing because of its ability to provide nutritious food in a short season. However, the potato crop requires profuse irrigation. In the UK, potato production currently uses about half of

\*University of Dundee





Figure 9 Phenotypic screening of extreme  $\delta^{13}\text{C}$  genotypes under controlled conditions.

all irrigation water. Further climate change is inevitable and, as this proceeds, summers are predicted to become drier and water for irrigation scarcer.

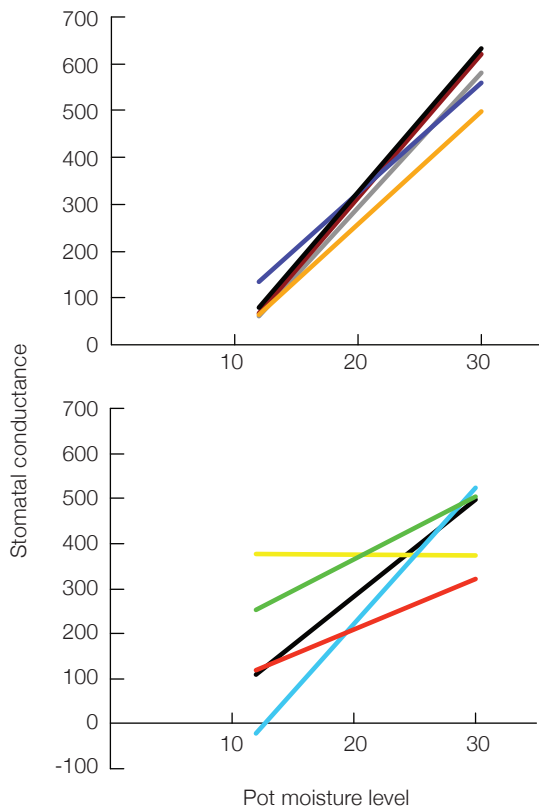
Yields of commercial potato varieties are often restricted by water availability. Their root systems are generally sparse and shallow, and they close their stomata, preventing photosynthetic carbon assimilation, while water is still available in the soil. To enable breeding of drought tolerant varieties, we are developing phenotypic screens that will allow us to explore the genetic basis of key traits for water use efficiency (WUE).

From field trials of a genetic mapping population performed in collaboration with Dr Andrew Thompson of Warwick-HRI, we initially identified ten genotypes with contrasting transpiration efficiencies based on leaf  $\delta^{13}\text{C}$  values. These genotypes were then cultivated under controlled glasshouse conditions in a soil typical of arable fields in Scotland (Fig. 9). After emergence, plants were grown for 30 days in soils watered to field capacity (30% volumetric content, -5 kPa water

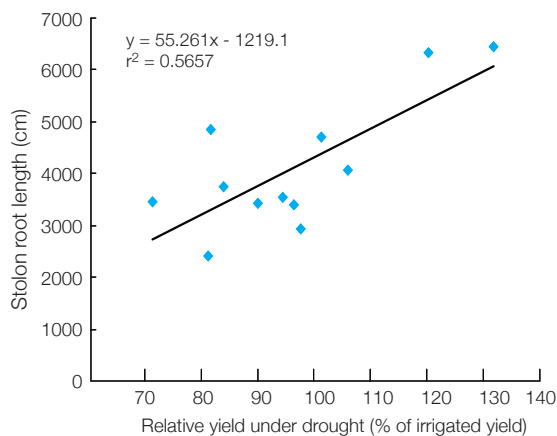
potential) before being divided into three groups irrigated to 30%, 20% (-300kPa, slight stress) and 12% (-1500kPa, wilting point) volumetric content. As WUE is a complex phenomenon, we evaluated a number of associated physiological and morphological traits. Tissue samples were also collected at different time points to determine differentially expressed genes at these moisture levels.

Response to water stress not only includes closing stomata but also reducing the density of stomata during leaf development. Preliminary data from our experiments shows that transpiration-efficient genotypes, as indicated by low leaf  $\delta^{13}\text{C}$  values, have consistently lower stomatal conductance at 12% volumetric soil moisture than transpiration-inefficient genotypes (Fig. 10). Thus transpiration-inefficient genotypes transpire more water at lower soil water content.

Another approach we are taking to improve the drought tolerance of potatoes is to screen genotypes for root length, specifically that associated with stolons. A



**Figure 10** Regression of stomatal conductance against pot moisture levels for the five selected clones with low and high  $\delta^{13}\text{C}$  values from a prior field experiment.



**Figure 11** Relationship between stolon root length and relative yield under drought conditions in the field experiment.

number of field trials have demonstrated that there is significant variation (~5-fold) in stolon root length between genotypes of potatoes. During last year's field season we demonstrated that stolon root length



**Figure 12** a) Potatoes grown under polytunnels to impose droughted treatments in 2009. b) Measurement of soil water content down the profile to establish the impact of the drought and irrigation treatment of soil water availability.

was strongly correlated with the ability of the potato plants to yield under droughted conditions (Fig. 11). This was achieved by growing potato cultivars with extreme rooting phenotypes under polytunnels with and without irrigation which developed strongly contrasting water availabilities in the field (Fig. 12). It is therefore apparent that as well as transpiration efficiency, rooting characteristics are likely to be critical for drought tolerance in potatoes.

These phenotypic screens in combination with microarray analysis will help us understand the basis of WUE and enable us to correlate WUE with gene expression in appropriate tissues. Identification and mapping of candidate genes that underlie WUE will ultimately enhance breeding for water efficient varieties in multiple environments.





SCRI's Balruddery Farm

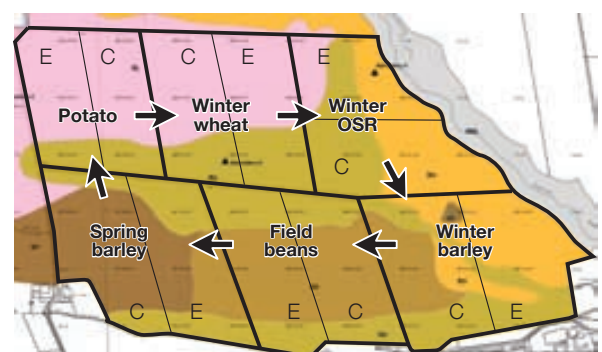
## A new platform for sustainability research at SCRI

**Cathy Hawes & David W. Hopkins**

The long-term viability of farming in Scotland depends on the sustainable management of our agricultural habitats. Intensification of arable systems to maximise crop yields in the short term has raised serious concerns about the functioning of arable systems and food security in the long term. The challenge now is to identify appropriate sustainable management practices and associated crop varieties that will allow farmers to achieve a balance between the potentially conflicting goals of maximising crop production, conserving arable biodiversity and maintaining ecosystem functions.

To do this, SCRI is establishing a new experimental research platform at Balruddery Farm near Dundee, for long-term studies on arable sustainability. The Centre

for Sustainable Cropping – Balruddery is the first of its scale in the UK and will provide a test-bed for new management practices and ‘sustainable’ crop varieties developed at SCRI. The six fields that make up SCRI's Centre for Sustainable Cropping cover approximately



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**Figure 13** Layout of fields and treatment (conventional [C] and experimental [E]) management methods within the sustainability platform at Balruddery Farm.



40 ha in the south-east corner of Balruddery Farm. Each field will be divided into two halves according to topography, boundary vegetation and soil type. The conventional and experimental management treatments will be randomly allocated to each half field. Four to five different varieties of the six crops in the rotation will then be sown across both treatments, providing a comparison of varietal differences in performance (Fig. 13). Background on the farm and the design of the platform was given in the SCRI Annual Report for 2008 and further details can be found at [www.scri.ac.uk/sustainability](http://www.scri.ac.uk/sustainability)

The experiment will start with the sowing of winter wheat, barley and oilseed rape varieties in autumn 2010 and beans, barley and potatoes in spring 2011. Maize was sown in April 2009 to provide a  $C_4$  input to the existing  $C_3$  based system that will be used as a signal to trace carbon fluxes and turnover under the two contrasting management systems. The maize crop grew well during 2009 and was cut at cob formation in September when the crop was between 1.2 and 1.8m high (Fig. 14). The entire crop residue was chopped and incorporated in the autumn. A second crop will be grown and incorporated in the same way in 2010 to boost  $C_4$  levels in the soil and extend the period over which the pulse can be detected during the first rotation. This signal will provide a tool to characterise the flux of carbon from plants to soils, the processing of these inputs through components of soil food webs and their return to the atmosphere through biological activity.



Figure 14 Maize at Balruddery grown to provide a  $C_4$  signature for measuring carbon dynamics.

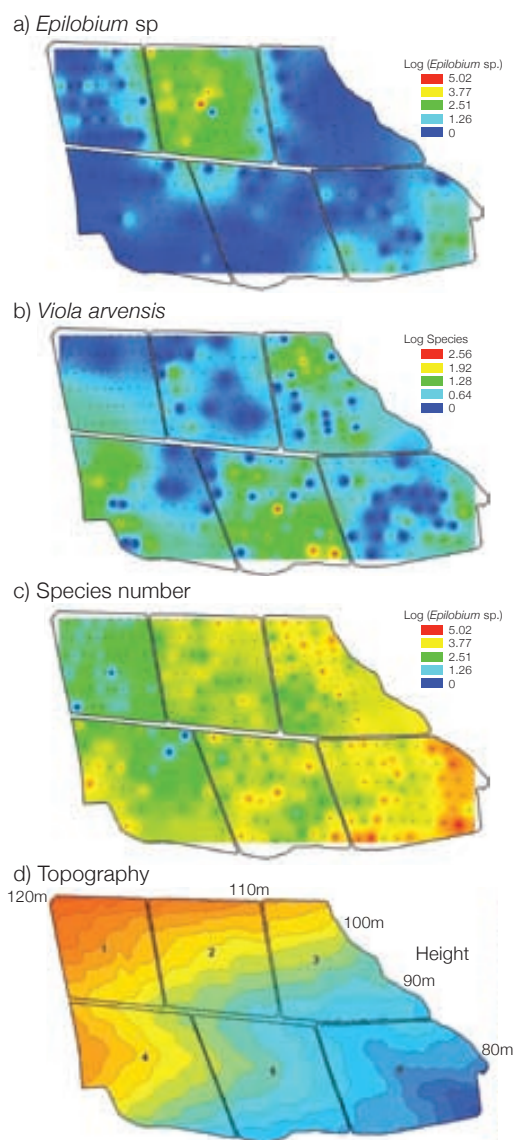


Figure 15 The distribution, abundance and diversity of arable weeds in the seedbank as assessed in spring 2009.

Baseline data is being collected on a wide range of system properties including arable plant and invertebrate biodiversity, soil microbial diversity and function, soil chemistry and biophysics, hydrology, nutrient and carbon fluxes and greenhouse gas emissions. The distribution, abundance and diversity of arable weeds in the seedbank was assessed in spring 2009 by identification of emerged weed seeds germinating from soil samples collected from a grid of 60 sample points across each field. The data shown include the weeds from the first flush of emergence only (Fig. 15). This is a good representation of the



arable seedbank diversity, but does not provide an estimate of absolute abundance. The full dataset will be reported in 2010. However, these data indicate a high degree of spatial variation across the site. *Epilobium* sp. (willowherb) is concentrated in one field (Fig. 15a), whereas *Viola arvensis* (field pansy) is more widely distributed (Fig. 15b). Species richness (Fig. 15c) is lowest in the north-west corner and increases towards the south-east, following the NW-SE slope across the Centre (Fig. 15d). Explanations for these patterns from field histories and erosion patterns will be sought and trends away from this baseline under conventional and experimental management will be measured annually. The data collected during 2009 and 2010 will provide the baseline against which to assess future changes

in soil health, biodiversity and system functions under experimental and conventional field management.

A major obstacle to the adoption of more sustainable farming practices in Scotland is the short-term economic situation of farmers which provides a strong incentive for intensive crop production methods. Maximising yield is therefore often prioritised over management for environmental sustainability, particularly as the latter is generally ill-defined. The aim of the Centre is to quantify the longer-term costs and benefits of new 'sustainable' crop varieties and management practices at a scale that is relevant to commercial farmers. This will provide the data necessary to identify a balance between the risk of potential short-term loss for long-term environmental and economic gain.



# Communications – Review of the Year

## **Phil Taylor**

SCRI enjoyed an astonishingly busy year from the perspective of the events and public engagements in which the staff participated.

It was also a watershed year in the Institute's history following the decisions of both the SCRI and Macaulay Land Use Research Institute's Governing Boards to accept in principle the formation of a new, research organisation.

The 'New Institute', as it has been tantalisingly christened, will combine the respective strengths of both

institutes while at the same time exploiting new scientific opportunities and the advantages of scale.

The Scottish Government's Cabinet Secretary for Rural Affairs and Environment, Richard Lochhead MSP, summed up the aspirations of all concerned when he said: "Both SCRI and the Macaulay already enjoy well-deserved reputations for excellence. Together they will be in an even stronger position to compete in the international arena and to address complex global issues, further raising Scotland's profile on the world stage."

It may be the case that SCRI's 2009 Annual Report is the penultimate edition. Happily there is no shortage of events, visits and achievements to fill the pages.

2009 was the first year in which several of SCRI's major stakeholder knowledge exchange events were held at Balruddery Farm. Balruddery was purchased by SCRI in 2008 and comprises 118 hectares of arable farm land located seven miles west of Dundee and between 70 and 160 m above sea level on the lower slopes of the Sidlaw Hills.



Directors and senior scientists from SCRI and the Macaulay Land Use Research Institute at their first joint meeting in Montrose.



Cereals in Practice which saw an encouraging growth in attendance in its first year at Balruddery.

Both Cereals in Practice and Potatoes in Practice were transferred to Balruddery and both events were hailed as successes.

Cereals in Practice (CiP) is organised jointly by SCRI, SAC (Scottish Agricultural College) and SSCR (Scottish Society for Crop Research). It is also supported by HGCA (Home-Grown Cereals Authority). It is the key event in the cereals industry calendar in Scotland.

More than 200 visitors arrived on the day, more than twice the number of visitors than in previous years and including a large group from Sweden.

Dr Keith Dawson, SAC's Principal Crops Consultant, who is also chairman of the SSCR Combinable and Energy Crops Sub-committee, said: "There was a tremendous amount of work done by all concerned to bring together the programme for the event.

"The whole aim of the work we do is to try to speed up the development of knowledge from the research lab through to the field and through on to the farm to benefit the consumer and the industry as a whole."

In August, Potatoes in Practice (PiP), the UK's largest potato industry field event, also made the move to Balruddery Farm. For the second year running, very heavy rain immediately before the event caused some difficulties. Happily a bright and windy day soon helped dry the ground. More than 600 visitors made the journey to this lovely corner of Angus. The biggest proportion was farmers, closely followed by those active in marketing and retail.

PiP was sent a message of support from Roseanna Cunningham MSP, Scotland's Environment Minister. She said: "Potatoes in Practice is a great example of Government and industry working in partnership to create a real Scottish success story. Participation in events such as this helps to ensure that Scotland will retain – and expand upon – its leading position." Later in the year SCRI and Mylnfield Research Services Ltd (MRS Ltd) joined forces at British Potato, the Potato Council's biennial show at Harrogate in Yorkshire.

Fruit for the Future, SCRI's knowledge exchange event tailored for the soft fruit sector, also saw some changes in 2009. It too has seen a steady growth in audience numbers and it was necessary to relocate from the



Commonwealth Potato Collection Assistant Curator, Gaynor McKenzie, with a special CPC display at Potatoes in Practice.





Guests at the 2009 Fruit for the Future event – the biggest to date.

Institute's biggest lecture theatre to a larger marquee set up on the lawns in front of the headquarters.



Guests on a tour of the polytunnels at the 2009 SCRI Director's Summer Soirée.

The installation of the marquee gave us the opportunity to host a summer evening soirée on the same evening on behalf of SCRI's Director, attended by more than 100 friends and stakeholders of SCRI.

Each year the Scottish Research Institutes host a distinguished lecture under the title Science for Life. This year it was the turn of SCRI to host the event and the invitation to speak was kindly accepted by Professor Emeritus Thomas Rosswall.

Swedish born Professor Rosswall, recently retired Executive Director of the International Council for Science (ICSU), speaks on behalf of the Challenge Programme of the Consultative Group on International Agricultural Research. This is a consortium of more than 2000 scientists in 100 countries with over \$500 million invested each year for research for development.

He told his audience at Dundee's science centre, Sensation, that the issues of food security and climate change had to be much more closely linked.



The central message of his lecture was that the global demand for food is expected to increase by 50% by 2030 as the population grows and consumption patterns change. He said there was an urgent need to connect climate change and food security both in scientific and policy contexts.

The Science for Life lecture is also the opportunity to run a postgraduate student competition testing oral and presentation skills. Happily for SCRI, the 2009 winner was our own Lindsay McMenemy, a member of the Environment Plant Interactions programme. Lindsay's presentation topic was: 'Plant mediated interactions with the large raspberry aphid.' Professor Rosswall made the prize presentation of a certificate and cheque.



Lindsay McMenemy, winner of the 2009 Scottish Research Institutes' Student PhD competition (middle) with her fellow contestants, Peter Gregory and guest speaker Professor Thomas Rosswall.

The runners up were: Paula Scott from the Rowett Institute of Nutrition and Health and Kerry Waylen from the Macaulay Land Use Research Institute.

Another award went to Susan McCallum of SCRI who won the Innovation prize, which is supported by the Mylnefield Trust and MRS Ltd. Susan's title was: 'Marker assisted breeding for sensory characteristics in red raspberry (*Rubus ideaus* L.)'.

For Lindsay McMenemy it was the second achievement of the year: she was awarded the runner-up prize in a student essay competition run by the Royal Entomological Society.

Another notable achievement was the award to SCRI's Dr Julie Graham of the Royal Horticultural Society's Jones-Bateman Cup. The trophy is awarded by the RHS every three years to recognise major advances and excellence in fruit research. It is the second time in recent years that SCRI has been recognised. A previous winner was Dr Rex Brennan for his work on blackcurrants.

Dr Graham's research focuses on identifying genetic markers for desirable traits in raspberries such as taste and pest and disease resistance. Her research has led to new more specific and timely breeding methods which are based on an understanding of the link between genotype, the genetics of the plant, and the phenotype, what the plant looks or tastes like.



Dr Julie Graham, winner of the RHS Jones-Bateman Trophy.

Four SCRI scientists won an outstanding paper award from the American Journal of Potato Research. (Morris, W.L., Ducreux, L.J.M., Bryan, G.J. and Taylor, M.A. 2008. Molecular dissection of sensory traits in the potato tuber. *American Journal of Potato Research* **85**, 286-297.) Mark Taylor, Laurence Ducreux and Wayne Morris are members of the Plant Products and Food Quality



programme and their colleague, Glenn Bryan, works with Genetics. Their former colleague Walter de Jong collected the award on the authors' behalf at the Potato Association of America annual meeting in August in New Brunswick, Canada.



An 'outstanding paper' award from the American Journal of Potato Research was received by (left to right) Laurence Ducreux, Wayne Morris, Glen Bryan and Mark Taylor.

Later in the year SCRI and the University of Dundee's College of Life Sciences hosted the annual Distinguished Lecturer Seminar. It was delivered by Professor Nick Harberd who was at the John Innes Centre for many years before moving to the University of Oxford to take up the Sibthorpe Professorship in Plant Sciences.

The busy season of SCRI events continued with a new, joint venture involving the Royal Botanic Garden Edinburgh. The 'Berry Festival' was held over a weekend in July and visitors to the Garden's exhibition centre were treated to the story of Scotland's rich and diverse berries.

Photographs, living plants and other exhibits celebrated the bewildering variety of raspberries, strawberries, blackcurrants, brambles, tayberries and other fruits and demonstrated the science behind Scotland's berry industry.

SCRI was also the driving force behind the biggest event in the 2009 calendar of activities organised by KnowledgeScotland. This is a knowledge exchange initiative funded by the Scottish Government and run jointly by all the Scottish Research Institutes.

'Science Policy Success' was a two day conference held at Dynamic Earth in the early autumn. One hundred and thirteen delegates watched presentations on the themes of health and wellbeing, climate change, biodiversity and sustainability. There was also an acclaimed presentation by the writer and broadcaster Lesley Riddoch who discussed the task of communicating complex information to the public.



Scottish Government Chief Scientist, Dr Anne Glover, addressing the Science Policy Success conference at Dynamic Earth in Edinburgh.

The post-conference evaluation found that 71% of those responding believed that the conference had achieved its aim of improving links between science and policy.

In September, scientists from all over the world travelled to Dundee for the meeting of the 'Biometrics in Plant Breeding' section of EUCARPIA, hosted by SCRI.

More than 100 delegates took part and also enjoyed a



Professor Mike Kearsey of the University of Birmingham speaking at the EUCARPIA conference dinner where he was honoured for his long and distinguished career.



Delegates at the EUCARPIA conference at their civic reception in Dundee.

civic reception hosted by the Lord Provost of Dundee, Councillor John Letford. Later they had a memorable dinner on board HM Frigate Unicorn, one of the last intact warships from the days of sail, and one of the six oldest ships in the world.

Towards the end of the year, a stakeholder day was organised on behalf of the Scottish Government's research initiative Programme 1: Profitable and Sustainable Agriculture – Plants. The day included short presentations from scientists contributing to RERAD's commissioned research and from relevant policy divisions.

Three other set-piece events helped to put SCRI in touch with its audiences.

The 2009 Royal Highland Show set a new attendance record over its four days with 176,522 people coming through the gates at the Royal Highland Centre, Ingliston, Edinburgh.

At the SAC Pavilion SCRI presented the 'timeline' of the Institute's history and also an interactive display

of the Living Field run by staff from Environment Plant Interactions. In the Royal Highland Education Trust



Her Majesty The Queen at the Royal Highland Show, Ingliston. The Queen spent nearly 10 minutes chatting to staff and Dundee school visitors at the SCRI stand in the Royal Highland Education Trust building.



building, a team from Communications introduced a younger audience to the work being done on berries, nutrition and health. The highlight for the team was a visit from Her Majesty the Queen who spent nearly 10 minutes chatting to staff and a party of visiting schoolchildren from Dundee.

For the second year running, SCRI joined the staff of Edward Baxter's Gilston Mains Farm in Fife for LEAF (Linking Environment and Farming) Open Farm Sunday. Once again the Living Field team was in action.



Celebrity guest Stephen Purdon from the BBC's River City soap with visitors to LEAF Open Farm Sunday.

The Institute's work on soft fruit was the centrepiece of the display mounted for the Dundee Flower and Food Festival in early September.

SCRI hosted a number of special guests during 2009 and they included members of parliaments far and



SAC Chairman, Lord Jamie Lindsay with the SCRI Director during a visit in July.



Members of the European Parliament on a visit to SCRI.

wide. Pete Wishart MP (Perth and North Perthshire) is a long time supporter of SCRI and was welcomed back to Invergowrie in September. Another very supportive politician, Alyn Smith MEP, was also welcomed back to SCRI during the year. In August the Director welcomed a visitor from further afield: Tony Windsor MP – a member of the Australian House of Representatives for New England.

On the day of Fruit for the Future SCRI hosted Lord Jamie Lindsay, a former Scottish Agriculture Minister and currently the Chairman of SAC. He met the Director and then was hosted by all the science programmes before being taken on a quick tour of the glasshouses, the Commonwealth Potato Collection and the Living Field.

SCRI's strong links with the Scottish Government were reflected in visits from Ron Stagg, Deputy Director, Research and Science Division, Scottish Government – Rural and Environment Research and Analysis Directorate and from Dennis Dick, a former Chairman of the Scottish Wildlife Trust and a member of the Scottish Biodiversity Committee which is advising the Government on the 2010 UN Year of Biodiversity.

One of the most memorable seminars to be held at SCRI during the year featured the renowned Professor Dianne Edwards, Head of the School of Earth, Ocean and Planetary Sciences at Cardiff University. Professor Edwards is also a fellow of the Royal Society and a trustee of the Natural History Museum and the National Botanical Garden of Wales.

Her lecture was one of the many events to celebrate the achievements of Women in Science, Technology, Engineering and Mathematics (STEM) that took place at



Eleanor Gilroy demonstrates 'You choose the news' - an interactive game on the theme of GM.

the Universities of Dundee, Abertay and St Andrews and at SCRI and the Sensation science centre.

SCRI's strong links with organisations furth of Scotland were underlined by involvement in a major Biotechnology and Biological Sciences Research Council (BBSRC) project and an award from the Monsanto Trust for a project in Africa.

SCRI and the University of Dundee are collaborating in a £27 million Sustainable Bioenergy Centre under the auspices of the BBSRC.

The centre was launched to provide the science to underpin and develop the important and emerging UK sustainable bioenergy sector. It represents the biggest ever single UK public investment in the field.

The University of Dundee, based at SCRI, was named as one of the six research hubs of academic and industrial partners to benefit from the investment. The Dundee-led team (which includes SCRI) aims to alter lignin production in barley to make it easier to produce bioenergy from waste straw without reducing the quality of the crop.

In another project SCRI is leading an international project to help potato farmers in Kenya. The mission is being supported with a grant from the Monsanto Fund totalling £186,000.

The ambition is to increase potato yields – and importantly farmers' incomes – by establishing systems



Dr Lesley Torrance of Plant Pathology with the Monsanto Fund's Bridget Badiou at the launch of an international project to help potato farmers in Kenya.

to support virus-free potato seed tuber production. The potato is the second most important food crop in Kenya after maize; 1.2 million tonnes are grown every year.

SCRI's Head of Plant Pathology, Dr Lesley Torrance is leading the project. She said: "There is massive scope for crop improvement by effective control of virus diseases and their aphid carriers. We hope to increase potato yields and thereby farmers' incomes on a sustainable basis."



Farewell to John: Dr John Bradshaw is presented with a leaving gift on his retiral after a distinguished career at SCRI.

During 2009 the Institute bade farewell to several long-serving and distinguished members of staff. Dr John Bradshaw, one of the world's leading plant breeders, retired at the end of March. John began his career in Scotland at the Scottish Plant Breeding Station in Pentlandsfield, Edinburgh 33 years ago. Latterly at SCRI he led the Potato Genetics research programme.

SCRI Director, Professor Peter Gregory, said John embodied what was best about SCRI. "John is an academic to his fingertips and has a love of his subject. He has the ability to communicate the passion that he has for his subject. But he's not just an academic who operates on his own. He interacts with many other people within the Institute. He conveys that passion to visitors and in scientific meetings and in particular he has converted that academic passion into commercially useful products that people want to buy. It is a marvellous testimony to his whole career that he has been able to do these things bringing great credit to both him and to the institute."

SCRI also said farewell to Mike De,Maine, who retired from SCRI at the end of April after long and distinguished service, latterly as the Health and Safety Manager.



Mike De,Maine's retirement.



Ronnie Forbes RSA takes a breather during the launch party for his SCRI portfolio.

Two members of staff celebrated 40 years service with SCRI: Alex Mills, a soft fruit specialist with Field Services, has planted and tended tens of thousands of raspberry canes and blackcurrant bushes. Ronnie Ogg, another member of the Invergowrie community, has also spent 40 years working in the glasshouses and has been a stalwart helper at all the Institute's public events.

Much of SCRI's work in genetics, plant pathology, environment plant interactions and food has now been summed up in stunning imagery by the Leverhulme Trust artist-in-residence, Ronnie Forbes RSA. Ronnie's time with SCRI came to an end, but his portfolio of work went on public show during the year, starting at the Hannah Maclure Centre in Dundee. The body of work produced during his residency – *De Rerum Natura: The Nature of Things* – forms a narrative around the way we see and understand the world. The work comprises large paintings, unique digital-collage prints and a film. More exhibitions of his SCRI work are planned in the year ahead.







# Biomathematics and Statistics Scotland

**David A. Elston**

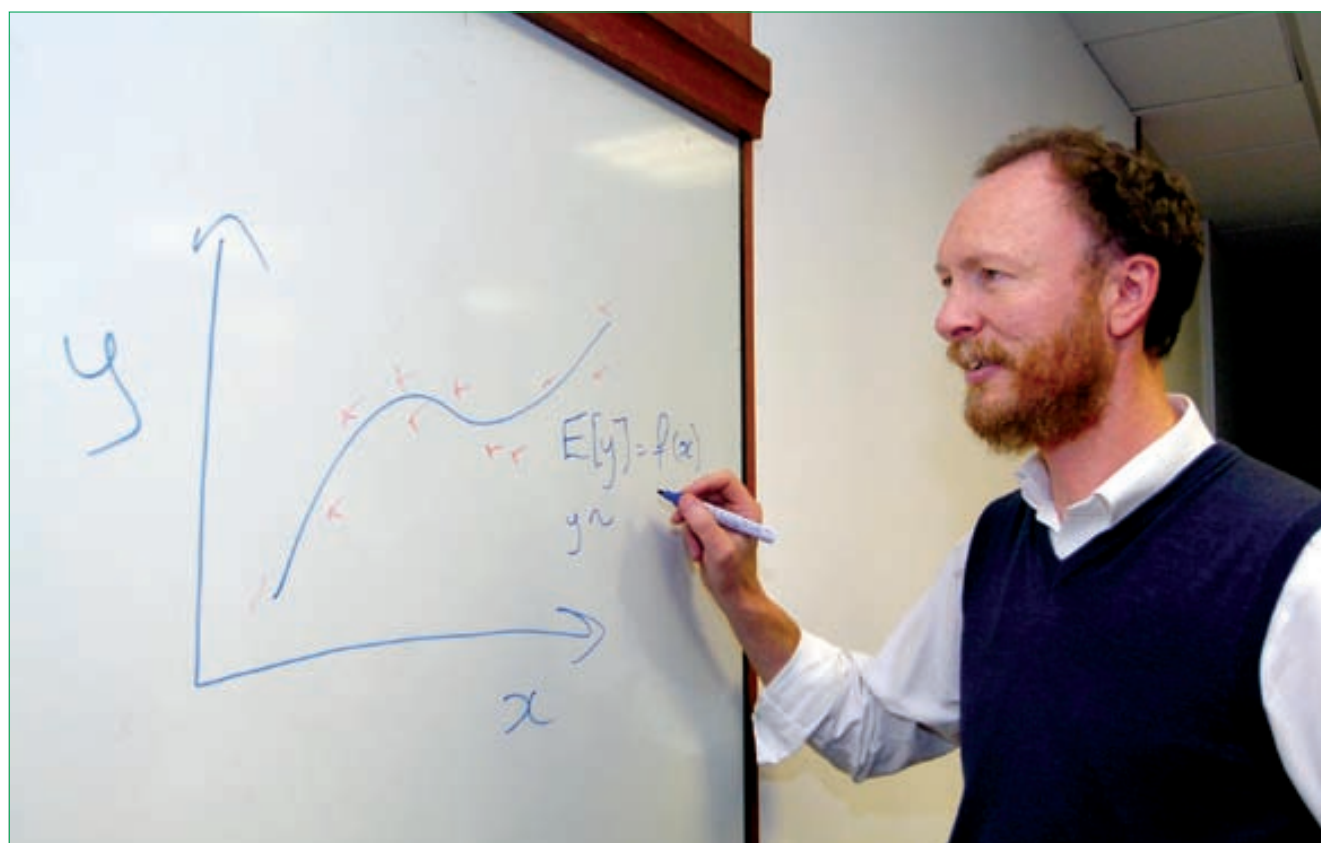
*Biomathematics & Statistics Scotland (BioSS; [www.bioss.ac.uk](http://www.bioss.ac.uk)) delivers research, consultancy and training in statistics, mathematical modelling and bioinformatics. Although BioSS forms part of the SCRI Group, its work is equally relevant to a number of other scientific organisations. Indeed BioSS plays a unique role in the Scottish research community. It bridges the gap between research in the mathematically based and traditionally more qualitative sciences. BioSS staff work within four broad application areas: plant science; animal health and welfare; ecology & environmental science; and human health & nutrition. Our programme of applied strategic research addresses generic issues encountered in these application areas and is managed in three broad themes: statistical bioinformatics; systems & process modelling; and statistical methodology.*

## Improving Quantitative Trait Locus (QTL) analysis for key crop traits

**Chris A. Hackett**

During 2009 BioSS marked up 20 years of collaboration with SCRI scientists in the area of linkage analysis and QTL mapping. The central idea of estimating genetic distances between DNA markers on the basis of recombination between them, and relating variation

at marker positions to variation in phenotypic traits to locate genetic regions affecting these traits, remains unchanged. However we have seen major changes in the types of population, the markers, the traits of interest and the statistical approaches needed. In this review we look back over this progression, and forward to new developments.



David Elston, Director of BioSS.

The first collaboration was an analysis of an  $F_2$  barley population, relating height and ear emergence to two markers on chromosome 4. This required the development of a novel mixture model with two unlinked genes interacting epistatically to control the vernalisation requirement. Subsequent collaborations have involved many more markers, as different techniques have developed: Restriction Fragment Length Polymorphisms (RFLPs); Random Amplified Polymorphic DNA (RAPD); Amplified Fragment Length Polymorphisms (AFLPs); Simple Sequence Repeats (SSRs). Single Nucleotide Polymorphism (SNP) analysis and Diversity Arrays Technology (DArT) now enable linkage maps of barley to be constructed with more than 1000 markers.

Barley is an ideal crop for QTL analysis, as studies on barley typically begin with homozygous parents, where recombination can be estimated with few complications. However research at SCRI has expanded to include QTL studies for all the major crops here. In raspberries and blackcurrants, mapping populations are usually derived by crossing two (outbreeding) parents and

studying their  $F_1$  progeny. Potato has the further complication that the breeders' potato is autotetraploid. BioSS and SCRI obtained funding to develop the theory for mapping in autotetraploid species, and our methods have been incorporated in the software TetraploidMap for Windows. This has been used by other groups to develop linkage maps and locate QTLs in plants such as alfalfa, leek, coffee and rose and will be developed further to meet the needs of a new SCRI project on blueberry.

The phenotypic traits under investigation have also changed in number and type. The first collaboration involved two highly heritable characters. Subsequent studies have looked at yield, and yield components, which are less heritable. More recent traits have involved more complex analysis in advance of the QTL mapping and have required the expertise of BioSS in other areas: for example the analysis of sensory data from a tasting panel assessing different potatoes, and the analysis of colour data using image analysis techniques. A variety of statistical approaches can be

used to look at modelling developmental processes: principal coordinates were used to summarise a series of repeated visual assessments of raspberry ripening, and analysed to identify QTLs affecting different stages of ripening. Another role of the preliminary analysis is to check for spatial trends across field trials, or temporal trends related to the ordering of samples through laboratory testing, which can conceal or bias QTL effects.

Mixed models are being increasingly used in QTL analyses. Their flexible correlation structure means that they can be used to represent the relationship between, say, yields measured in different locations or under different treatment regimes in a factorial experiment, so enabling tests for QTL by environment interactions. These interactions can, in turn, be modelled in a mixed model framework as functions of environmental variables such as temperature or rainfall. Mixed

models can also be used to combine trait values from related populations, taking into account the degree of relatedness: the scale of this can range from combining two or three large mapping populations that share a common parent through to association mapping of germplasm collections.

In the future, the challenges are most likely to come from the increasing dimensions of phenotypic data as 'omics technologies lead to measurements of metabolites or gene expressions, or image analysis methods such as LemnaTec give detailed pictures of crop development. Dealing with these large, potentially multivariate data sets will be challenging both statistically and computationally. We anticipate further developments in the types and quantities of marker data, and changes in the target traits as issues of sustainability and food security in a changing climate become increasingly important.

# Mylnefield Research Services Ltd

**Nigel W. Kerby & Jonathan B. Snape**

Mylnefield Research Services (MRS) Ltd, the wholly-owned commercial subsidiary of SCRI, celebrated its twentieth anniversary in 2009. It was established in 1989 by Professor John Hillman, the then Director of SCRI, to enhance competitiveness, understand and fulfil the needs of industry and maximise the value of SCRI intellectual assets and resources. Agricultural food production has changed radically over this period and the underpinning research has seen significant advances. The need for SCRI to be able to convert its excellent science into commercial products and services that meet industrial and societal needs, while stimulating economic development, has never been

more acute. MRS has consistently generated profits that are reinvested in SCRI. The formation of a new research institute comprising SCRI and the Macaulay Land Use Research Institute in 2011 will ensure we are well positioned to meet the global challenges of food and energy security and climate change.

The twentieth anniversary of MRS was celebrated at the Director's summer soir  e on the lawn at SCRI in August when local dignitaries, key stakeholders, past and present employees and directors enjoyed an evening of fine food (including potatoes and fruit bred by MRS and its partners) and entertainment.



Dr Nigel Kerby with Mr Murray Thomson and Mr Peter Berry.



MRS uses a diverse range of routes to commercialise SCRI expertise and intellectual property (IP), including: Spin out companies, for example EnPrint Ltd.; Joint Ventures, for example Danasia Berry (Beijing) Corporation; collaborative research projects for example long-term breeding contracts with GlaxoSmithKline and Greenvale AP Plc; licensing, for example we have granted 473 licences to plant varieties bred at SCRI; sale of products, for example barley molecular markers, plant virus antibodies; analytical services for example stable isotope analysis, lipid analysis; field and glasshouse services, for example supply of high-health plants, trialling new agrochemicals; consultancy, for example statistical services from BioSS.

#### Finances

The turnover of MRS was £1.9 million in 2008/2009.

MRS transferred £0.9 million to the SCRI Group, including £225k of Gift Aid. As in previous years, contract research (45%) was the biggest contributor to income, followed by royalties (26%) and analytical services (20%). In addition, MRS retained £91k (after tax) to increase significantly company reserves for future investments,

#### Knowledge transfer and exploitation

In March 2009, Dr Richard Gregson joined the MRS team. Richard, a trained patent examiner with a PhD in chemistry, is employed by Genecom and seconded to MRS with the sole aim of generating income from the commercialisation of SCRI's IP and resources. Genecom also provide funds to develop commercial ideas and in 2009, a project led by Dr Wolfram Meier-Augenstein was approved to develop methodologies to determine the authenticity of Scotch whisky using stable isotopes.

#### Analytical services

The early part of 2009 saw difficult trading conditions for Mylnefield Lipid Analysis. The impact of the financial crisis and the resultant recession saw a reduction in plantings of oil seed crops for use in health foods and nutraceuticals, and a major customer in the pharmaceutical sector closed its UK office and terminated its contracts with MRS. The second half

of the year saw business pick up again, and following investment in a new TLC machine that attracted new customers, October 2009 was the most successful month to date for the business.

In addition, MRS was audited by the Medical and Healthcare products Regulatory Authority (MHRA) and can now operate to Good Clinical Practice. MRS is now working towards obtaining Good Manufacturing Practice and Good Laboratory Practice accreditation and has retained the services of Marilyn Emery Consultants to assist in this process.

Under the leadership of Dr Wolfram Meier-Augenstein, the stable isotope analysis business has shown steady growth that has necessitated the investment in new equipment and extra technical assistance. This business has been branded as Mylnefield Isotope Signatures and new marketing material and a website ([www.isotopesignatures.com](http://www.isotopesignatures.com)) have been developed. It is anticipated that combining increased marketing effort with extra analytical capacity will increase sample throughput and lead to further growth in 2010.



The launch of the spin-out company EnPrint©. From left to right: Dr Keith Winton, Dr Rayne Longhurst, Dr Tim Daniell and Dr David M Roberts.

#### Associated businesses

EnPrint Ltd was established in April 2009 with a £150k investment from the Genomia Seed Fund. EnPrint are developing laboratory-based analytical services for analysing complex populations of microorganisms based on methods pioneered by Dr Tim Daniell and reduced to practice by Dr David Roberts. These techniques have attracted considerable interest from



environment agencies and water companies in the UK and elsewhere in Europe. The challenge in 2010 will be to convert this interest into commercial contracts and raise sufficient finance to enable the company to develop a sustainable range of services.

MRS has a 10% stake in a cooperative joint venture, Danasia Berry (Beijing) Corporation Limited. 2009 saw the production of 57 tonnes of fruit in the Hairau district of Beijing and the establishment of new plantations. In addition, there has been a change in emphasis

from summer fruiting varieties to primocane and

significant investments in the infrastructure of farms and juvenile plants. Trials of SCRI 'Glen' varieties were established in 2009 and experiments on overwintering cane to protect from severe frosts were initiated. It is early days for this venture but MRS and our Danish partners – Berrifine Corporation and the Industrialisation Fund for Developing Countries (IFU) – are determined to increase both the quality and quantity of products through selecting appropriate varieties, extension services, transferring new technologies (for example protected cropping and drip irrigation) and training.

### Human Resources

MRS attained a Bronze Award for its Investors in People submission. This accolade is testament to the commitment and dedication of all MRS staff. Of course, there is always room for improvement and in 2010 we will be looking at new ways to improve.

### Plant breeding

MRS's commitment to plant breeding was evidenced by the employment of two new PhD plant breeders. Vanessa Young (potatoes) and Hazel Bull (cereals) will be undergoing rigorous training over the next five years

to ensure that MRS has access to plant breeders with the skills required to exploit fully the data generated by genomics. It is anticipated that in 2010 an additional PhD plant breeder focused on soft fruit will be recruited.

A new raspberry breeding consortium was established in 2009 that has guaranteed funding for raspberry breeding for at least the next five years. The consortium comprises the Scottish Government, the Horticultural Development Company and most of the major raspberry propagators, growers and marketing groups in the UK. For the first time this breeding programme will utilise molecular markers developed at SCRI by Dr Julie Graham and her team. It is hoped that these developments will lead to a new generation of raspberry varieties that build on the market leading 'Glen' series of raspberries.



Finlay Dale (right) with John Bradshaw.

Following the retirement of Dr John Bradshaw in April, Dr Finlay Dale was seconded to MRS for 50% of his time in recognition of his commitment to delivering eight potato breeding programmes. Two major potato breeding programmes with McCain Potato and Greenvale AP were renewed in 2009. The potato breeding team was strengthened by the appointment of Jane Robertson. Two blackcurrant breeding programmes (one fresh, one processing) were ably managed by Dr Rex Brennan and look poised to deliver exciting new varieties in the coming years.

### Licensing and royalties

Royalty income in 2008/2009 was the highest ever at £498k driven mainly by income from the raspberry



Loch Tay

variety Glen Lyon in Spain and Glen Ample in the UK and elsewhere in Europe. Income from blackberries showed strong growth following the release of Loch Tay and income from potatoes showed a steady increase due to the popularity of the new 'Vales' varieties marketed by our long-standing partner Greenvale AP Plc.

In 2009, three new potato varieties (Inca Bella, Chinchá and Tabitha) were admitted on to the National List and two new varieties were submitted for National List Trials. EU Plant Variety Rights were applied for in relation to three varieties of blackcurrants, one potato variety and two varieties of the catch crop *Solanum sisymbriifolium*. Following the success of the arrangement to collect

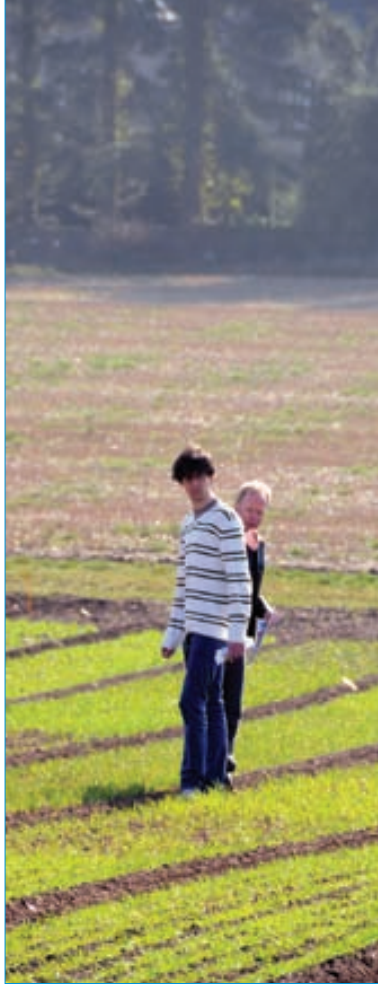
royalties from Glen Lyon in Spain, in late 2009 MRS concluded an agreement with the Spanish company Eurosemillas to act as agent for MRS raspberries and blackberries in Spanish and Portuguese speaking countries around the world.

#### Contract research

Despite the recession, income from contract research managed to grow by 18%. This is despite one potato breeding company going into administration and several potential major collaborations being put on hold pending more favourable economic conditions. Although many SCRI scientists contributed to this impressive performance, special mention should be made of Dr Alison Lees (potato pathology), Dr Julie Graham (soft fruit genetics) and Dr Derek Stewart (food quality).

#### Acknowledgements

MRS gratefully acknowledges the support of all SCRI staff, for their significant contribution to the success of the company. MRS would also like to thank its customers and sponsors for their continued support. Finally, we would like to thank the non-executive Directors of MRS who contribute their valuable time without recompense.





# Division of Plant Sciences University of Dundee

**John W. S. Brown**

*The Division of Plant Sciences has been very successful in attracting external research grant funding of over £9 million over the last 18–24 months. The grants are mainly from the BBSRC but also include EU funding and a major grant from the Global Climate and Energy project (GCEP) and will run for between 3–5 years. Claire Halpin has been the major contributor with £5.5M for projects on lignin biosynthesis and recombination. One aspect of much of this funding is the collaboration with SCRI research groups reflecting the special partnership between the College of Life Sciences of the University of Dundee and SCRI.*

A major highlight was the January launch of the BBSRC Sustainable Bioenergy Centre (BSBEC), a virtual Centre with six research hubs. The Dundee centre is headed by Claire Halpin and is a collaboration with Robbie Waugh and Derek Stewart at SCRI. BSBEC is supported by £28 million of BBSRC funding, has many links with industrial partners and is focused on complementary research areas relevant to sustainable bioenergy.

We are pleased to welcome a new Fellow of the Royal Society of Edinburgh, Edgar Huitema, who joined the Division in October. His area of research is the function of a new class of effector proteins, the crinklers, in the plant pathogen, *Phytophthora*. He is associated with the Plant Pathology programme of SCRI and his work will complement that of Paul Birch in advancing our understanding of effectors and the defence



mechanisms that they target for suppression. This year, Paul Birch's research group was one of the leading groups involved in obtaining the genome sequence of *Phytophthora infestans*, the major pathogen of potato (published earlier this year in *Nature*). We also say a sad goodbye to Lyn Jones, Professor of Plant Ecology, who retired at the end of September. Lyn will remain active as he follows ongoing interests and collaborations.

The grant income success has also meant a rapid increase in the size of the Division to over 45 people. A challenge has been to reinforce the identity of the Division while many of its Principal Investigators are situated in different parts of the SCRI campus – mainly to reflect their research areas and opportunities for interaction and collaboration. The recent refurbishment of a corridor which houses three of the Plant Sciences groups alongside SCRI colleagues, has allowed many of the newly recruited scientists and technicians to

be accommodated. In addition, it has also created a central hub for the Division where Morven Pearson (the Divisional Secretary) and Sandie Gray (recently appointed as Lab Manager) are situated alongside John Brown and Claire Halpin (the Head and Deputy Head of Division).





The Masters level MRes teaching course in 'Crops for the Future' began this October with its first intake of four students from Scotland, Poland, Pakistan and India. The course is a joint venture between the College of Life Sciences/Division of Plant Sciences and SCRI and builds on the unique combination of expertise of both organisations. Our MRes is highly topical as issues of food security and sustainability continue to dominate scientific policy. It aims to give students a strong grounding in modern plant and crop biology including genetics, genomics and new approaches to breeding and crop improvement.

Other scientific highlights include Gordon Simpson's work on the function of FPA in flowering time control and the demonstration that it controls RNA 3' end formation which has led to a successful grant to look at wider aspects of mRNA 3' processing in regulating



expression. John Brown's group has discovered mRNAs and aberrant mRNAs in the nucleolus raising the possibility of novel functions for the nuclear compartment in regulating plant gene expression. Andy Flavell has developed a PCR based technology for Next Generation Sequencing of hundreds of candidate gene alleles in hundreds of barley genotypes as a basis for identifying alleles responsible for traits important to the breeder. Finally, Steve Hubbard has developed a number of interactions with SCRI colleagues in the Environment Plant Interactions Programme in particular looking at the dynamics of the interaction between the potato aphid (*Macrosiphon euphorbiae*) and its parasitoids, endosymbionts and plant pathogens and the impact of secondary bacterial endosymbionts on fitness of the cabbage aphid (*Brevicoryne brassicae*) and its capacity to resist parasitism and to transmit viral pathogens.



# Postgraduate studies at SCRI

**Craig G. Simpson & Tracy A. Valentine**

Postgraduate training at SCRI provides an environment that allows technical and intellectual competences to flourish and equips students with modern scientific skills suitable for a range of scientific careers. Training in research skills and techniques is an essential element in the development of a research student. Students have the opportunity to use the considerable knowledge, techniques and technologies available at SCRI and their associated universities. The outcome is a partnership that makes a substantial and original contribution to knowledge in the research areas at SCRI.

The Institute is involved in the training of around 60 students at present, with about half supported through the very successful and highly competitive joint studentship programme. This programme is funded jointly through SCRI and the student's university. It has led to the highest student population at SCRI to date. This highly motivated group enhances scientific life at SCRI and positively challenges SCRI staff through vibrant scientific questioning and by developing novel techniques. The rigorous and demanding nature of the studentship is monitored through a process that







Susan McCallum won the Innovation Prize at the 2009 Student Competition.

requires a written report and short presentation with a selected impartial scientific advisor every six months. These meetings challenge the student with a different point of view and provide a record of their scientific and academic progress. The meetings are complementary to the monitoring system at the University at which the postgraduate student is registered.

The students at SCRI have established a committee that holds meetings and offers a chance for students to discuss aspects of postgraduate life and concerns. Two students are also members of the Institute University Interactions Committee (UIC) that deals with all aspects of postgraduate research at SCRI. They provide a valuable link between students and the function of the UIC. Informative occasional presentations were given in 2009 by members of staff to give advice on topics and skills relevant to postgraduate research.

SCRI played host to this year's Postgraduate Student Competition as part of the Scottish Research Institutes' annual Science for Life lecture. The competition involved an incredibly high standard of talks, confidently

presented and defended. This was very much a celebration of the work going on around the Scottish Research Institutes. SCRI was represented by Susan McCallum 'Marker assisted breeding for sensory characteristics in red raspberry (*Rubus idaeus* L.)' and Lindsay McMenemy 'Plant mediated interactions with the large raspberry aphid', our winners of the internal SCRI student competition. Lindsay went on to win the competition following on from her success as runner-up in the 2008 Student Essay Competition run by the Royal Entomological Society.



The moment SCRI's Lindsay McMenemy learned she was the winner of the 2009 Scottish Research Institutes' Student PhD competition.

This year we welcomed postgraduate students to the Master of Research (MRes) course titled 'Crops for the Future' for the first time. This is a joint venture initiated between the University of Dundee College of Life Sciences and SCRI. The course provides a framework for advanced study in practical and theoretical aspects of crop bioscience through lectures from SCRI and Dundee University staff and through practical demonstration of modern agricultural and plant science practices.

# The Scottish Society for Crop Research

## Bill Macfarlane Smith

The Society has a role in raising awareness of items of interest and concern to the agriculture, horticulture and forestry industries. This is partially achieved by funding the Annual SSCR Lecture immediately following the Annual General Meeting. This year's AGM was held on Wednesday 27 May and the Annual Lecture was given by Professor Annie Anderson of the Centre of Public Health and Nutrition Research, the Division of Therapeutics and Medicine at the University of Dundee Ninewells Hospital and Medical School. The title of the lecture was, 'The challenge of achieving a healthy and sustainable diet in Scotland'.

The Society currently supports research on a range of topics, including research into differential cultivar response to soil physical constraints in winter and spring barley, integrated pest and pathogen manage-

ment in raspberry, the possible exploitation of avoidance cues used by vine and clay coloured weevils, the genetic cause of crumbly fruit in raspberry, and a long running programme of support for raspberry breeding. Other work supported includes training in methods of barley transformation, the assessment of the effect of adverse weather conditions on distilling quality in wheat and support for a study of techniques to reduce pollution from tramlines in combinable crops. Funding continues on a biennial basis for a keynote speaker at the Crop Protection in Northern Britain Conference next to be held 23–24 February 2010.

All meetings organised by the Society were well supported, in virtually every case attracting greater numbers than in previous years.

'Cereals in Practice' was held on 9 July 2009 at the Institute's new Balruddery Farm, and was attended by 207 visitors. This combined event with SAC included static demonstrations of machinery as well as variety and other plant demonstrations.

The 'Fruit for the Future' event held on 16 July 2009 attracted 120 breeders, growers, end-users and processors. The increased attendance was so large that existing facilities are no longer adequate and alternatives are being considered for future years. Research on raspberries, strawberries, blackcurrants and gooseberries was displayed, and a useful opportunity provided for the tasting assessment of prospective new varieties of raspberry.



Fruit for the Future, 2009.



Cereals in Practice, 2009.

'Potatoes in Practice' was held on 13 August, also located at the new Balruddery Farm. This was attended by over 650 growers, breeders, scientists and advisers, which probably represents about the maximum numbers for such an outdoor event in northern Britain. Support was again provided by Potato Council Ltd, SAC, CSC Crop Protection Ltd, SCRI and the Society.



Potatoes in Practice

Demonstrations on cultivars, agronomy and related research on pests and diseases of the crop, continued to be major features, along with a further increase in static demonstrations of potato crop machinery and other equipment.

The Potato and Soft Fruit Crop Sub-Committees held other half day meetings during the year to provide information on current research and permit interaction between end users, growers, processors and scientists.

The Combinable and Energy Crops winter event at Battleby on 17 February attracted over 70 delegates who listened to an excellent series of presentations on matters of current concern to the cereal industry, including the dramatic changes to pesticide availability.

In the year ahead, the Society will be reviewing its remit, to ensure that it is fit for the 21st century and continues to meet the objectives and aspirations of its members.



# Publications

Publications listed between 1 October 2008 and 31 September 2009 are classified in the following manner:

- J Papers describing original research in refereed journals.
- R Books, book chapters and reviews in books – providing each has been edited externally
- P Published proceedings of contributions to conferences or learned societies (including published abstracts)
- O Popular articles and unrefereed publications

**Aalders, I., Hough, R.L., Towers, W., Black, H.I.J., Ball, B.C., Griffiths, B.S., Hopkins, D.W., Lilly, A., McKenzie, B.M., Rees, R.M., Sinclair, A. & Campbell, C.D.** 2009. Considerations for Scottish soil monitoring in the European context. *European Journal of Soil Science* **60**, 833-843. <http://dx.doi.org/10.1111/j.1365-2389.2009.01183.x> J

**Abecia, L., Balcells, J., Fondevila, M., Belenguer, A., Holtrop, G. & Lobley, G.E.** 2008. Contribution of gut microbial lysine to liver and milk amino acids in lactating does. *British Journal of Nutrition* **100**, 977-983. <http://dx.doi.org/10.1017/S0007114508957986> J

**Adam, M.A.M., Phillips, M.S., Tzortzakakis, M. & Blok, V.C.** 2009. Characterisation of *mjap* genes encoding novel secreted proteins from the root-knot nematode, *Meloidogyne javanica*. *Nematology* **11**, 253-265. <http://dx.doi.org/10.1163/156854109X429583> J

**Al-Mrabeh, A., Ziegler, A., Cowan, G.H. & Torrance, L.** 2009. A fully recombinant ELISA using in vivo biotinylated antibody fragments for the detection of *Potato leaf roll virus*. *Journal of Virological Methods* **159**, 200-205. <http://dx.doi.org/10.1016/j.jviromet.2009.03.025> J

**Andrews, J., Lynn, J., Parsons, N., Ryder, C., White, P.J. & Thompson, A.J.** 2008. Genetic analysis of root traits for water capture in the genus *Solanum*. *Association of Applied Biologists, Resource Capture by Crops: Integrated Approaches*, Sutton Bonington, UK, 10-12 September 2008 (Poster). P

**Andrews, M., Lea, P.J., Raven, J.A. & Azevedo, R.A.** 2009. Nitrogen use efficiency. 3. Nitrogen fixation: genes and costs. *Annals of Applied Biology* **155**, 1-13. <http://dx.doi.org/10.1111/j.1744-7348.2009.00338.x> J

**Antunez-Lamas, M., Cabrera, E., Lopez-Solanilla, E., Solano, R., Gonzalez-Melendi, P., Chico, J.M., Toth, I.K., Birch, P.R.J., Pritchard, L., Liu, H. & Rodriguez-Palenzuela, P.** 2009. Bacterial chemoattraction towards jasmonate plays a role in the entry of *Dickeya dadantii* through wounded tissues. *Molecular Microbiology* **74**, 662-671. <http://dx.doi.org/10.1111/j.1365-2958.2009.06888.x> J

**Ashman, M.R., Hallett, P.D., Brookes, P.C. & Allen, J.** 2009. Evaluating soil stabilisation by biological processes using step-wise aggregate fractionation. *Soil and Tillage Research* **102**, 209-215. <http://dx.doi.org/10.1016/j.still.2008.07.005> J

**Askarianzadeh, A., Birch, A.N.E., Ramsay, G. & Minaeimoghadam, M.** 2009. Study of wild *Solanum* species to identify sources of resistance against the green potato aphid, *Myzus persicae*. *Plant Protection and Plant Health in Europe*, Berlin, Germany, 14-16 May 2009. P

**Avrova, A.O., Boevink, P.C., Young, V., Grenville-Briggs, L.J., van West, P., Birch, P.R.J. & Whisson, S.C.** 2008. A novel *Phytophthora infestans* haustorium-specific membrane protein is required for infection of potato. *Cellular Microbiology* **10**, 2271-2284. <http://dx.doi.org/10.1111/j.1462-5822.2008.01206.x> J

**Baccini, M., Bachmaier, E.M., Biggeri, A., Boekschoten, M.V., Bouwman, F.G., Brennan, L., Caesar, R., Cinti, S., Coort, S.L., Crosley, K., Daniel, H., Drevon, C.A., Duthie, S., Eijssen, L., Elliott, R.M., van Erk, M., Evelo, C., Gibney, M., Heim, C., Horgan, G.W., Johnson, I.T., Kelder, T., Kleemann, R., Kooistra, T., van Iersel, M.P., Mariman, E.C., Mayer, C., McLoughlin, G., Müller, M., Mulholland, F., van Ommen, B., Polley, A.C., Pujos-Guillot, E., Rubio-Aliaga, I., Roche, H.M., de Roos, B., Sailer, M., Tonini, G., Williams, L.M. & de Wit, N.** 2008. The NuGO proof of principle study package: a collaborative research effort of the European Nutrigenomics Organisation. *Genes and Nutrition* **3**, 147-151. <http://dx.doi.org/10.1007/s12263-008-0102-5> J

**Baggaley, N.J., Langan, S.J., Fitter, M.N., Potts, J.M. & Dunn, S.M.** 2009. Long-term trends in hydro-climatology of a major Scottish mountain river. *Science of the Total Environment* **407**, 4633-4641. <http://dx.doi.org/10.1016/j.scitotenv.2009.04.015> J

**Bahar, B., Moloney, A.P., Monahan, F.J., Harrison, S.M., Zazzo, A., Scrimgeour, C.M., Begley, I.S. & Schmidt, O.** 2009. Turnover of carbon, nitrogen and sulfur in bovine longissimus dorsi and psoas major muscles: Implications for isotopic authentication of meat. *Journal of Animal Science* **87**, 905-913. <http://dx.doi.org/10.2527/jas.2008-1360> J

**Barker, H., McGeachy, K.D., Toplak, N., Gruden, K., Žel, J. & Browning, I.** 2009. Comparison of genome sequence of PVY isolates with biological properties. *American Journal of Potato Research* **86**, 227-238. <http://dx.doi.org/10.1007/s12230-009-9076-0> J

**Barnett, C.M., Bengough, A.G. & McKenzie, B.M.** 2009. Quantitative image analysis of earthworm-mediated soil displacement. *Biology and Fertility of Soils* **45**, 821-828. <http://dx.doi.org/10.1007/s00374-009-0392-9> J

**Barré, P. & Hallett, P.D.** 2009. Rheological stabilisation of wet soils by model root and fungal exudates depends on clay mineralogy. *European Journal of Soil Science* **60**, 525-538. <http://dx.doi.org/10.1111/j.1365-2389.2009.01151.x> J

**Barré, P., McKenzie, B.M. & Hallett, P.D.** 2009. Earthworms bring compacted and loose soils to a similar mechanical state. *Soil Biology and Biochemistry* **41**, 656-658. <http://dx.doi.org/10.1016/j.soilbio.2008.12.015> J



- Bartley, P.M., Wright, S.E., Maley, S.W., Buxton, D., Nath, M. & Innes, E.A.** 2009. The development of immune responses in Balb/c mice following inoculation with attenuated or virulent *Neospora caninum* tachyzoites. *Parasite Immunology* **31**, 392-401. <http://dx.doi.org/10.1111/j.1365-3024.2009.01115.x> J
- Basnayake, S., Maclean, D.J., Whisson, S.C. & Drenth, A.** 2009. Identification and occurrence of the LTR-*Copia*-like retrotransposon, *PSCR* and other *Copia*-like elements in the genome of *Phytophthora sojae*. *Current Genetics* **55**, 521-536. <http://dx.doi.org/10.1007/s00294-009-0263-9> J
- Battino, M., Beekwilder, J., Denoyes-Rothan, B., Laimer, M., McDougall, G.J. & Mezzetti, B.** 2009. Bioactive compounds in berries relevant to human health. *World Conference of Public Health Nutrition*, Barcelona, Spain, 25-27 September 2006. *Nutrition Reviews* **67**, S145-S150. <http://dx.doi.org/10.1111/j.1753-4887.2009.00178.x> P
- Beardall, J., Allen, D., Bragg, J., Finkel, Z.V., Flynn, K.J., Quigg, A., Rees, T.A.V., Richardson, A. & Raven, J.A.** 2009. Allometry and stoichiometry of unicellular, colonial and multicellular phytoplankton. *New Phytologist* **181**, 295-309. <http://dx.doi.org/10.1111/j.1469-8137.2008.02660.x> J
- Bengough, A.G.** 2009. Scaling root growth responses from seedlings to field. *Society for Experimental Biology Annual Main Meeting*, Glasgow, UK, 28 June-1 July 2009. P
- Bengough, A.G., Valentine, T.A., McKenzie, B.M., Hallett, P.D., Dietrich, R., White, P.J. & Jones, H.G.** 2009. Physical limitations to root growth: screening, scaling and reality. *7th International Symposium of the Society for Root Research: Root Research and Applications*, Vienna, Austria, 2-4 September 2009 (Abstract). P
- Berry, P. & White, P.J.** 2008. Components of variation in nitrogen use efficiency in *B. napus*. *Oilseed Rape Genetic Improvement Network (OREGIN) 6th Stakeholder Forum Meeting on "Assessing low input systems for breeding and trialling - the N and P economy"*, NIAB, Cambridge, UK, 21 November 2008 (Talk). P
- Berry, P., Foulkes, J., White, P.J., Spink, J. & Teakle, G.** 2008. Breeding for improved nitrogen use efficiency in oilseed rape. *Association of Applied Biologists, Resource Capture by Crops: Integrated Approaches*, Sutton Bonington, UK, 10-12 September 2008 (Talk). P
- Berry, P., Teakle, G., Foulkes, J., White, P.J. & Spink, J.** 2008. Breeding for improved nitrogen use efficiency in oilseed rape. *5th ISHS International Symposium on Brassicas and 16th Crucifer Genetics Workshop*, Lillehammer, Norway, 8-12 September 2008, 40 (Talk). P
- Bingham, I.J. & Newton, A.C.** 2009. Crop tolerance of foliar pathogens: possible mechanisms and potential for exploitation. In: D. Walters, ed. *Disease Control in Crops: Biological and Environmentally Friendly Approaches*, Wiley-Blackwell, Chichester, UK, 142-161. R
- Bingham, I.J., Rees, R.M. & Bengough, A.G.** 2009. Influence of soil compaction on the dynamics of root growth and mortality in spring barley. *7th International Symposium of the Society for Root Research: Root Research and Applications*, Vienna, Austria, 2-4 September 2009. P
- Bingham, I.J., Wu, L.F., Thomas, W.T.B., White, P.J. & Hackett, C.A.** 2009. Nitrogen use efficiency of spring barley: changes associated with over 75 years of barley breeding. *MONOGRAM Network Workshop*, Burwalls, Bristol, UK, 29 April-1 May 2009. P
- Birch, A.N.E.** 2009. Book Review: Crop Wild Relative Conservation and Use. Edited by N. Maxted, B.V. Ford-Lloyd, J.M. Iriando, M.E. Dulloo & J. Turk. CABI, Wallingford (2008). *Experimental Agriculture* **45**, 237. <http://dx.doi.org/10.1017/S001447970800731X> O
- Birch, A.N.E.** 2009. Book review: Integrated Pest Management. Concepts, Tactics, Strategies and Case Studies. Edited by E.B. Radcliffe, W.D. Hutchinson & R.E. Cancelada. CUP, Cambridge, (2009). *Experimental Agriculture* **45**, 511-512. <http://dx.doi.org/10.1017/S0014479709990317> O
- Birch, A.N.E.** 2009. Innovative trap for raspberry growers. *The Fruit Grower* April 2009, 33-34. O
- Birch, A.N.E.** 2009. IPM in ecological engineering: using pest-resistant crops with sustainable biocontrol. *ISBCA 09, 3rd International Symposium on Biological Control of Arthropods*, Christchurch University, New Zealand, 8-13 February 2009. P
- Birch, A.N.E. & Allen, J.** 2009. Pests in the firing line. *Fresh Produce Journal* July 2009, 36-37. O
- Birch, A.N.E., Shepherd, T., Hancock, R.D. & Goszcz, K.** 2009. Understanding sugar sensing in induced plant defences and stress tolerance. *International Society of Chemical Ecology*, Neuchâtel, Switzerland, 23-25 August 2009. P
- Birch, P.R.J. & Avrova, A.O.** 2009. Gene expression profiling. In: K.H. Lamour & S. Kamoun, eds. *Oomycete Genetics and Genomics: Diversity, Interactions and Research Tools*, Wiley-Blackwell, Hoboken, New Jersey, USA, 477-492. R
- Birch, P.R.J., Armstrong, M., Bos, J., Boevink, P., Gilroy, E.M., Taylor, R.M., Wawra, S., Pritchard, L., Conti, L., Ewan, R., Whisson, S.C., van West, P., Sadanandom, A. & Kamoun, S.** 2009. Towards understanding the virulence functions of RXLR effectors of the oomycete plant pathogen *Phytophthora infestans* (Review). *Journal of Experimental Botany* **60**, 1133-1140. <http://dx.doi.org/10.1093/jxb/ern353> J
- Blower, T.R., Fineran, P.C., Johnson, M.J., Toth, I.K., Humphreys, D.P. & Salmond, G.P.C.** 2009. Mutagenesis and functional characterisation of the RNA and protein components of the *toxIN* abortive infection and toxin-antitoxin locus of *Erwinia*. *Journal of Bacteriology* **191**, 6029-6039. <http://dx.doi.org/10.1128/JB.00720-09> J
- Bowen, S.R., Gregorich, E.G. & Hopkins, D.W.** 2009. Biochemical properties and biodegradation of dissolved organic matter from soils. *Biology and Fertility of Soils* **45**, 733-742. <http://dx.doi.org/10.1007/s00374-009-0387-6> J
- Bradshaw, J.E.** 2008. Looking for the next number one potato cultivar in Great Britain. *SAC Information Note* **105**. O
- Bradshaw, J.E.** 2009. A genetic perspective on yield plateau in potato (Review). *Potato Journal (Indian)* **36**, 79-94. J
- Bradshaw, J.E.** 2009. Breeding for field resistance to late blight of potato at SCRI. *III International Late Blight Conference*, Beijing, China, 3-6 April 2008. *Acta Horticulturae* **834**, 87-100. P
- Bradshaw, J.E. & Ramsay, G.** 2009. Potato origin and production. In: J. Singh & L. Kaur, eds. *Advances in Potato Chemistry and Technology*, Academic Press, Burlington, MA, USA, 1-26. R



- Bradshaw, J.E., Dale, M.F.B. & Mackay, G.R.** 2009. Improving the yield, processing quality and disease and pest resistance of potatoes by genotypic recurrent selection. *18th General Congress of the European Association for Research on Plant Breeding*, Valencia, Spain, 9-12 September 2009. *Euphytica* **170**, 215-227. <http://dx.doi.org/10.1007/s10681-009-9925-4> P
- Bradshaw, J.E., Titley, M. & Wilson, R.N.** 2009. Single seed descent as a breeding method for swedes (*Brassica napus* L. var. *napobrassica* Peterm.). *Euphytica* **169**, 387-401. <http://dx.doi.org/10.1007/s10681-009-9971-y> J
- Braga, R.A., Dupuy, L., Pasqual, M. & Cardoso, R.R.** 2009. Live biospeckle laser imaging of root tissues. *European Biophysics Journal* **38**, 679-686. <http://dx.doi.org/10.1007/s00249-009-0426-0> J
- Brennan, R.M.** 2009. Book Review: Encyclopedia of Fruit and Nuts. Edited by J. Janick & R.E. Paull. CABI, Wallingford (2008). *Experimental Agriculture* **45**, 238. <http://dx.doi.org/10.1017/S0014479708007345> O
- Brennan, R.M.** 2009. Book Review: The Peach: Botany, Production and Uses. Edited by D.R. Layne & D. Bassi. CABI, Wallingford (2008). *Experimental Agriculture* **45**, 379-380. <http://dx.doi.org/10.1017/S0014479709007777> O
- Brennan, R.M., Jorgensen, L., Gordon, S.L., Loades, K., Hackett, C.A., & Russell, J.R.** 2009. The development of a PCR-based marker linked to resistance to the blackcurrant gall mite (*Cecidophyopsis ribis* Acari: Eriophyidae). *Theoretical and Applied Genetics* **118**, 205-211. <http://dx.doi.org/10.1007/s00122-008-0889-x> J
- Brierley, J.L., Stewart, J.A. & Lees, A.K.** 2009. Quantifying potato pathogen DNA in soil. *Applied Soil Ecology* **41**, 234-238. <http://dx.doi.org/10.1016/j.apsoil.2008.11.004> J
- Broadley, M.R., Fairweather-Tait, S.J., Foot, I., Hart, D.J., Hurst, R., Knott, P., McGrath, S.P., Meacham, M.C., Norman, K., Mowat, H., Scott, P., Stroud, J.L., Tucker, M., White, P.J. & Zhao, F.J.** 2009. Biofortification of UK wheat with selenium. *Medical Geology 2009, British Geological Survey*, Nottingham, UK, 19-20 March 2009. P
- Broadley, M.R., Hammond, J.P., King, G.J., Bowen, H.C., Hayden, R., Spracklen, W.P., Lochlainn, S. & White, P.J.** 2009. Biofortifying *Brassica* with calcium (Ca) and magnesium (Mg). *Abstracts and Programme of the XVI International Plant Nutrition Colloquium: Plant Nutrition for Sustainable Development and Global Health*, Sacramento, California, USA, 26-30 August 2009. 203 P
- Broadley, M.R., King, G., Hammond, J.P. & White, P.J.** 2008. Improving nutrient acquisition and mineral content in vegetable *Brassica*. *University of Nottingham/Chinese Vegetable Study*, Nottingham, UK, 11 December 2008 (Talk). P
- Broadley, M.R., White, P.J., Hammond, J.P., Graham, N.S., Bowen, H.C., Emmerson, Z.F., Fray, R.G., Iannetta, P.P.M., McNicol, J.W. & May, S.T.** 2008. Evidence of neutral transcriptome evolution in plants. *New Phytologist* **180**, 587-593. <http://dx.doi.org/10.1111/j.1469-8137.2008.02640.x> J
- Broadley, M.R., White, P.J., Hammond, J.P., Graham, N., Bowen, H.C., Emmerson, Z.F., Fray, R.G., Iannetta, P.P.M., McNicol, J.W. & May, S.** 2008. Evidence of neutral transcriptome evolution in plants. *BBSRC Systems Biology Grantholder Workshop*, Nottingham, UK, 15-17 December 2008 (Poster). P
- Brown, J.W.S., Simpson, C.G., Lewandowska, D., Clark, G., Fuller, J., Kim, S.-H., Pendle, A., Shaw, P.J. & Koroleva, O.** 2009. The plant nucleolus, mRNA and nonsense-mediated decay. *Eurasnet UK RNA Splicing Workshop*, Lake District, UK, 23-25 January 2009, 10. P
- Brown, K.L., Wathne, G.J., Sales, J., Bruce, M.E. & Mabbott, N.A.** 2009. The effects of host age on follicular dendritic cell status dramatically impair scrapie agent neuroinvasion in aged mice. *Journal of Immunology* **183**, 5199-5207. <http://dx.doi.org/10.4049/jimmunol.0802695> J
- Brown, L.K., George, T.S., Wishart, J., Thompson, J.A., Wright, G.M., Thomas, W.T.B., Lyon, J., Forster, B.P., Alexander, J. & White, P.J.** 2009. Genotypic variation in root traits involved in phosphorus utilization by barley. *7th International Symposium of the Society for Root Research: Root Research and Applications*, Vienna, Austria, 2-4 September 2009 (Abstract). P
- Brueggeman, R., Druka, A., Nirmala, J., Cavileer, T., Drader, T., Rostoks, N., Mirlohi, A., Bennypaul, H., Gill, U., Kudrna, D., Whitelaw, C., Kilian, A., Han, F., Sun, Y., Gill, K., Steffenson, B. & Kleinhofs, A.** 2008. The stem rust resistance gene *Rpg5* encodes a protein with nucleotide binding site, leucine-rich and protein kinase domains. *Proceedings of the National Academy of Sciences, USA* **105**, 14970-14975. <http://dx.doi.org/10.1073/pnas.0807270105> J
- Butler, A., Doherty, R.M. & Marion, G.** 2009. Model averaging to combine simulations of future global vegetation carbon stocks. *Environmetrics* **20**, 791-811. <http://dx.doi.org/10.1002/env.953> J
- Campbell, R., Morris, W.L., Ducreux, L., Ramsay, G., Bryan, G.J. & Taylor, M.A.** 2009. The role of carotenoid cleavage products in tuber apical meristem activation and deactivation processes. *4th Plant Dormancy Symposium*, Fargo, North Dakota, USA, 8-10 June 2009 (Poster). P
- Canto, T., Aranda, M.A. & Fereres, A.** 2009. Climate change effects on physiology and population processes of hosts and vectors that influence the spread of hemipteran-borne plant viruses. *Global Change Biology* **15**, 1884-1894. <http://dx.doi.org/10.1111/j.1365-2486.2008.01820.x> J
- Caron-Lormier, G., Bohan, D.A., Hawes, C., Raybould, A., Houghton, A.J. & Humphry, R.W.** 2009. How might we model an ecosystem? *Ecological Modelling* **220**, 1935-1949. <http://dx.doi.org/10.1016/j.ecolmodel.2009.04.021> J
- Chapman, S.N., Faulkner, C., Kaiserli, E., Garcia-Mata, C., Savenkov, E.I., Roberts, A.G., Oparka, K.J. & Christie, J.M.** 2008. The photoreversible fluorescent protein iLOV outperforms GFP as a reporter of plant virus infection. *Proceedings of the National Academy of Sciences, USA* **105**, 20038-20043. <http://dx.doi.org/10.1073/pnas.0807551105> J
- Chatterjee, A., Horgan, G.W. & Theobald, C.M.** 2008. Exposure assessment for pesticide intake from multiple food products: a Bayesian latent-variable approach. *Risk Analysis* **28**, 1727-1736. <http://dx.doi.org/10.1111/j.1539-6924.2008.01124.x> J
- Chen, C., Hajirezaei, M.R., Zhan, M.I., Hornyik, C., Debast, S., Lacomme, C., Fernie, A.R., Sonnewald, U. & Börnke, F.** 2008. RNA interference-mediated repression of sucrose-phosphatase in transgenic potato tubers (*Solanum tuberosum*) strongly affects the hexose-to-sucrose ratio upon cold storage with only minor effects on total soluble carbohydrate accumulation. *Plant Cell and Environment* **31**, 165-176. <http://dx.doi.org/10.1111/j.1365-3040.2007.01747.x> J

- Chen, X., Griffiths, B.S., Daniell, T.J., Neilson, R. & O'Flaherty, V.** 2009. High-throughput sequencing of soil nematode communities for ecological research. *British Society of Soil Science and Soil Science Society of Ireland Autumn Meeting*, Wexford, Ireland, 9-11 September 2009. P
- Chichkova, N.V., Galiullina, R.A., Taliansky, M.E. & Vartapetian, A.B.** 2008. Tissue disruption activates a plant caspase-like protease with TATD cleavage specificity. *Plant Stress* **2**, 89-95. J
- Chung, B.N. & Palukaitis, P.** 2009. Production of PVY-specific small interfering RNAs and resistance to PVY in transgenic tobacco expressing multiple, fused, virus-derived double-stranded RNAs. *Association of Applied Biologists. Advances in Plant Virology*, Harrogate, UK, 1-3 April 2009. P
- Clark, E., Karley, A.J., Daniell, T.J., Wishart, J. & Hubbard, S.F.** 2008. The bacteria associated with the cabbage aphid (*Brevicoryne brassicae*): their community composition and influence on aphid performance. *Entomological Society of America Annual Meeting*, Reno, NV, USA, 16-19 November 2008. P
- Clark, E., Karley, A.J., Daniell, T.J., Wishart, J. & Hubbard, S.F.** 2009. The community composition and influence on aphid performance of the bacteria associated with the cabbage aphid, *Brevicoryne brassicae*. *8th International Symposium on Aphids*, Catania, Italy, 8-12 June 2009. P
- Clark, K.E., Koricheva, J., Jones, T.H., Hartley, S.E. & Johnson, S.N.** 2009. Are there patterns in above and belowground insect herbivore interactions? A meta-analysis approach. *Ento'09 National Meeting of the Royal Entomological Society*, University of Sheffield, Sheffield, UK, 15-17 July 2009. P
- Clayton, S.J., Read, D.B., Murray, P.J. & Gregory, P.J.** 2008. Exudation of alcohol and aldehyde sugars from roots of defoliated *Lolium perenne* L. grown under sterile conditions. *Journal of Chemical Ecology* **34**, 1411-1421. <http://dx.doi.org/10.1007/s10886-008-9536-x> J
- Coates, E.M., McDougall, G.J. & Shepherd, T.** 2009. Metabolomic profiling of HT29 colon adenocarcinoma cells. *NUGO SYSDIET Metabolomics Workshop*, Copenhagen, Denmark, 24-27 February 2009 (Poster). P
- Cock, P.J.A., Antao, T., Chang, J.T., Chapman, B.A., Cox, C.J., Dalko, A., Friedberg, I., Hamelryck, T., Kauff, F., Wilczynski, B. & de Hoon, M.J.L.** 2009. Biopython: freely available Python tools for computational molecular biology and bioinformatics. *Bioinformatics* **25**, 1422-1423. <http://dx.doi.org/10.1093/bioinformatics/btp163> J
- Cockell, C.S., Kaltenegger, L. & Raven, J.A.** 2009. Cryptic photosynthesis – extrasolar planetary oxygen without a surface biological signature. *Astrobiology* **9**, 623-636. <http://dx.doi.org/10.1089/ast.2008.0273> J
- Cockell, C.S., Léger, A., Fridlund, M., Herbst, T.M., Kaltenegger, L., Absil, O., Beichman, C., Benz, W., Blanc, M., Brack, A., Chelli, A., Colangeli, L., Cottin, H., du Foresto, F.C., Danchi, W.C., Defrère, D., den Herder, J.W., Eiroa, C., Greaves, J., Henning, T., Johnston, K.J., Jones, H., Labadie, L., Lammer, H., Launhardt, R., Lawson, P., Lay, O.P., LeDuigou, J.-M., Liseau, R., Malbet, F., Martin, S.R., Mawet, D., Mourard, D., Moutou, C., Mugnier, L.M., Olliver, M., Paresce, F., Quirrenbach, A., Rabbia, Y.D., Raven, J.A., Rottgering, H.J.A., Rouan, D., Santos, N.C., Selsis, F., Serabyn, E., Shibai, H., Tamura, M., Thiébaud, E., Westall, F. & White, G.J.** 2009. Darwin - a mission to detect and search for life on extrasolar planets. *Astrobiology* **9**, 1-22. <http://dx.doi.org/10.1089/ast.2007.0227> J
- Comadran, J.** 2008. Association mapping of *Hordeum vulgare* in Mediterranean environments. *University of Dundee*, PhD Thesis. O
- Comadran, J., Thomas, W.T.B., van Eeuwijk, F.A., Ceccarelli, S., Grando, S., Stanca, A.M., Pecchioni, N., Akar, T., Al-Yassin, A., Benbelkacem, A., Ouabbou, H., Bort, J., Romagosa, I., Hackett, C.A. & Russell, J.R.** 2009. Patterns of genetic diversity and linkage disequilibrium in a highly structured *Hordeum vulgare* association-mapping population for the Mediterranean basin. *Theoretical and Applied Genetics* **119**, 175-187. <http://dx.doi.org/10.1007/s00122-009-1027-0> J
- Cook, N., Karley, A.J., Russell, J.R. & Hubbard, S.F.** 2008. Population genetics of farmland sawflies. *Entomological Society of America Annual Meeting*, Reno, NV, USA, 16-19 November 2008. P
- Cooke, D.E.L., Lees, A.K., Hansen, J.G., Lassen, P., Andersson, B. & Bakonyi, J.** 2008. Eucablight pathogen database update. *11th Euroblight Workshop*, Hamar, Norway, 28-31 October 2008. P
- Cooke, D.E.L., Lees, A.K., Shaw, D.S., Taylor, M., Bain, R.A. & Ritchie, J.** 2008. Drivers and possible consequences of a changing population of *Phytophthora infestans* on the GB potato industry. *9th International Congress of Plant Pathology*, Turin, Italy, 24-29 August 2008. P
- Cooke, D.E.L., Lees, A.K., Zhan, J. & Birch, P.R.J.** 2008. Molecular tools for analysis of populations of *Phytophthora infestans* at a local and global scale. *Global Potato Conference 2008*, New Delhi, India, 9-12 December 2008. P
- Cornulier, T., Elston, D.A., Arcese, P., Benton, T.G., Douglas, D.J.T., Lambin, X., Reid, J., Robinson, R.A. & Sutherland, W.J.** 2009. Estimating the annual number of breeding attempts from breeding dates using mixture models. *Ecology Letters* **12**, 1184-1193. <http://dx.doi.org/10.1111/j.1461-0248.2009.01377.x> J
- Cousens, C., Thonur, L., Imlach, S., Crawford, J., Sales, J. & Griffiths, D.J.** 2009. Jaagsiekte sheep retrovirus is present at high concentration in lung fluid produced by ovine pulmonary adenocarcinoma-affected sheep and can survive for several weeks at ambient temperatures. *Research in Veterinary Science* **87**, 154-156. <http://dx.doi.org/10.1016/j.rvsc.2008.11.007> J
- Crampton, B.G., Hein, I. & Berger, D.K.** 2009. Salicylic acid confers resistance to a biotrophic rust pathogen, *Puccinia substriata*, in pearl millet (*Pennisetum glaucum*). *Molecular Plant Pathology* **10**, 291-304. <http://dx.doi.org/10.1111/j.1364-3703.2008.00532.x> J
- Crosley, L.K., Duthie, S.J., Polley, A.C., Bouwman, F.G., Heim, C., Mulholland, F., Horgan, G., Johnson, I.T., Mariman, E.C., Elliott, R.M., Daniel, H. & de Roos, B.** 2009. Variation in protein levels obtained from human blood cells and biofluids for platelet, peripheral blood mononuclear cell, plasma, urine and saliva proteomics. *Genes and Nutrition* **4**, 95-102. <http://dx.doi.org/10.1007/s12263-009-0121-x> J
- Cross, J., Berrie, A., Xu, X., Fitzgerald, J., O'Neill, T.M., Wedgood, E., Allen, J., Hall, D., Farman, D., Birch, A.N.E., Mitchell, C., Jorna, C.S., Shepherd, T., Boonham, N. &**





**Spence, N.J.** 2009. Free of pests, diseases – and residues. *HDC News* February 2009, 22-24. O

**Cullen, D.W., Squire, G.R., McNicol, J.W., Jacobs, J.H., Osborne, J.L., Ford, L., Ramsay, G., Scrimgeour, C.M. & Young, M.W.** 2008. Development and validation of gas chromatography and real-time quantitative PCR for the quantification of landscape-scale gene flow from varieties of high erucic acid (HEAR) oilseed rape. *Journal of the Science of Food and Agriculture* **88**, 2253-2264.  
<http://dx.doi.org/10.1002/jsfa.3340> J

**Cummings, S.P., Gyaneshwar, P., Vinuesa, P., Farruggia, F.T., Andrews, M., Humphry, D., Elliott, G.N., Nelson, A., Orr, C., Pettitt, D., Shah, G.R., Santos, S.R., Krishnan, H.B., Odee, D., Moreira, F.M.S., Sprent, J.I., Young, J.P.W. & James, E.K.** 2009. Nodulation of *Sesbania* species by *Rhizobium* (*Agrobacterium*) strain IRBG74 and other Rhizobia. *Environmental Microbiology* **11**, 2510-2525.  
<http://dx.doi.org/10.1111/j.1462-2920.2009.01975.x> J

**Daniell, T.J.** 2009. Book Review: Mycorrhizae in Crop Production. Edited by C. Hamek & C. Plenchette. Haworth Food and Agricultural Products Press, Binghampton, NY, USA (2007). *Experimental Agriculture* **45**, 239.  
<http://dx.doi.org/10.1017/S0014479708007369> O

**Davidson, R.S., Marion, G., White, P.C.L. & Hutchings, M.R.** 2009. Use of host population reduction to control wildlife infection: rabbits and paratuberculosis. *Epidemiology and Infection* **137**, 131-138.  
<http://dx.doi.org/10.1017/S0950268808000642> J

**Davies, H.V., Bryan, G.J. & Taylor, M.A.** 2008. Advances in functional genomics and genetic modification of potato (Review). *Potato Research* **51**, 283-299.  
<http://dx.doi.org/10.1007/s11540-008-9112-3> J

**Davis, J., Armengaud, P., Newton, A.C., White, P.J. & Amtmann, A.** 2009. Potassium deficiency and defence signaling. *Regulatory Oxylipins: An International Symposium*, Lausanne, Switzerland, 4-6 June 2009 (Poster). P

**Davis, J., Armengaud, P., Newton, A.C., White, P.J. & Amtmann, A.** 2009. Potassium deficiency and JA-dependent responses to biotic stress in barley. *University of Glasgow Post Graduate Symposium 2009*, Glasgow, UK, 9 June 2009 (Poster). P

**Dawson, I.K., Hedley, P.E., Guarino, L. & Jaenicke, H.** 2009. Does biotechnology have a role in the promotion of underutilised crops? *Food Policy* **34**, 319-328.  
<http://dx.doi.org/10.1016/j.foodpol.2009.02.003> J

**Deakin, S.J., Stewart, D., Ford, I., MacRury, S. & Megson, I.L.** 2009. Delphinidin, but not resveratrol protects vascular endothelial cells from hydroxyl radical-induced death. *2nd International Conference: Berries and Human Health, International Berry Health Benefits Symposium*, Monterey, California, USA, 21-24 June 2009 (Abstract). P

**Deakin, S.J., Stewart, D., Ford, I., MacRury, S. & Megson, I.L.** 2009. Nitric oxide sensitivity is preserved in porcine coronary artery pre-treated with delphinidin but not resveratrol. *2nd International Conference: Berries and Human Health, International Berry Health Benefits Symposium*, Monterey, California, USA, 21-24 June 2009 (Abstract). P

**Deakin, S.J., Stewart, D., Ford, I., MacRury, S. & Megson, I.L.** 2009. Nitric oxide sensitivity is preserved in porcine coronary artery pre-treated with delphinidin but not resveratrol,

*Society of Free Radical Research: Free Radicals, Health and Lifestyle*, Rome, Italy, 26-29 August 2009 (Abstract). *Free Radical Research* **43**, 1256-1257. P

**Debeljak, M., Kocev, D., Towers, W., Jones, M., Griffiths, B.S. & Hallett, P.D.** 2009. Potential of multi-objective models for risk-based mapping of the resilience characteristics of soils: demonstration at a national level. *Soil Use and Management* **25**, 66-77.  
<http://dx.doi.org/10.1111/j.1475-2743.2009.00196.x> J

**Debeljak, M., Squire, G.R., Leprince, F. & Brus, R.** 2008. Application of qualitative modeling in ecosystem-based environmental risk assessment of GMOs with respect to EU regulations. *10th International Symposium on Biosafety of Genetically Modified Organisms*, Wellington, New Zealand, 16-21 November 2008. P

**Dennis, P.G., Hirsch, P.R., Smith, S.J., Taylor, R.G., Valsami-Jones, E. & Miller, A.J.** 2009. Linking rhizoplane pH and bacterial density at the microhabitat scale. *Journal of Microbiological Methods* **76**, 101-104.  
<http://dx.doi.org/10.1016/j.mimet.2008.09.013> J

**Derridj, S., Elad, Y. & Birch, A.N.E.** 2009. Sugar signalling and a new way for vegetable and fruit induced resistance against insects, pathogens and nematodes. *IOBC/WPRS Working Group "Induced resistance in plants against insects and diseases"*, Granada, Spain, 12-16 May 2009. P

**Dimier, C., Brunet, C., Geider, R.D. & Raven, J.A.** 2009. Growth and photoregulation dynamics of the picoeukaryote *Pelagomonas calceolata* in fluctuating light. *Limnology and Oceanography* **54**, 823-836. J

**Dixon, E.R., Junquera, D., Martinez, A., Blackwell, M.S.A., Kemp, H., Meier-Augenstein, W., Duffy, A. & Bol, R.** 2009. Field scale spatial distribution of soil  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  under permanent grassland. *Simsug 2009*, University of Glasgow, Glasgow, 14-15 January 2009. P

**Dobson, G., Shepherd, T., Verrall, S.R., Conner, S., McNicol, J.W., Ramsay, G., Shepherd, L.V.T., Davies, H.V. & Stewart, D.** 2008. Phytochemical diversity in tubers of potato cultivars and landraces using a GC-MS metabolomics approach. *Journal of Agricultural and Food Chemistry* **56**, 10280-10291. <http://dx.doi.org/10.1021/jf801370b> J

**Donn, S., Griffiths, B.S., Neilson, R. & Daniell, T.J.** 2009. T-RFLP analysis of nematode assemblages. *British Society for Soil Science Spring Conference. Predicting the Future for Highly Organic Soils*, Edinburgh, UK, 5-7 May 2009. P

**Druka, A., Druka, I., Centeno, A.G., Li, H., Sun, Z., Thomas, W.T.B., Bonar, N., Steffenson, B.J., Ullrich, S.E., Kleinhofs, A., Wise, R.P., Close, T.J., Potokina, E., Luo, Z., Wagner, C., Schweizer, G.F., Marshall, D.F., Kearsley, M.J., Williams, R.W. & Waugh, R.** 2008. Towards systems genetic analyses in barley: Integration of phenotypic, expression and genotype data into GeneNetwork. *BMC Genetics* **9**, 73-73.  
<http://dx.doi.org/10.1186/1471-2156-9-73> J

**Du, Z., Chen, F., Zhao, Z., Liao, Q., Palukaitis, P. & Chen, J.** 2008. The 2b protein and the C-terminus of the 2a protein of *Cucumber mosaic virus* subgroup I strains both play a role in the viral RNA accumulation and induction of symptoms. *Virology* **380**, 363-370.  
<http://dx.doi.org/10.1016/j.virol.2008.07.036> J

**Ducreux, L., Morris, W.L., Prosser, I.M., Morris, J.A., Beale, M.H., Wright, F., Shepherd, T., Bryan, G.J.,**



- Hedley, P.E. & Taylor, M.A.** 2008. Expression profiling of potato germplasm differentiated in quality traits leads to the identification of candidate flavour and texture genes. *Journal of Experimental Botany* **59**, 4219-4231.  
<http://dx.doi.org/10.1093/jxb/ern264> J
- Duncan, S.H., Lobley, G.E., Holtrop, G., Ince, J., Johnstone, A.M., Louis, P. & Flint, H.J.** 2008. Human colonic microbiota associated with diet, obesity and weight loss. *International Journal of Obesity* **32**, 1720-1724.  
<http://dx.doi.org/10.1038/ijo.2008.155> J
- Dupuy, L.** 2009. Book Review: An Introduction to Mathematical Models in Ecology and Evolution. Time and space. 2nd edition by M. Gillman. Wiley-Blackwell, Chichester (2009). *Experimental Agriculture*, **45**, 509.  
<http://dx.doi.org/10.1017/S001447970999021> O
- Dupuy, L.** 2009. Imaging the 3D kinematics of circummutation in maize roots. *Society for Experimental Biology Annual Main Meeting*, Glasgow, UK, 28 June-1 July 2009. P
- Dupuy, L.** 2009. New approaches for the modelling of root architecture. *The 20th International Conference on Arabidopsis Research*, EICC, Edinburgh, UK, 30 June-4 July 2009. P
- Dupuy, L., Federici, F. & Haseloff, J.** 2009. Novel tools for plant tissue engineering. *BioSysBio Conference 2009*, University of Cambridge, Cambridge, UK, 23-25 March 2009 (Talk). P
- Dupuy, L., Vignes, M., McKenzie, B.M. & White, P.J.** 2009. The dynamics of plant root meristems. *French Annual Meeting of Biology, Informatics and Mathematics*, Nantes, France, 9-11 June 2009. P
- Dupuy, L., White, P.J., McKenzie, B.M. & Vignes, M.** 2009. Meristematic fronts: a new model of the plant architectural development, *Society for Experimental Biology Annual Main Meeting*, Glasgow, UK, 28 June -1 July 2009. *Comparative Biochemistry and Physiology* **153**, 219. P
- Eaton, S.L., Anderson, M.J., Hamilton, S., González, L., Sales, J., Jeffrey, M., Reid, H.W., Rocchi, M.A. & Chianini, F.** 2009. CD21 B cell populations are altered following subcutaneous scrapie inoculation in sheep. *Veterinary Immunology and Immunopathology* **131**, 105-109.  
<http://dx.doi.org/10.1016/j.vetimm.2009.02.012> J
- Fankem, H., Din, N., Nwaga, D., Daniell, T.J. & George, T.S.** 2009. Phosphate solubilization by rhizosphere fungi (*Aspergillus* and *Penicillium*) isolated from Cameroonian soils and their effect on growth and nutrient uptake of barley. *8th International Plant Growth-Promoting Rhizobacteria Workshop*, Portland, Oregon, USA, 17-22 May 2009. P
- Fankem, H., Din, N., Nwaga, D., Daniell, T.J. & George, T.S.** 2009. Phosphate solubilization by rhizosphere fungi (*Aspergillus* and *Penicillium*) isolated from Cameroonian soils and their effect on growth and nutrient uptake of barley. *15th Molecular Microbial Ecology Group Meeting*, University of Aberdeen, Aberdeen, UK, 29-30 July 2009. P
- Fankem, H., Din, N., Nwaga, D., Daniell, T.J. & George, T.S.** 2009. Preliminary selection of phosphate solubilising plant growth promoting microorganisms. *Society for Experimental Biology Annual Main Meeting*, Glasgow, UK, 28 June-1 July 2009. P
- Farmer, N., Curran, J., Lucy, D., Daeid, N.N. & Meier-Augenstein, W.** 2009. Stable isotope profiling of burnt wooden safety matches. *Science and Justice* **49**, 107-113.  
<http://dx.doi.org/10.1016/j.scijus.2009.03.007> J
- Federici, F., Dupuy, L. & Haseloff, J.** 2008. Novel tools for plant tissue engineering, *Synthetic Biology 4.0. The Fourth International Meeting on Synthetic Biology*, University of Science & Technology, Kowloon, Hong Kong, 10-12 October 2008 (Poster). P
- Fenton, B.** 2008. Insecticide resistant aphids. *SAC Information Note* **113**. O
- Fenton, B., Malloch, G., Flynn, A. & Van Toor, R.** 2009. Progress in field diagnostics for potato virus detection and epidemiology. *Association of Applied Biologists. Advances in Plant Virology*, Harrogate, UK, 1-3 April 2009. P
- Foito, A., Byrne, S., Shepherd, T., Stewart, D. & Barth, S.** 2009. Drought stress in perennial ryegrass - a metabolomics perspective, *Monogram Network Workshop 2009*, Bristol, UK, 29 April-1 May 2009. P
- Foito, A., Byrne, S., Shepherd, T., Stewart, D. & Barth, S.** 2009. Transcriptional and metabolic profiles of *Lolium perenne* L. genotypes in response to a PEG induced water stress. *Plant Biotechnology Journal* **7**, 719-732.  
<http://dx.doi.org/10.1111/j.1467-7652.2009.00437.x> J
- Foito, A., Byrne, S., Stewart, D. & Barth, S.** 2009. Transcriptional and metabolic profiles of *Lolium perenne* L. genotypes in response to a PEG induced water stress. *28th EUCARPIA Meeting of Fodder Crops and Grasses*, La Rochelle, France, 11-14 May 2009 (Abstract). P
- Foito, A., Byrne, S., Stewart, D. & Barth, S.** 2009. Transcriptional and metabolic profiles of *Lolium perenne* L. genotypes in response to a PEG induced water stress. *International Plant Abiotic Stress Conference*, Vienna, Austria, 8-11 February 2009 (Poster). P
- Fry, W.E., Grünwald, N.J., Cooke, D.E.L., McLeod, A., Forbes, G. A. & Cao, K.** 2009. Population genetics and population diversity of *Phytophthora infestans*. In: K.H. Lamour & S.Kamoun, eds. *Oomycete Genetics and Genomics: Diversity, Interactions and Research Tools*. Wiley-Blackwell, Hoboken, New Jersey, USA, 139-163. R
- García-Ruiz, R., Ochoa, V., Viñegla, B., Hinojosa, M.B., Peña-Santiago, R., Liébanas, G., Linares, J.C. & Carreira, J.A.** 2009. Soil enzymes, nematode community and selected physico-chemical properties as soil quality indicators in organic and conventional olive oil farming: influence of seasonality and site features. *Applied Soil Ecology* **41**, 305-314.  
<http://dx.doi.org/10.1016/j.apsoil.2008.12.004> J
- Garnett, M.H., Hartley, I.P., Hopkins, D.W., Sommerkorn, M. & Wookey, P.A.** 2009. A passive sampling method for radiocarbon analysis of soil respiration using molecular sieve. *Soil Biology and Biochemistry* **41**, 1450-1456.  
<http://dx.doi.org/10.1016/j.soilbio.2009.03.024> J
- George, T.S.** 2009. The genetic component of variation in phosphorus use by barley is strongly influenced by environment. *Royal Society of Edinburgh: Annual Research Awards Reception*, Edinburgh, UK, 2 September 2009. P
- George, T.S., Brown, L.K., Daniell, T.J., Gregory, P.J. & Richardson, A.E.** 2009. Microbial community structure in the rhizosphere of tobacco plants engineered to release fungal phytase from roots. *7th International Symposium of the Society for Root Research: Root Research and Applications*, Vienna, Austria, 2-4 September 2009. P



- George, T.S., Brown, L.K., Newton, A.C., Hallett, P.D., Thomas, W.T.B. & White, P.J.** 2009. Impact of soil tillage on the robustness of the genetic component of variation in phosphorus use efficiency in barley (*Hordeum vulgare* L.). *Abstracts and Programme of the XVI International Plant Nutrition Colloquium "Plant Nutrition for Sustainable Development and Global Health"*, Sacramento, California, USA, 26-30 August 2009. 175. P
- George, T.S., Gregory, P.J., Hocking, P. & Richardson, A.E.** 2008. Variation in root-associated phosphatase activities in wheat contributes to the utilisation of organic P substrates *in vitro*, but does not explain differences in the P-nutrition of plants when grown in different soils. *Environmental and Experimental Botany* **64**, 239-249.  
<http://dx.doi.org/10.1016/j.envexpbot.2008.05.002> J
- George, T.S., Wishart, J., Brown, L.K., Ramsay, G., Bradshaw, J.E., Gregory, P.J. & White, P.J.** 2009. Variation in drought tolerance in potatoes. *Potatoes in Practice*, Scottish Crop Research Institute, Dundee, UK, 13 August 2009. P
- Glasbey, C.A.** 2009. Two-dimensional generalisations of dynamic programming for image analysis. *Statistics and Computing* **19**, 49-56.  
<http://dx.doi.org/10.1007/s11222-008-9068-9> J
- Glasbey, C.A. & Khondoker, M.R.** 2009. Efficiency of functional regression estimators for combining multiple laser scans of cDNA microarrays. *Biometrical Journal* **51**, 45-55.  
<http://dx.doi.org/10.1002/bimj.200710444> J
- Gordon, D.C. & Hallett, P.D.** 2009. Rise in CO<sub>2</sub> affects soil water transport through repellency. *2nd International Conference Biohydrology 2009: A Changing Climate for Biology and Soil Hydrology*, Bratislava, Slovakia, 21-24 September 2009. *Biologia* **64**, 532-535.  
<http://dx.doi.org/10.2478/s11756-009-0115-6> P
- Graham, J.** 2009. Book Review: Principles and practices of plant genomics. Volume 1. Genome mapping. Edited by C. Kole and A.G. Abbott. Science Publishers, Enfield, NH, USA (2008). *Experimental Agriculture* **45**, 131.  
<http://dx.doi.org/10.1017/S0014479708007217> O
- Graham, J. & Jennings, S.N.** 2009. Raspberry breeding. In: S.M. Jain & M. Priyadarshan, eds. *Breeding Plantation Tree Crops: Temperate Species*, IBH & Science Publication Inc., Oxford, UK, 233-248.  
[http://dx.doi.org/10.1007/978-0-387-71203-1\\_7](http://dx.doi.org/10.1007/978-0-387-71203-1_7) R
- Graham, J., Hackett, C.A., Smith, K., Woodhead, M., Hein, I. & McCallum, S.** 2009. Mapping QTLs for developmental traits in raspberry from bud break to ripe fruit. *Theoretical and Applied Genetics* **118**, 1143-1155.  
<http://dx.doi.org/10.1007/s00122-009-0969-6> J
- Graham, J., Ratnaparkhe, M.B. & Powell, W.** 2009. Molecular mapping and breeding of physiological traits. In: C. Kole & A.G. Abbott, eds. *Principles and Practices of Plant Genomics. Volume 2. Molecular Breeding*, Science Publishers Inc., Enfield, New Hampshire, USA, 217-241. R
- Graham, J., Woodhead, M., Smith, K., Russell, J.R., Marshall, B., Ramsay, G. & Squire, G.R.** 2009. New insight into wild red raspberry populations using simple sequence repeat markers. *Journal of the American Society for Horticultural Science* **134**, 109-119. J
- Granger, S., Bol, R., Meier-Augenstein, W. & Kemp, H.** 2009. The hydrological response of heavy clay soils to rainfall as assessed using  $\delta^2\text{H}$ . *Simsug 2009*, University of Glasgow, Glasgow, UK, 14-15 January 2009. P
- Granum, E., Roberts, K., Raven, J.A. & Leegood, R.C.** 2009. Primary carbon and nitrogen metabolic gene expression in the diatom *Thalassiosira pseudonana* (Bacillariophyceae): Diel periodicity and effects of inorganic carbon and nitrogen. *Journal of Phycology* **45**, 1083-1092.  
<http://dx.doi.org/10.1111/j.1529-8817.2009.00728.x> J
- Gregory, A.S., Watts, C.W., Griffiths, B.S., Hallett, P.D., Kuan, H.L. & Whitmore, A.P.** 2009. The effect of long-term soil management on the physical and biological resilience of a range of arable and grassland soils in England. *Geoderma* **153**, 172-185. <http://dx.doi.org/10.1016/j.geoderma.2009.08.002> J
- Gregory, P.J.** 2009. Measuring root system architecture: opportunities and challenges. *7th International Symposium of the Society for Root Research: Root Research and Applications*, Vienna, Austria, 2-4 September 2009. P
- Gregory, P.J. & Ingram, J.S.I.** 2008. Climate change and the current 'food crisis'. *CAB Reviews: Perspectives in Agriculture, Veterinary Science, Nutrition and Natural Resources* **3**, 090, 1-10. J
- Gregory, P.J., Bengough, A.G., Grinev, D.V., Schmidt, S., Thomas, W.T.B., Wojciechowski, T. & Young, I.M.** 2009. Root phenomics of crops - opportunities and challenges. *Functional Plant Biology* **36**, 922-929. J
- Gregory, P.J., Johnson, S.N., Newton, A.C. & Ingram, J.S.I.** 2008. Climate change: the challenge for international agriculture. *Association of Applied Biologists. Effects of Climate Change on Plants: Implications for Agriculture*, Rothamsted Research, Harpenden, UK, 12-13 November 2008. P
- Gregory, P.J., Johnson, S.N., Newton, A.C. & Ingram, J.S.I.** 2009. Integrating pests and pathogens into the climate change/food security debate. *Journal of Experimental Botany* **60**, 2827-2838. <http://dx.doi.org/10.1093/jxb/erp080> J
- Grouffaud, S., van West, P., Avrova, A.O., Birch, P.R.J. & Whisson, S.C.** 2008. *Plasmodium falciparum* and *Hyaloperonospora parasitica* effector translocation motifs are functional in *Phytophthora infestans*. *Microbiology* **154**, 3743-3751. <http://dx.doi.org/10.1099/mic.0.2008/021964-0> J
- Guilioni, L., Jones, H.G., Leinonen, I. & Lhomme, J.P.** 2008. On the relationships between stomatal resistance and leaf temperatures in thermography. *Agricultural and Forest Meteorology* **148**, 1908-1912.  
<http://dx.doi.org/10.1016/j.agrformet.2008.07.009> J
- Haas, B.J., Kamoun, S., Zody, M.C., Jiang, R.H.Y., Handsaker, R.E., Cano, L.M., Grabherr, M., Kodira, C.D., Raffaele, S., Torto-Alalibo, T., Bozkurt, T.O., Ah-Fong, A.M.V., Alvarado, L., Anderson, V.L., Armstrong, M.R., Avrova, A.O., Baxter, L., Beynon, J., Boevink, P.C., Bollman, S.R., Bos, J.I.B., Bulone, V., Cai, G.H., Cakir, C., Carrington, J.C., Chawner, M., Conti, L., Costanzo, S., Ewan, R., Fahlgren, N., Fischbach, M.A., Fugelstad, J., Gilroy, E.M., Gnerre, S., Green, P.J., Grenville-Briggs, L.J., Griffith, J., Grunwald, N.J., Horn, K., Horner, N.R., Hu, C.H., Huitema, E., Jeong, D.H., Jones, A.M.E., Jones, J.D.G., Jones, R.W., Karlsson, E.K., Kunjeti, S.G., Lamour, K., Liu, Z.Y., Ma, L.J., MacLean, D., Chibucos, M.C., McDonald, H., McWalters, J., Meijer, H.J.G., Morgan, W., Morris, P.F., Munro, C.A., O'Neill, K., Ospina-Giraldo, M., Pinzón, A., Pritchard, L., Ramsahoye,**

- B., Ren, Q.H., Restrepo, S., Roy, S., Sadanandom, A., Savidor, A., Schornack, S., Schwartz, D.C., Schumann, U.D., Schwessinger, B., Seyer, L., Sharpe, T., Silvar, C., Song, J., Studholme, D.J., Sykes, S., Thines, M., van de Vondervoort, P.J.I., Phuntumart, V., Wawra, S., Weide, R., Win, J., Young, C., Zhou, S.G., Fry, W., Meyers, B.C., van West, P., Ristaino, J., Govers, F., Birch, P.R.J., Whisson, S.C., Judelson, H.S. & Nusbaum, C. 2009. Genome sequence and analysis of the Irish potato famine pathogen *Phytophthora infestans*. *Nature* **461**, 393-398. <http://dx.doi.org/10.1038/nature08358> J
- Hall, S., English, L.C. & Hopkins, D.W. 2009. Progress towards understanding microbial activity in a raised mire. *British Society of Soil Science: Predicting the Future for Highly Organic Soils*, Edinburgh, UK, 5-7 May 2009. P
- Hallett, P.D., Feeney, D., Bengough, A.G., Rillig, M.C., Scrimgeour, C.M. & Young, I.M. 2009. Disentangling the impact of AM fungi versus roots on soil structure and water transport. *Plant and Soil* **314**, 183-196. <http://dx.doi.org/10.1007/s11104-008-9717-y> J
- Hallett, P.D., Lichner, L., Rajkai, K., Schaumann, G.E., Škvarenina, J. & Tesar, M. 2009. Foreword to the thematic issue on Biohydrology. *2nd International Conference Biohydrology 2009: A Changing Climate for Biology and Soil Hydrology*, Bratislava, Slovakia, 21-24 September 2009. *Biologia* **64**, 415-418. <http://dx.doi.org/10.2478/s11756-009-0116-5> P
- Hallett, P.D., Loades, K., Mickovski, S.B., Bengough, A.G., Bransby, M.F., Davies, M.C.R. & Sonnenberg, R. 2009. An assessment of models that predict soil reinforcement by plant roots. *European Geosciences Union*, Vienna, Austria, 18-22 April 2009. P
- Hallett, P.D., Sun, B., Dennis, P.G. & Hopkins, D.W. 2009. Gelifluction of soil on the Antarctic Peninsula. *European Geosciences Union*, Vienna, Austria, 18-22 April 2009. P
- Hammond, J.P. & White, P.J. 2008. Assessing the potential for breeding Brassica crops with improved P use efficiency. *Oilseed Rape Genetic Improvement Network (OREGIN) 6th Stakeholder Forum Meeting on "Assessing low input systems for breeding and trialling - the N and P economy"*, NIAB, Cambridge, UK, 21 November 2008 (Talk). P
- Hammond, J.P., Broadley, M.R., Bowen, H.C., Hayden, R., Spracklen, W.P. & White, P.J. 2009. A molecular diagnostic for phosphorus deficiency in potatoes, *Abstracts and Programme of the XVI International Plant Nutrition Colloquium "Plant Nutrition for Sustainable Development and Global Health"*, Sacramento, California, USA, 26-30 August 2009. 93. P
- Hammond, J.P., Broadley, M.R., Bowen, H.C., Hayden, R., Spracklen, W.P. & White, P.J. 2009. Chips and Ps: A diagnostic array for phosphorus (P) deficiency in potatoes. *Phoenix 2009 Symposium: Protein Complexes in Signalling and Development*, Glasgow, UK, 25-27 June 2009. P
- Hammond, J.P., Broadley, M.R., White, P.J., King, G.J., Bowen, H.C., Hayden, R., Meacham, M.C., Mead, A., Overs, T., Spracklen, W.P. & Greenwood, D.J. 2009. Shoot yield drives phosphorus use efficiency in *Brassica oleracea* and correlates with root architecture traits. *Journal of Experimental Botany* **60**, 1953-1965. <http://dx.doi.org/10.1093/jxb/erp083> J
- Hammond, J.P., Broadley, M.R., White, P.J., King, G.J., Bowen, H.C., Spracklen, W.P., Hayden, R., Meacham, M. & Overs, T. 2008. Phosphorus use efficiency in *Brassica*. *Association of Applied Biologists, Resource Capture by Crops: Integrated Approaches*, Sutton Bonington, UK, 10-12 September 2008 (Talk). P
- Hammond, J.P., White, P.J. & Broadley, M.R. 2009. Getting the most out of soil nutrients. *Horticulture Week* 27 February 2009, 30-31. O
- Hansen, J.G., Andersson, B.A., Bain, R., Schmiedl, J., Soellinger, J., Ritchie, F., Bucena, L., Çakir, E., Cooke, L., Dubois, L., Filippov, A., Hannukkala, A., Hausladen, H., Hansvater, E., Heldak, J., Hermansen, A., Kapsa, J., Pliakhnevich, M., Koppel, M., Lees, A.K., Musa, T., Ronis, A., Schepers, H., Vogelaar, K. & Vanhaverbeke, P. 2008. The development and control of late blight (*Phytophthora infestans*) in Europe in 2007 and 2008. *11th Euroblight Workshop*, Hamar, Norway, 28-31 October 2008. P
- Hargreaves, C.E., Gregory, P.J. & Bengough, A.G. 2009. Measuring root traits in barley (*Hordeum vulgare* ssp. *vulgare* and ssp. *spontaneum*) seedlings using gel chambers, soil sacs and x-ray microtomography. *Plant and Soil* **316**, 285-297. <http://dx.doi.org/10.1007/s11104-008-9780-4> J
- Harrison, B.D. 2009. Plant virology in the 20th century. *Association of Applied Biologists. Advances in Plant Virology*, Harrogate, UK, 1-3 April 2009. P
- Hartley, I.P., Garnett, M.H., Hopkins, D.W., Sommerkorn, M. & Wookey, P.A. 2009. Plant-soil interactions, positive priming effects and patterns of soil C storage in Arctic Sweden. *International Symposium on Soil Organic Matter Dynamics: Land Use, Management and Global Change*, Colorado State University, Colorado, USA, 6-10 July 2009. P
- Hartley, I.P., Hopkins, D.W., Garnett, M.H., Sommerkorn, M. & Wookey, P.A. 2009. No evidence for compensatory thermal adaptation of soil microbial respiration in the study of Bradford *et al.* (2008). *Ecology Letters* **12**, E12-E14. <http://dx.doi.org/10.1111/j.1461-0248.2009.01300.x> J
- Haupt, S., Ziegler, A., Cowan, G.H. & Torrance, L. 2009. Studies of the role and function of *Barley stripe mosaic virus* encoded proteins in replication and movement using GFP fusions. In: B.W. Hicks, ed. *Viral Applications of Green Fluorescent Protein. Methods in Molecular Biology Volume* 515, Springer, Berlin, Germany, 287-297. <http://dx.doi.org/10.1007/978-1-59745-559-6> R
- Hawes, C. 2009. Book Review: Essentials of ecology. 3rd edition. By C.R. Townsend, M. Begon and J.L. Harper. Blackwell Publishing, Oxford (2008). *Experimental Agriculture* **45**, 128. <http://dx.doi.org/10.1017/S001447970800714X> O
- Hawes, C., Houghton, A.J., Bohan, D.A. & Squire, G.R. 2009. Functional approaches for assessing plant and invertebrate abundance patterns in arable systems. *Basic and Applied Ecology* **10**, 34-42. <http://dx.doi.org/10.1016/j.baae.2007.11.007> J
- Hein, I., Gilroy, E.M., Armstrong, M.R. & Birch, P.R.J. 2009. The zig-zag-zig in oomycete-plant interactions. *Molecular Plant Pathology* **10**, 547-562. <http://dx.doi.org/10.1111/J.1364-3703.2009.00547.X> J
- Hillier, J., Hawes, C., Squire, G.R., Hilton, A., Wale, S. & Smith, P. 2009. The carbon footprints of food crop production. *International Journal of Agricultural Sustainability* **7**, 107-118. <http://dx.doi.org/10.3763/ijas.2009.0419> J
- Hinsinger, P., Bengough, A.G., Vetterlein, D. & Young, I.M. 2009. Rhizosphere: biophysics, biogeochemistry and





ecological relevance. *Plant and Soil* **321**, 117-152.  
<http://dx.doi.org/10.1007/s11104-008-9885-9> J

**Holden, N., Pritchard, L. & Toth, I.K.** 2008. Colonisation outwith the colon: plants as an alternative environmental reservoir for human pathogenic enterobacteria. *FEMS Microbiology Reviews* **33**, 689-703.  
<http://dx.doi.org/10.1111/j.1574-6976.2008.00153.x> J

**Hopkins, D.W.** 2009. Soil organic carbon stocks and dynamics in long-term experimental grassland plots. *British Society for Soil Science Spring Conference. Predicting the Future for Highly Organic Soils*, Edinburgh, UK, 5-7 May 2009. P

**Hopkins, D.W., Dennis, P.G., Hartley, I.P. & Wookey, P.A.** 2009. Soil microbes and organic matter dynamics at the ends of the earth. *Soil Organic Matters International Conference*, Rothamsted Research, Harpenden, UK, 23-25 June 2009. P

**Hopkins, D.W., Sparrow, A.D., Gregorich, E.G., Elberling, B., Novis, P., Fraser, F., Scrimgeour, C.M., Dennis, P.G., Meier-Augenstein, W. & Greenfield, L.G.** 2009. Isotopic evidence for the provenance and turnover of organic carbon by soil microorganisms in the Antarctic dry valleys. *Environmental Microbiology* **11**, 597-608.  
<http://dx.doi.org/10.1111/j.1462-2920.2008.01830.x> J

**Hopkins, D.W., Waite, I.S., McNicol, J.W. & O'Donnell, A.G.** 2009. Soil organic carbon stocks and dynamics in long-term experimental grassland plots. *International Symposium on Soil Organic Matter Dynamics: Land Use, Management and Global Change*, Colorado State University, Colorado, USA, 6-10 July 2009. P

**Hopkins, D.W., Waite, I.S., McNicol, J.W., Poulton, P.R., MacDonald, A.J. & O'Donnell, A.G.** 2009. Soil organic carbon contents in long-term experimental grassland plots in the UK (Palace Leas and Park Grass) have not changed consistently in recent decades. *Global Change Biology* **15**, 1739-1754.  
<http://dx.doi.org/10.1111/j.1365-2486.2008.01809.x> J

**Hopkins, D.W., Waite, I.S. & O'Donnell, A.G.** 2009. Soil organic carbon stocks and dynamics in long-term experimental grassland plots. *2009 International Symposium on Environmental Science & Technology*, Shanghai, China, 2-5 June 2009. *Progress in Environmental Science and Technology* **2**, 1801-1807. P

**Hornyik, C., Terzi, L.C., Rataj, K., Marshall, J. & Simpson, G.G.** 2009. FPA controls pre-mRNA 3' end site selection. *20th International Conference on Arabidopsis Research*, Edinburgh, UK, 30 June-4 July 2009 (Poster). P

**Hudacsek, P., Bransby, M.F., Hallett, P.D. & Bengough, A.G.** 2009. Centrifuge modelling of climatic effects on clay embankments. *Proceedings of the Institution of Civil Engineers – Engineering Sustainability* **162**, 91-100.  
<http://dx.doi.org/10.1680/ensu.2009.162.2.91> J

**Hughes, V., Bannantine, J.P., Denham, S., Smith, S., Garcia-Sanchez, A., Sales, J., Paustian, M.L., McLean, K. & Stevenson, K.** 2008. Immunogenicity of proteome-determined *Mycobacterium avium* subsp *paratuberculosis*-specific proteins in sheep with Paratuberculosis. *Clinical and Vaccine Immunology* **15**, 1824-1833.  
<http://dx.doi.org/10.1128/CVI.00099-08> J

**Iannetta, P.P.M. & Squire, G.R.** 2008. Scottish Government funded weed research. *Proceedings of AAB Conference*:

*The future of weed research in the UK*, Home Grown Cereals Authority HQ, London, UK, 19 November 2008 (Invited Talk). P

**Iglesias-Rodriguez, M.D., Buitenhuis, E.T., Raven, J.A., Schofield, O., Poulton, A.J., Gibbs, S., Halloran, P.R. & de Baar, H.J.W.** 2008. Response to comment on 'Phytoplankton calcification in a high CO<sub>2</sub> world'. *Science* **322**, 1466.  
<http://dx.doi.org/10.1126/science.1161501> J

**Johnson, S.N. & Barton, A.** 2009. Climate change in the underworld: effects of elevated CO<sub>2</sub> on root-feeding insects. *Ento'09 National Meeting of the Royal Entomological Society*, University of Sheffield, Sheffield, UK, 15-17 July 2009. P

**Johnson, S.N. & Murray, P.J.** 2008. *Root Feeders: An Ecosystem Perspective*, CABI, Wallingford, UK. 272pp. R

**Johnson, S.N., Bezemer, T.M. & Jones, T.H.** 2008. Linking aboveground and belowground herbivory. In: S.N. Johnson & P.J. Murray, eds. *Root Feeders: An Ecosystem Perspective*, CABI, Wallingford, UK, 153-170. R

**Johnson, S.N., Hawes, C. & Karley, A.J.** 2009. Reappraising the role of plant nutrients as mediators of interactions between root- and foliar-feeding insects. *Functional Ecology* **23**, 699-706. <http://dx.doi.org/10.1111/j.1365-2435.2009.01550.x> J

**Jones, H.G. & Brennan, R.M.** 2009. Potential impacts of climate change on soft fruit production: the example of winter chill in *Ribes*. *COST 863 Euroberry Workshop on Berry Production in Changing Climate Conditions and Cultivation Systems*, Geisenheim, Germany, 29-31 October 2008. *Acta Horticulturae* **838**, 27-33. P

**Jones, H.G., Serraj, R., Loveys, B.R., Xiong, L.H., Wheaton, A. & Price, A.H.** 2009. Thermal infrared imaging of crop canopies for the remote diagnosis and quantification of plant responses to water stress in the field. *Functional Plant Biology* **36**, 978-989. <http://dx.doi.org/10.1071/FP09123> J

**Kalyna, M., Maranova, M., Simpson, C.G., Brown, J.W.S. & Barta, A.** 2009. Arabidopsis SR protein atRSp31: identification of RNA targets and its role in stress response. *Plant Biology 2009*, Hawaii, USA, 18-22 July 2009 (Poster). P

**Kalyna, M., Maranova, M., Simpson, C.G., Brown, J.W.S. & Barta, A.** 2009. Arabidopsis SR protein atRSp31: identification of RNA targets and its role in stress response. *RNA 2009*, Madison, Wisconsin, USA, 26-31 May 2009 (Poster). P

**Karjalainen, R.O., Stewart, D., McDougall, G.J., Hilz, H., Anttonen, M., Saviranta, N., Mattila, P. & Törrönen, R.** 2008. Understanding health-promoting bioactive compounds in blackcurrants and their agronomic improvement. *COST 863 Euroberry. Bioactive Compounds in Berry Fruits: Genetic Control, Breeding, Cultivar, Analytical Aspects and Human Health*, Zurich, Switzerland, 3-6 December 2008. P

**Karley, A.J. & White, P.J.** 2009. Moving cationic minerals to edible tissues: potassium, magnesium, calcium (Review). *Current Opinion in Plant Biology* **12**, 291-298.  
<http://dx.doi.org/10.1016/j.pbi.2009.04.013> J

**Kassim, A., Poette, J., Paterson, A., Zait, D., McCallum, S., Woodhead, M., Smith, K., Hackett, C.A. & Graham, J.** 2009. Environmental and seasonal influences on red raspberry anthocyanin antioxidant contents and identification of quantitative traits loci (QTL). *Molecular Nutrition and Food Research* **53**, 625-634.  
<http://dx.doi.org/10.1002/mnfr.200800174> J



- Kärenlampi, S. & White, P. J.** 2009. Potato proteins, lipids and minerals. In: J. Singh & L. Kaur, eds. *Advances in Potato Chemistry and Technology*, Academic Press, Burlington, MA, USA, 99-126. R
- Kerchev, P., Foyer, C.H., Fenton, B. & Hancock, R.D.** 2009. Investigating roles for reactive oxygen in plant-aphid interactions. *Plant ROS 2009: Society for Free Radical Research International Plant Oxygen Group Meeting on Reactive Oxygen and Nitrogen Species*, Helsinki, Finland, 8-10 July 2009 (Poster). P
- Kerchev, P., Foyer, C.H., Fenton, B. & Hancock, R.D.** 2009. Reactive oxygen and antioxidants modulate the interaction between *Myzus persicae* (Sulzer) and plant hosts. *8th International Symposium on Aphids*, Catania, Italy, 8-12 June 2009 (Poster). P
- Kiær, L.P., Felber, F., Flavell, A.J., Guadagnuolo, R., Guiatti, D., Hauser, T.P., Olivieri, A.M., Scotti, I., Syed, N.H., Vischi, M., van de Wiel, C. & Jørgensen, R.B.** 2009. Spontaneous gene flow and population structure in wild and cultivated chicory, *Cichorium intybus* L. *Genetic Resources and Crop Evolution* **56**, 405-419.  
<http://dx.doi.org/10.1007/s10722-008-9375-1> J
- Kikuchi, T., Li, H., Karim, N., Kennedy, M.W., Moens, M. & Jones, J.T.** 2009. Identification of putative expansin-like genes from the pine wood nematode, *Bursaphelenchus xylophilus*, and evolution of the expansin gene family within the Nematoda. *Nematology* **11**, 355-364.  
<http://dx.doi.org/10.1163/156854109X446953> J
- Kim, S.-H., Koroleva, O.A., Lewandowska, D., Pendle, A.F., Clark, G.P., Simpson, C.G., Shaw, P.J. & Brown, J.W.S.** 2009. Aberrant mRNA transcripts and the nonsense-mediated decay proteins UPF2 and UPF3 are enriched in the *Arabidopsis* nucleolus. *Plant Cell* **21**, 2045-2057.  
<http://dx.doi.org/10.1105/tpc.109.067736> J
- Kim, S.-H., Lewandowska, D., Clark, G., Pendle, A., Koroleva, O., Shaw, P.J. & Brown, J.W.S.** 2009. Aberrant mRNAs in the plant nucleolus. *RNA 2009*, Madison, Wisconsin, USA, 26-31 May 2009 (Poster). P
- Kim, S.-H., Lewandowska, D., Clark, G., Simpson, C.G., Pendle, A., Koroleva, O., Shaw, P.J. & Brown, J.W.S.** 2009. Aberrant mRNAs in the plant nucleolus. *20th International Conference on Arabidopsis Research*, Edinburgh, UK, 30 June-4 July 2009 (Poster). P
- Kopecký, D., Bartoš, J., Lukaszewski, A.J., Baird, J.H., Cernoch, V., Kölliker, R., Rognli, O.A., Blois, H., Caig, V., Lubberstedt, T., Studer, B., Shaw, P., Doležel, J. & Kilian, A.** 2009. Development and mapping of DArT markers within the *Festuca - Lolium* complex. *BMC Genomics* **10**, 473.  
<http://dx.doi.org/10.1186/1471-2164-10-473> J
- Koroleva, O.A., Calder, G., Pendle, A.F., Kim, S.-H., Lewandowska, D., Simpson, C.G., Jones, I.M., Brown, J.W.S. & Shaw, P.J.** 2009. Dynamic behaviour of *Arabidopsis* eIF4A-III, putative core protein of exon junction complex: fast relocation to nucleolus and splicing speckles under hypoxia. *Plant Cell* **21**, 1592-1606.  
<http://dx.doi.org/10.1105/tpc.108.060434> J
- Koza, A., Hallett, P.D., Moon, C.D. & Spiers, A.J.** 2009. Characterisation of a novel air-liquid interface biofilm of *Pseudomonas fluorescens* SBW25. *Microbiology* **155**, 1397-1406. <http://dx.doi.org/10.1099/mic.0.025064-0> J
- Laimer, M., Marzban, G., Herndl, A., Beekwilder, J., McDougall, G.J., Stewart, D., Quiles, J.L., Krüger, E., Atkinson, C., Nestby, R., Toldam-Anderson, T.B., Harsan, E., Heinonen, M., Olsson, M., Juranic, Z., Battino, M. & Mezzetti, B.** 2009. Euroberry research: from genomics to sustainable production, quality and health WG4: Bioactive compounds of berry fruit affecting human health. *COST 863 Management Workshop on Defining Needs of Berry Industries*, Lisbon, Portugal, 19-21 March 2009. P
- Lawson, D.J. & Jensen, H.J.** 2009. The role of weak selection and high mutation rates in nearly neutral evolution. *Journal of Theoretical Biology* **257**, 696-703.  
<http://dx.doi.org/10.1016/j.jtbi.2008.12.029> J
- Lees, A.K.** 2008. Powdery scab strains. *SAC Information Note* **108**. O
- Lees, A.K., Cooke, D.E.L., Stewart, J.A., Sullivan, L. & Carnegie, S.** 2008. *Phytophthora infestans* population changes: implications. *11th Euroblight Workshop*, Hamar, Norway, 28-31 October 2008. P
- Lees, A.K., Sullivan, L. & Cullen, D.W.** 2009. A quantitative polymerase chain reaction assay for the detection of *Polyscytalum pustulans*, the cause of skin spot disease of potato. *Journal of Phytopathology* **157**, 154-158.  
<http://dx.doi.org/10.1111/j.1439-0434.2008.01459.x> J
- Lees, A.K., Sullivan, L., Williams, N.A. & Cooke, D.E.L.** 2008. Using molecular tools to investigate the role of tuber and soil-borne inoculum of *Phytophthora infestans*. *9th International Congress of Plant Pathology*, Turin, Italy, 24-29 August 2008. P
- Lehrach, W.P. & Husmeier, D.** 2009. Segmenting bacterial and viral DNA sequence alignments with a trans-dimensional phylogenetic factorial hidden Markov model. *Journal of the Royal Statistical Society Series C - Applied Statistics* **58**, 307-327. <http://dx.doi.org/10.1111/j.1467-9876.2008.00648.x> J
- Lewis, F.I., Gunn, G.J., McKendrick, I.J. & Murray, F.M.** 2009. Bayesian inference for within-herd prevalence of *Leptospira interrogans* serovar Hardjo using bulk milk antibody testing. *Biostatistics* **10**, 719-728.  
<http://dx.doi.org/10.1093/biostatistics/kxp026> J
- Lewsey, M., Palukaitis, P. & Carr, J. P.** 2008. Plant-virus interactions: Defence and counter-defence. In: J. Parker, ed. *Annual Plant Reviews, Volume 34, Molecular Aspects of Plant Disease Resistance*, Wiley-Blackwell, Oxford, UK, 134-176. R
- Lewsey, M., Surette, M., Robertson, F.C., Ziebell, H., Choi, S.H., Ryu, K.H., Canto, T., Palukaitis, P., Payne, T., Walsh, J.A. & Carr, J.P.** 2009. The role of the *Cucumber mosaic virus* 2b protein in viral movement and symptom induction. *Molecular Plant-Microbe Interactions* **22**, 642-654.  
<http://dx.doi.org/10.1094/MPMI-22-6-0642> J
- Liu, H., Coulthurst, S.J., Pritchard, L., Hedley, P.E., Ravensdale, M., Humphris, S.N., Burr, T., Takle, G.W., Brurberg, M.B., Birch, P.R.J., Salmond, G.P.C. & Toth, I.K.** 2008. Quorum sensing coordinates brute force and stealth modes of infection in the plant pathogen *Pectobacterium atrosepticum*. *PLoS Pathogens* **4**, e1000093.  
<http://dx.doi.org/10.1371/journal.ppat.1000093> J
- Lochlainn, S., Bowen, H.C., Fray, R.G., Hammond, J.P., King, G.J., White, P.J. & Broadley, M.R.** 2008. Natural genetic variation in zinc (Zn) accumulation by Brassicaceae. *5th ISHS International Symposium on Brassicas and 16th Crucifer*



Genetics Workshop, Lillehammer, Norway, 8-12 September 2008, 131 (Poster). P

**Lokossou, A.A., Park, T.H., van Arkel, G., Arens, M., Ruyter-Spira, C., Morales, J., Whisson, S.C., Birch, P.R.J., Visser, R.G.F., Jacobsen, E. & van der Vossen, E.A.G.** 2009. Exploiting knowledge of *R/Avr* genes to rapidly clone a new LZ-NBS-LRR family of late blight resistance genes from potato linkage group IV. *Molecular Plant-Microbe Interactions* **22**, 630-641. <http://dx.doi.org/10.1094/MPMI-22-6-0630> J

**Longhurst, R., Roberts, D. & Daniell, T.J.** 2009. EnPrint secures start-up funding. *Young Company Finance Newsletter*. O

**Low, J.C., Chambers, J., McKelvey, W.A.C., McKendrick, I.J. & Jeffrey, M.** 2009. Failure to transmit scrapie infection by transferring preimplantation embryos from naturally infected donor sheep. *Theriogenology* **72**, 809-816. <http://dx.doi.org/10.1016/j.theriogenology.2009.05.017> J

**Ma'shum, M., Tisdall, J.M., Borrell, A.K., McKenzie, B.M., Gill, J.S., Kusnarta, I.G.M., Sukartono, M. & Van Cooten, D.E.** 2009. Rice responses to soil management in rice-based cropping system in the semi-arid tropics of southern Lombok, Eastern Indonesia. *Field Crops Research* **110**, 197-206. <http://dx.doi.org/10.1016/j.fcr.2008.08.003> J

**Maberly, S.C., Ball, L.A., Raven, J.A. & Sültemeyer, D.** 2009. Inorganic carbon acquisition by chrysophytes. *Journal of Phycology* **45**, 1052-1061. <http://dx.doi.org/10.1111/j.1529-8817.2009.00734.x> J

**MacFarlane, S.A. & McGavin, W.J.** 2009. Genome activation by *Raspberry bushy dwarf virus* coat protein. *Journal of General Virology* **90**, 747-753. <http://dx.doi.org/10.1099/vir.0.007195-0> J

**MacFarlane, S.A. & McGavin, W.J.** 2009. Sequencing studies for the identification and characterization of new and old *Rubus* viruses. *21st International Conference on Virus and other Graft Transmissible Diseases of Fruit Crops*, Neustadt, Germany, 5-10 July 2009 (Talk). P

**MacFarlane, S. A. & Neilson, R.** 2009. Testing of transmission of Tobraviruses by nematodes. In: R. Coico, T. Kowalik, J.M. Quarles, B. Stevenson & R.K. Taylor, eds. *Current Protocols in Microbiology*, John Wiley & Sons, Inc., Chichester, UK, 16B.5.1-16B.5.16. <http://dx.doi.org/10.1002/9780471729259.mc16b05s12> R

**Madsen, C.T., Stephens, J., Hornyik, C., Shaw, J., Collinge, D.B., Lacomme, C. & Albrechtsen, M.** 2009. Identification and characterization of barley RNA-directed RNA polymerases. *Biochimica et Biophysica Acta* **1789**, 375-385. <http://dx.doi.org/10.1016/j.bbagr.2009.03.003> J

**Maestre, F.T., Bowker, M., Puche, M.D., Bowker, M., Hinojosa, M.B., Martínez, I., García-Palacios, P., Castillo, A.P., Soliveres, S., Luzuriaga, A.L., Sánchez, A.M., Carreira, J.A., Gallardo, A. & Escudero, A.** 2009. Shrub encroachment can reverse desertification in semi-arid Mediterranean grasslands. *Ecology Letters* **12**, 930-941. <http://dx.doi.org/10.1111/j.1461-0248.2009.01352.x> J

**Mahajan, A., Currie, C.G., Mackie, S., Tree, J., McAteer, S., McKendrick, I., McNeilly, T.N., Roe, A., La Ragione, R.M., Woodward, M.J., Gally, D.L. & Smith, D.G.E.** 2009. An investigation of the expression and adhesin function of H7 flagella in the interaction of *Escherichia coli* O157:H7 with bovine intestinal epithelium. *Cellular Microbiology* **11**, 121-137. <http://dx.doi.org/10.1111/j.1462-5822.2008.01244.x> J

**Mahmood-ul-Hassan, M. & Gregory, P.J.** 2008. The role of rainfall intensity and soil in determining rates of flow through cryoturbated chalk. *Pakistan Journal of Science and Industrial Research* **51**, 235-241. J

**Mankin, R.W., Johnson, S.N., Grinev, D.V. & Gregory, P.J.** 2008. New experimental techniques for studying root herbivores. In: S.N. Johnson & P.J. Murray, eds. *Root Feeders: An Ecosystem Perspective*, CABI, Wallingford, UK, 20-32. R

**Maranova, M., Kalyna, M., Simpson, C.G., Brown, J.W.S. & Barta, A.** 2008. Identifying RNA targets of the *Arabidopsis thaliana* splicing factor atRSP31. *RNA 2008. 13th Annual Meeting of the RNA Society*, Berlin, Germany, 28 July-3 August 2008. P

**Maranova, M., Kalyna, M., Simpson, C.G., Brown, J.W.S. & Barta, A.** 2009. *Arabidopsis* SR protein AtRSP31: Identification of RNA targets and its role in plant development. *4th Annual EURASNET Meeting*, Assisi, Italy, 23-24 April 2009 (Poster). P

**Margaritopoulos, J.T., Kasprovicz, L., Malloch, G.L. & Fenton, B.** 2009. Tracking the global dispersal of a cosmopolitan insect pest, the peach potato aphid. *BMC Ecology* **9**, 13. <http://dx.doi.org/10.1186/1472-6785-9-13> J

**Marion, G., Smith, L.A., Swain, D.L., Davidson, R.S. & Hutchings, M.R.** 2008. Agent-based modelling of foraging behaviour: the impact of spatial heterogeneity on disease risks from faeces in grazing systems. *Journal of Agricultural Science* **146**, 507-520. <http://dx.doi.org/10.1017/S0021859608008022> J

**Marshall, B., Newton, A.C. & Zhan, J.** 2009. Quantitative evolution of aggressiveness of powdery mildew under two-cultivar barley mixtures. *Plant Pathology* **58**, 378-388. <http://dx.doi.org/10.1111/j.1365-3059.2008.01953.x> J

**Martin, P. & Johnson, S.E.** 2009. Will climate change accelerate the breakdown of aphid-resistance in raspberry? *Ento'09 National Meeting of the Royal Entomological Society*, University of Sheffield, Sheffield, UK, 15-17 July 2009 (Poster). P

**Martinussen, I., Stewart, D. & McDougall, G.J.** 2009. Metabolomic approach to identifying bioactive compounds in berries: advances toward fruit nutritional enhancement. *2nd International Conference: Berries and Human Health. International Berry Health Benefits Symposium*, Monterey, California, USA, 21-24 June 2009 (Abstract). P

**McDougall, G.J.** 2009. Book Review: Health Benefits of Organic Food: Effects of the Environment. Edited by I. Givens, S. Baxter, A.M. Minihane & E. Shaw. CABI, Wallingford (2008). *Experimental Agriculture* **45**, 375. <http://dx.doi.org/10.1017/S0014479709007650> O

**McDougall, G.J. & Stewart, D.** 2009. Bioactive components from berries: human health effects. *Euro Food Chem XV. Food for the Future*, Copenhagen, Denmark, 5-8 July 2009 (Talk). P

**McDougall, G.J., Kulkarni, N.N. & Stewart, D.** 2008. Current developments on the inhibitory effects of berry polyphenols on digestive enzymes. *Biofactors* **34**, 73-80. J

**McDougall, G.J., Kulkarni, N.N. & Stewart, D.** 2009. Berry polyphenols inhibit pancreatic lipase activity *in vitro*. *Food Chemistry* **115**, 193-199. <http://dx.doi.org/10.1016/j.foodchem.2008.11.093> J

**McDougall, G.J., Martinussen, I. & Stewart, D.** 2008. Development of high throughput analyses of

polyphenol composition in berries using abbreviated mass spectrophotometry techniques. *COST 863 Euroberry. Bioactive Compounds in Berry Fruits: Genetic Control, Breeding, Cultivar, Analytical Aspects and Human Health*, Zurich, Switzerland, 3-5 December 2008. P

**McGavin, W.J. & MacFarlane, S.A.** 2009. *Rubus chlorotic mottle virus*, a new Sobemovirus infecting raspberry and bramble. *Virus Research* **139**, 10-13.  
<http://dx.doi.org/10.1016/j.virusres.2008.09.004> J

**McKenzie, B.M.** 2009. Book review: Water and Cereals in Drylands. Edited by P Koohafkan & B.A. Stewart. Earthscan, London (2008). *Experimental Agriculture* **45**, 514.  
<http://dx.doi.org/10.1017/S0014479709990366> O

**McKenzie, B.M., Bengough, A.G., Hallett, P.D., Thomas, W.T.B., Forster, B.P. & McNicol, J.W.** 2009. Deep rooting and drought screening of cereal crops: A novel field-based method and its application. *Field Crops Research* **112**, 165-171. <http://dx.doi.org/10.1016/j.fcr.2009.02.012> J

**McKenzie, B.M., Kühner, S., MacKenzie, K., Peth, S. & Horn, R.** 2009. Soil compaction by uniaxial loading and the survival of the earthworm *Aporrectodea caliginosa*. *Soil and Tillage Research* **104**, 320-323.  
<http://dx.doi.org/10.1016/j.still.2009.04.004> J

**McKenzie, B.M., Valentine, T.A., Bengough, A.G. & Krol, M.** 2009. Root plasticity to water and its relevance for drought tolerance. *7th International Symposium of the Society for Root Research: Root Research and Applications*, Vienna, Austria, 2-4 September 2009. P

**McKinley, J., Ruffell, A., Harrison, M., Meier-Augenstein, W., Kemp, H., Graham, C. & Barry, L.** 2009. Spatial thinking in search methodology: a case study of the 'no body murder enquiry', west of Ireland. In: K. Ritz, L. Dawson & D. Miller, eds. *Criminal and Environmental Soil Forensics*, Springer, Dordrecht, The Netherlands, 285-302.  
[http://dx.doi.org/10.1007/978-1-4020-9204-6\\_18](http://dx.doi.org/10.1007/978-1-4020-9204-6_18) R

**McLellan, H.** 2008. The involvement of Cathepsin B-like genes in disease resistance in *Arabidopsis thaliana*. *University of Edinburgh*, PhD Thesis. O

**McLellan, H., Gilroy, E.M., Yun, B.W., Birch, P.R.J. & Loake, G.J.** 2009. Functional redundancy in the *Arabidopsis* Cathepsin B gene family contributes to basal defence, the hypersensitive response and senescence. *New Phytologist* **183**, 408-418.  
<http://dx.doi.org/10.1111/j.1469-8137.2009.02865.x> J

**McMenemy, L.S., MacFarlane, S.A., Hartley, S.E. & Johnson, S.N.** 2009. Co-operation between plant enemies - do raspberry viruses attract more aphid vectors. *8th International Symposium on Aphids*, Catania, Italy, 8-12 June 2009. P

**McMenemy, L.S., MacFarlane, S.A., Hartley, S.E. & Johnson, S.E.** 2009. Plant mediated interactions with the large raspberry aphid. *Ento'09 National Meeting of the Royal Entomological Society*, University of Sheffield, Sheffield, UK, 15-17 July 2009. P

**McMenemy, L.S., Mitchell, C. & Johnson, S.N.** 2009. Biology of the European large raspberry aphid (*Amphorophora idaei*); its role in virus transmission and resistance breakdown in red raspberry. *Agricultural and Forest Entomology* **11**, 61-71.  
<http://dx.doi.org/10.1111/j.1461-9563.2008.00409.x> J

**Meier-Augenstein, W. & Kemp, H.F.** 2009. Human provenancing based on stable isotope forensic intelligence. *5th European Academy of Forensic Science (EAFS)*, Glasgow, UK, 8-11 September 2009 (Talk). P

**Meier-Augenstein, W. & Kemp, H.F.** 2009. Human provenancing: it's elemental. *Copernicus Meetings*, European Geosciences Union, Vienna, Austria, 19-24 April 2009. P

**Meier-Augenstein, W., Kemp, H.F. & Lock, C.M.** 2009. N<sub>2</sub>: A potential pitfall for bulk <sup>2</sup>H isotope analysis of explosives and other nitrogen-rich compounds by continuous-flow isotope-ratio mass spectrometry. *Rapid Communications in Mass Spectrometry* **23**, 2011-2016.  
<http://dx.doi.org/10.1002/rcm.4112> J

**Messéan, A., Squire, G.R., Perry, J.N., Angevin, F., Gomez, M., Townend, P., Sausse, C., Breckling, B., Langrell, S., Dzeroski, S. & Sweet, J.** 2009. Sustainable introduction of GM crops into European agriculture: a summary report of the FP6 SIGMEA research project. *Oléagineux, Corps Gras, Lipides* **16**, 37-51. J

**Mickovski, S.B., Hallett, P.D., Bransby, M.F., Davies, M.C.R., Sonnenberg, R. & Bengough, A.G.** 2009. Mechanical reinforcement of soil by willow roots: impacts of root properties and root failure mechanism. *Soil Science Society of America Journal* **73**, 1276-1285.  
<http://dx.doi.org/10.2136/sssaj2008.0172> J

**Milbourne, D., Bradshaw, J.E. & Hackett, C.A.** 2009. Molecular mapping and breeding in polyploid crop plants. In: C. Kole & A.G. Abbott, eds. *Principles and Practices of Plant Genomics. Volume 2. Molecular Breeding*, Science Publishers Inc., Enfield, New Hampshire, USA, Chapter 10. R

**Milne, I., Lindner, D., Bayer, M., Husmeier, D., McGuire, G., Marshall, D.F. & Wright, F.** 2009. TOPALi v2: a rich graphical interface for evolutionary analyses of multiple alignments on HPC clusters and multi-core desktops. *Bioinformatics* **25**, 126-127.  
<http://dx.doi.org/10.1093/bioinformatics/btn575> J

**Mitchell, C. & Johnson, S.N.** 2009. Combining biological control and plant resistance to control the large raspberry aphid. *Ento'09 National Meeting of the Royal Entomological Society*, University of Sheffield, Sheffield, UK, 15-17 July 2009. P

**Moir, H.J., Gibbins, C.N., Buffington, J.M., Webb, J.H., Soulsby, C. & Brewer, M.J.** 2009. A new method to identify the fluvial regimes used by spawning salmonids. *Canadian Journal of Fisheries and Aquatic Sciences* **66**, 1404-1408.  
<http://dx.doi.org/10.1139/F09-136> J

**Morales, J.G.** 2008. Mechanisms of virulence and avirulence in the biotrophic interaction between potato and the late blight pathogen *Phytophthora infestans*. *University of Dundee*, PhD Thesis. O

**Morris, W.L. & Taylor, M.A.** 2009. The effects of processing and storage on potato tuber umami content: implications for product flavour and reduced salt formation. *Excellence in Food Manufacture 2009*, Central Science Laboratory, York, UK, 19 May 2009. P

**Morris, W.L., Ducreux, L., Hedley, P.E., Morris, J., Bryan, G.J., Ross, H.A. & Taylor, M.** 2009. Investigating key determinants of potato flavour and texture. *Euro Food Chem XV - Food for the Future Conference*, Copenhagen, Denmark, 5-8 July 2009 (Poster). P





- Murray, P.J., Clegg, C.D., Bristow, A.W., Gregory, P.J., Headon, D.M. & Clayton, S.J. 2009. Movement of newly assimilated carbon in the grass *Lolium perenne* and its incorporation into rhizosphere microbial DNA. *Simsug 2009*, University of Glasgow, Glasgow, UK, 14-15 January 2009. P
- Nart, P., Naylor, S.W., Huntley, J.F., McKendrick, I.J., Gally, D.L. & Low, J.C. 2008. Responses of cattle to gastrointestinal colonization by *Escherichia coli* O157:H7. *Infection and Immunity* **76**, 5366-5372. <http://dx.doi.org/10.1128/IAI.01223-07> J
- Neumann, G., George, T.S. & Plassard, C. 2009. Strategies and methods for studying the rhizosphere - the plant science toolbox. *Plant and Soil* **321**, 431-456. <http://dx.doi.org/10.1007/s11104-009-9953-9> J
- Newton, A.C. 2009. Book Review: Plant pathology. Concepts and laboratory exercises. 2nd edition. Edited by R.N. Trigiano, M.T. Windham and A.S. Windham. CRC Press/Taylor and Francis, Boca Raton, Florida, USA (2008). *Experimental Agriculture* **45**, 130-131. <http://dx.doi.org/10.1017/S0014479708007205> O
- Newton, A.C. 2009. Exploiting diversity in cereal production. *Scottish Society for Crop Research. Farming in Tough Times*, Perth, UK, 17 February 2009, (Abstract & Talk). P
- Newton, A.C. 2009. Plant disease control through the use of variety mixtures. In: D. Walters, ed. *Disease Control in Crops: Biological and Environmentally Friendly Approaches*, Wiley-Blackwell, Chichester, UK, 162-171. R
- Newton, A.C. & Guy, D.C. 2009. The effects of uneven patchy cultivar mixtures on disease control and yield in winter barley. *Field Crops Research* **110**, 225-228. <http://dx.doi.org/10.1016/j.fcr.2008.09.002> J
- Newton, A.C., Begg, G.S. & Swanston, J.S. 2009. Deployment of diversity for enhanced crop function. *Annals of Applied Biology* **154**, 309-322. <http://dx.doi.org/10.1111/j.1744-7348.2008.00303.x> J
- Orfanus, T., Bedrna, Z., Lichner, L., Hallett, P.D., Knava, K. & Sebin, M. 2008. Spatial variability of water repellency in pine forest soil. *Soil and Water* **3**, S123-S129. J
- Osterrieder, A., Carvalho, C.M., Latijnhouwers, M., Johansen, J.N., Stubbs, C., Botchway, S. & Hawes, C. 2009. Fluorescence lifetime imaging of interactions between Golgi tethering factors and small GTPases in plants. *Traffic* **10**, 1034-1046. <http://dx.doi.org/10.1111/j.1600-0854.2009.00930.x> J
- Öpik, M., Metsis, M., Daniell, T.J., Zobel, M. & Moora, M. 2009. Large-scale parallel 454 sequencing reveals host ecological group specificity of arbuscular mycorrhizal fungi in a boreonemoral forest. *New Phytologist* **184**, 424-437. <http://dx.doi.org/10.1111/j.1469-8137.2009.02920.x> J
- Palukaitis, P. 2008. Multiple pathways of resistance to TMV in *N* gene tobacco. *International Meeting of the Federation of Korean Microbiological Societies*, Seoul, Korea, 16-17 October 2008, 98-100 (Abstract). P
- Palukaitis, P. 2008. Multiple pathways of resistance to TMV in *N* gene tobacco. *Korean Society of Plant Pathology. International Symposium and Annual Meeting - New Approaches to Plant Disease Management*, Muju, Korea, 23-24 October 2008, 20-22 (Abstract). P
- Pawlett, M., Hopkins, D.W., Moffett, B.F. & Harris, J.A. 2009. The effect of earthworms and liming on soil microbial communities. *Biology and Fertility of Soils* **45**, 361-369. <http://dx.doi.org/10.1007/s00374-008-0339-6> J
- Peter, K.A., Gray, S.M., Gildow, F.E. & Palukaitis, P. 2008. Phloem limitation of *Potato leafroll virus* is selected for by the virus. *XIVth International Congress of Virology*, Istanbul, Turkey, 10-15 August 2008, 167 (Abstract). P
- Peter, K.A., Gildow, F.E., Palukaitis, P. & Gray, S.M. 2009. The C-terminus of the Poliovirus P5 readthrough domain limits virus infection to the phloem. *Journal of Virology* **83**, 5419-5429. <http://dx.doi.org/10.1128/JVI.02312-08> J
- Polechová, J., Barton, N. & Marion, G. 2009. Species' range: adaptation in space and time. *American Naturalist* **174**, E186-E204. <http://dx.doi.org/10.1086/605958> J
- Potokina, E., Druka, A., Luo, Z.W., Moscou, M., Wise, R., Waugh, R. & Kearsley, M.J. 2008. Tissue-dependent limited pleiotropy affects gene expression in barley. *Plant Journal* **56**, 287-296. <http://dx.doi.org/10.1111/j.1365-3113X.2008.03601.x> J
- Potokina, E., Druka, A., Luo, Z., Waugh, R. & Kearsley, M.J. 2009. The transcriptome analysis of barley (*Hordeum vulgare* L.) using the Affymetrix Barley1 GeneChip. *Russian Journal of Genetics* **45**, 1317-1328. <http://dx.doi.org/10.1134/S1022795409110064> J
- Prashar, A., Wishart, J., George, T.S., Brown, L.K., Thompson, J.A., Ramsay, G. & Bradshaw, J.E. 2009. Root architecture in potato: potential for improving resource capture and QTL mapping. *Plant Abiotic Stress - from signalling to development. International Conference*, Tartu, Estonia, 14-17 May 2009. P
- Pritchard, L., Liu, H., Booth, C., Douglas, E., François, P., Schrenzel, J., Hedley, P.E., Birch, P.R.J. & Toth, I.K. 2009. Microarray comparative genomic hybridisation analysis incorporating genomic organisation, and application to enterobacterial plant pathogens. *PLoS Computational Biology* **5**, e1000473. <http://dx.doi.org/10.1371/journal.pcbi.1000473> J
- Pswarayi, A., van Eeuwijk, F.A., Ceccarelli, S., Grando, S., Comadran, J., Russell, J.R., Francia, E., Pecchioni, N., Li Destri, O., Akar, T., Al-Yassin, A., Benbelkacem, A., Choumane, W., Karrou, M., Ouabbou, H., Bort, J., Araus, J.L., Molina-Cano, J.-L., Thomas, W.T.B. & Romagosa, I. 2008. Barley adaptation and improvement in the Mediterranean basin. *Plant Breeding* **127**, 554-560. <http://dx.doi.org/10.1111/j.1439-0523.2008.01522.x> J
- Raczynska, K.D., Konieczna, M., Ciesiolka, A., Szewc, L., Lewandowska, D., Simpson, C.G., Szweykowska-Kulinska, Z., Brown, J.W.S. & Jarmolowski, A. 2009. Alternative splicing in the *Arabidopsis thaliana* *HYL1* mutant. *4th Annual EURASNET Meeting*, Assisi, Italy, 23-24 April 2009 (Poster). P
- Rakhshanderhoo, F., Squires, J. & Palukaitis, P. 2008. RNA-dependent RNA polymerase - 1 can play a defensive role in systemic viral infection. *9th International Congress of Plant Pathology*, Torino, Italy, 24-29 August 2008. P
- Rakhshanderhoo, F., Takeshita, M., Squires, J. & Palukaitis, P. 2009. The influence of RNA-dependent RNA polymerase 1 on *Potato virus Y* infection and on other antiviral response genes. *Molecular Plant-Microbe Interactions* **22**, 1312-1318. <http://dx.doi.org/10.1094/MPMI-22-10-1312> J



- Ramirez-Farias, C., Slezak, K., Fuller, Z., Duncan, A., Holtrop, G. & Louis, P.** 2009. Effect of inulin on the human gut microbiota: stimulation of *Bifidobacterium adolescentis* and *Faecalibacterium prausnitzii*. *British Journal of Nutrition* **101**, 541-550. <http://dx.doi.org/10.1017/S0007114508019880> J
- Ramsay, G.** 2009. Book Review: Conserving plant genetic resources in protected areas. Edited by J.M. Iriondo, N. Maxted and M.E. Dulloo. CABI, Wallingford, Oxon (2008). *Experimental Agriculture* **45**, 510. <http://dx.doi.org/10.1017/S0014479709990287> O
- Raven, J.A.** 2009. Contributions of anoxygenic and oxygenic phototrophy and chemolithotrophy to carbon and oxygen fluxes in aquatic environments. *8th International Workshop of the Group for Aquatic Primary Productivity*, Eilat, Israel, April 2008. *Aquatic Microbial Ecology* **56**, 177-192. <http://dx.doi.org/10.3354/ame01315> P
- Raven, J.A.** 2009. Functional evolution of photochemical energy transformations in oxygen-producing organisms. *Functional Plant Biology* **36**, 505-515. <http://dx.doi.org/10.1071/FP09087> J
- Raven, J.A.** 2009. Horsetails get the wind up. *New Phytologist* **184**, 6-9. <http://dx.doi.org/10.1111/j.1469-8137.2009.3012.x> J
- Raven, J.A.** 2009. The evolution of plants. *Society for Experimental Biology Annual Main Meeting*, Glasgow, UK, 28 June-1 July 2009. P
- Raven, J.A. & Giordano, M.** 2009. Biomineralization by photosynthetic organisms: Evidence of coevolution of the organisms and their environment? *Geobiology* **7**, 140-154. <http://dx.doi.org/10.1111/j.1472-4669.2008.00181.x> J
- Raven, J.A., Beardall, J., Flynn, K.J. & Maberly, S.C.** 2009. Phagotrophy in the origins of photosynthesis in eukaryotes and as a complementary mode of nutrition in phototrophs: relation to Darwin's insectivorous plants. *Journal of Experimental Botany* **60**, 3975-3987. <http://dx.doi.org/10.1093/jxb/erp282> J
- Ravensdale, M.** 2008. A molecular study of the cfl and cfa gene cluster in *Pectobacterium atrosepticum*. University of Dundee, PhD Thesis. O
- Regeai, S.O., Dolan, K.M., Fitzpatrick, D.A., Browne, J.A., Jones, J.T. & Burnell, A.M.** 2009. Novel primers for the amplification of nuclear DNA introns in the entomopathogenic nematode *Heterorhabditis bacteriophora* and their cross-amplification in seven other *Heterorhabditis* species. *Molecular Ecology Resources* **9**, 421-424. <http://dx.doi.org/10.1111/j.1755-0998.2008.02431.x> J
- Rehman, S., Butterbach, P., Popeijus, H., Overmars, H., Davis, E.L., Jones, J.T., Goverse, A., Bakker, J. & Smant, G.** 2009. Identification and characterization of the most abundant cellulases in stylet secretions from *Globodera rostochiensis*. *Phytopathology* **99**, 194-202. <http://dx.doi.org/10.1094/PHYTO-99-2-0194> J
- Richardson, A.E., Hocking, P.J., Simpson, R.J. & George, T.S.** 2009. Plant mechanisms to optimise access to soil phosphorus. *Crop and Pasture Science* **60**, 124-143. <http://dx.doi.org/10.1071/CP07125> J
- Roberts, A.G., Cowan, G.H., Savenkov, E.I., Ziegler, A., Chapman, S., Wright, K.M. & Torrance, L.** 2009. Role of TGB1 in the cell-cell and long-distance movement of *Potato mop top virus*. *Association of Applied Biologists. Advances in Plant Virology*, Harrogate, UK, 1-3 April 2009. P
- Romagosa, I., van Eeuwijk, F.A. & Thomas, W.T.B.** 2009. Statistical analyses of genotype by environment data. In: M.J. Carena, ed. *Cereals, Handbook of Plant Breeding*, Springer, Dordrecht, The Netherlands, 291-332. R
- Ross-Adams, H., Pasdar, A., Mayer, C.D. & MacLeod, M.J.** 2009. Validation of MAP2K6 as a novel stroke candidate gene. *International Stroke Conference 2009*, San Diego, California, USA, 17-20 February 2009. *Stroke* **40**, E179. P
- Rosso, M.N., Jones, J.T. & Abad, P.** 2009. RNAi and functional genomics in plant parasitic nematodes. *Annual Review of Phytopathology* **47**, 207-232. <http://dx.doi.org/10.1146/annurev.phyto.112408.132605> J
- Russell, J.R., Kadu, C.A.C., Jamnadass, R., Booth, A., Cordeiro, N.J., Woodhead, M. & Dawson, I.K.** 2009. AFLP and SSR diversity in the African fruit tree *Allanblackia*: Implications for management of a genus newly subject to domestication for the edible oil industry. *Tree Genetics and Genomes* **5**, 517-527. <http://dx.doi.org/10.1007/s11295-009-0205-1> J
- Ryder, C., Barker, B., Teakle, G., Lynn, J., Pink, D.A.C., Deswarte, J.C., Farquhar, G., White, P.J. & Thompson, A.J.** 2008. Water-use-efficiency genes in *Brassica oleracea*. *5th ISHS International Symposium on Brassicas and 16th Crucifer Genetics Workshop*, Lillehammer, Norway, 8-12 September 2008, 42 (Talk). P
- Salse, J., Abrouk, M., Bolot, S., Guilhot, N., Courcelle, E., Faraut, T., Waugh, R., Close, T.J., Messing, J. & Feuillet, C.** 2009. Reconstruction of monocotyledonous proto-chromosomes reveals faster evolution in plants than in animals. *Proceedings of the National Academy of Sciences, USA* **106**, 14908-14913. <http://dx.doi.org/10.1073/pnas.0902350106> J
- Santos, C., Tavares, L.R., Fortalezas, S., Carillho, D., Pontes, V., McDougall, G.J., Stewart, D. & Ferreira, R.B.** 2009. Neuroprotective and MMP-9 inhibitory activity of hydroethanolic extract of *Arbustus unedo* leaves. *57th International Congress and Annual Meeting of the Society for Medicinal Plant Research and Natural Product Research*, Geneva, Switzerland, 16-20 August 2009. *Planta Medica* **75**, 924. P
- Schena, L., Cardle, L. & Cooke, D.E.L.** 2008. Use of genome sequence data in the design and testing of SSR markers for *Phytophthora* species. *BMC Genomics* **9**, 620. <http://dx.doi.org/10.1186/1471-2164-9-620> J
- Schreiber, A.W.J., Sutton, T., Caldo, R.A., Kalashyan, E., Lovell, B., Mayo, G., Muehlbauer, G., Druka, A., Waugh, R., Wise, R.P., Langridge, P. & Baumann, U.** 2009. Comparative transcriptomics in the Triticeae. *BMC Genomics* **10**, 285. <http://dx.doi.org/10.1186/1471-2164-10-285> J
- Schulte, D., Close, T.J., Graner, A., Langridge, P., Matsumoto, T., Muehlbauer, G.J., Sato, K., Schulman, A.H., Waugh, R., Wise, R.P. & Stein, N.** 2009. International Barley Sequencing Consortium – at the threshold of efficient access to the barley genome. *Plant Physiology* **149**, 142-147. <http://dx.doi.org/10.1104/pp.108.128967> J
- Shand, C.A., Stutter, M., George, T.S., Mackay, G., Haygarth, P., Bol, R. & Dixon, L.** 2009. <sup>31</sup>P NMR study of phosphorus in soils. *42nd International Union of Pure and Applied Chemistry 2009 Meeting*, SECC, Glasgow, UK, 2-7 August 2009. P
- Shaw, D.S., Evans, D., Bufo, C. & Cooke, D.E.L.** 2008. Can UK populations of *P. infestans* mate? *11th Euroblight Workshop*, Hamar, Norway, 28-31 October 2008. P



- Shaw, J., Canetta, E., Kim, S.-H., Kalinina, N.O., Adya, A.K., Brown, J.W.S. & Taliansky, M.E.** 2009. A plant virus movement protein forms ringlike complexes with major nucleolar protein, fibrillarin *in vitro*. *Association of Applied Biologists. Advances in Plant Virology*, Harrogate, UK, 1-3 April 2009. P
- Shepherd, T., Birch, A.N.E., Jorna, C.S., Mitchell, C., Cross, J., Hall, D. & Farman, D.** 2009. Profiling of raspberry cane wound volatiles using a combination of SPME and GC-TOF-MS. *Metabomeeting 2009*, Norwich, UK, 5-8 July 2009. P
- Shi, B.-J., Symons, R.H. & Palukaitis, P.** 2009. Stability and competitiveness of interval recombinant RNAs derived from a chimeric cucumovirus. *Virus Research* **140**, 216-221. <http://dx.doi.org/10.1016/j.virusres.2008.11.009> J
- Simpson, C.G., Chapman, S., Liney, M., Davidson, D., Lewandowska, D. & Brown, J.W.S.** 2009. *Arabidopsis* PTB-like 1 (AtPTBL1) negatively regulates splicing inclusion of a plant mini-exon. *20th International Conference on Arabidopsis Research*, Edinburgh, UK, 30 June-4 July 2009 (Poster). P
- Simpson, C.G., Kalyna, M., Fuller, J., Davidson, D., Barta, A. & Brown, J.W.S.** 2009. Intron retention in *Arabidopsis* mRNA transcripts. *20th International Conference on Arabidopsis Research*, Edinburgh, UK, 30 June-4 July 2009 (Poster). P
- Simpson, C.G., Kalyna, M., Fuller, J., Davidson, D., Barta, A. & Brown, J.W.S.** 2009. The frequency of intron retention in *Arabidopsis* is over-estimated. *4th Annual EURASNET Meeting*, Assisi, Italy, 23-24 April 2009 (Poster). P
- Simpson, C.G., Kalyna, M., Fuller, J., Davidson, D., Barta, A. & Brown, J.W.S.** 2009. The frequency of intron retention in *Arabidopsis* is over-estimated. *RNA 2009*, Madison, Wisconsin, USA, 26-31 May 2009 (Poster). P
- Simpson, C.G., Kalyna, M., Lewandowska, D., Kusenda, B., Fuller, J., Cardle, L., McNicol, J.W., Clark, G., Barta, A. & Brown, J.W.S.** 2009. Alternative splicing and NMD in *Arabidopsis*. *RNA 2009*, Madison, Wisconsin, USA, 26-31 May 2009 (Poster). P
- Simpson, C.G., Kalyna, M., Lewandowska, D., Kusenda, B., Fuller, J., Cardle, L., McNicol, J.W., Clark, G., Barta, A. & Brown, J.W.S.** 2009. Alternative splicing and NMD in *Arabidopsis*. *4th Annual EURASNET Meeting*, Assisi, Italy, 23-24 April 2009 (Poster). P
- Simpson, C.G., Kalyna, M., Lewandowska, D., Kusenda, B., Fuller, J., Cardle, L., McNicol, J.W., Clark, G., Barta, A. & Brown, J.W.S.** 2009. Alternative splicing and NMD in *Arabidopsis*. *20th International Conference on Arabidopsis Research*, Edinburgh, UK, 30 June-4 July, 2009 (Poster). P
- Simpson, C.G., Kalyna, M., Lewandowska, D., Kusenda, B., Fuller, J., Cardle, L., McNicol, J.W., Clark, G., Syed, N.H., Marshall, J., Barta, A. & Brown, J.W.S.** 2009. Alternative splicing and NMD in *Arabidopsis*. *RNA UK 2010*, Lake District, UK, 22-24 January 2009 (Talk). P
- Sir, M., Lichner, L., Tesar, M., Hallett, P.D. & Martinkova, M.** 2009. Simulation of phytomass productivity based on the optimum temperature for plant growth in a cold climate. *2nd International Conference Biohydrology 2009*, Bratislava, Slovakia, 21-24 September 2009. *Biologia* **64**, 615-619. P
- Smith, L.A., Marion, G., Swain, D.L., White, P.C.L. & Hutchings, M.R.** 2009. Inter- and intra-specific exposure to parasites and pathogens via the faecal-oral route: a consequence of behaviour in a patchy environment. *Epidemiology and Infection* **137**, 630-643. <http://dx.doi.org/10.1017/S0950268808001313> J
- Smith, L.A., Marion, G., Swain, D.L., White, P.C.L. & Hutchings, M.R.** 2009. Livestock grazing behavior and inter- versus intraspecific disease risk via the fecal oral route. *Behavioral Ecology* **20**, 426-432. <http://dx.doi.org/10.1093/beheco/arn143> J
- Song, B.K., Hein, I., Druka, A., Waugh, R., Marshall, D.F., Nadarajah, K., Yap, S.J. & Ratnam, W.** 2009. The 172 kb genomic DNA region of the *O. rufipogon* *ylg1.1* locus: Comparative sequence analysis with *O. sativa* ssp. *japonica* and *O. sativa* ssp. *indica*. *Functional & Integrative Genomics* **9**, 97-108. <http://dx.doi.org/10.1007/s10142-008-0091-x> J
- Spezia, L.** 2009. Reversible jump and the label switching problem in hidden Markov models. *Journal of Statistical Planning and Inference* **139**, 2305-2315. <http://dx.doi.org/10.1016/j.jspi.2008.10.016> J
- Sprent, J.I.** 2009. An interdisciplinary look at legumes and their bacterial symbionts: Some thoughts from Big Sky. *New Phytologist* **184**, 15-17. <http://dx.doi.org/10.1111/j.1469-8137.2009.03006.x> J
- Squire, G.R.** 2009. Book Review: Sustainable rural systems. Sustainable agriculture and rural communities. Edited by G.M. Robinson. Ashgate Publishing, Aldershot (2008). *Experimental Agriculture* **45**, 379. O
- Squire, G.R., Hawes, C., Begg, G.S. & Young, M.W.** 2009. Cumulative impact of GM herbicide-tolerant cropping on arable plants assessed through species-based and functional taxonomies. *Environmental Science and Pollution Research* **16**, 85-94. <http://dx.doi.org/10.1007/s11356-008-0072-6> J
- Squires, J. & Palukaitis, P.** 2009. The effect of CaMV integration in *Arabidopsis* on the growth and fecundity of plants and the stability and expression of GFP transgene. *Association of Applied Biologists. Advances in Plant Virology*, Harrogate, UK, 1-3 April 2009, 16 (Abstract). P
- Srinivasan, D.G., Año, L., Fenton, B., Jaubert-Possamai, S., Tagu, D. & Edwards, O.** 2009. Molecular method of facultative parthenogenesis of pea aphid, *Acyrtosiphon pisum*. *8th International Symposium on Aphids*, Catania, Italy, 8-12 June 2009. P
- Staley, J.T. & Johnson, S.N.** 2008. Climate change impacts on root herbivores. In: S.N. Johnson & P.J. Murray, eds. *Root Feeders: An Ecosystem Perspective*, CABI, Wallingford, UK, 192-213. R
- Stark, L.A. & Taliansky, M.E.** 2008. Old and new faces of the nucleolus. Workshop on the nucleolus and disease. *EMBO Reports* **10**, 35-40. <http://dx.doi.org/10.1038/embor.2008.230> J
- Stewart, D.** 2009. Blackcurrant - a source of human health beneficial phytochemicals. *2nd International Conference: Berries and Human Health. International Berry Health Benefits Symposium*, Monterey, California, USA, 21-24 June 2009 (Abstract). P
- Stokes, A., Atger, C., Bengough, A.G., Fourcaud, T. & Sidle, R.C.** 2009. Desirable plant root traits for protecting natural and engineered slopes against landslides (Invited review). *Plant and Soil* **324**, 1-30. <http://dx.doi.org/10.1007/s11104-009-0159-y> J

- Subramanian, N.K., Ramsay, G., White, P.J. & Broadley, M.R.** 2008. Studies on the distribution of minerals in potato tubers. *Society for Experimental Biology Plant Transport Group Meeting*, Manchester, UK, 3-5 September 2008 (Poster). P
- Subramanian, N.K., Ramsay, G., White, P.J. & Broadley, M.R.** 2008. Variation in mineral content within potato tubers and between genotypes. *Global Potato Conference 2008*, New Delhi, India, 9-12 December 2008 (Poster). P
- Subramanian, N.K., Ramsay, G., White, P.J. & Broadley, M.R.** 2009. Exploiting genetic variation for elevated mineral concentrations in potatoes. *EUCARPIA, XIV Meeting of the Biometrics in Plant Breeding Section Biometrics Meeting*, Dundee, 2-4 September 2009 (Poster). P
- Sun, B., Hallett, P.D., Caul, S., Barnett, C.M. & Hopkins, D.W.** 2009. Distribution of soil carbon and microbial biomass under different tillage regimes. *International Symposium on Soil Organic Matter Dynamics: Land Use, Management and Global Change*, Colorado State University, Colorado, USA, 6-10 July 2009. P
- Sun, B., Hallett, P.D., Caul, S., Barnett, C.M. & Hopkins, D.W.** 2009. Distribution of soil carbon and microbial biomass under different tillage regimes. *Soil Organic Matters International Conference*, Rothamsted Research, Harpenden, UK, 23-25 June 2009. P
- Swanson, M.M., Fraser, G., Daniell, T.J., Torrance, L., Gregory, P.J. & Taliansky, M.E.** 2009. Viruses in soils: morphological diversity and abundance in the rhizosphere. *Annals of Applied Biology* **155**, 51-60. <http://dx.doi.org/10.1111/j.1744-7348.2009.00319.x> J
- Tavares, L.R., Pimpao, R.C., Santos, C., McDougall, G.J., Stewart, D. & Ferreira, R.B.** 2009. Phytochemical characterisation of *Juniperus* spp. leaves. *57th International Congress and Annual Meeting of the Society for Medicinal Plant Research and Natural Product Research*, Geneva, Switzerland, 16-20 August 2009. *Planta Medica* **75**, 923. P
- Tavares, L.R., Santos, C.N., McDougall, G.J., Fortalezas, S., Carrilho, D., Stewart, D. & Ferreira, R.B.** 2009. Phytochemical profiling of different tissues from Portuguese endemic *Rubus* species. *COST 863 Management Workshop on Defining Needs of Berry Industries*, Lisbon, Portugal, 19-21 March 2009 (Talk). P
- Tavares, L.R., Santos, C.N., McDougall, G.J., Stewart, D. & Ferreira, R.B.** 2008. Portuguese endemic wild blackberries as an alternative source of polyphenols and antioxidant activity. *COST 863 Euroberry. Bioactive Compounds in Berry Fruits: Genetic Control, Breeding, Cultivar, Analytical Aspects and Human Health*, Zurich, Switzerland, 3-6 December 2008. P
- Teakle, G. & White, P.J.** 2008. Nitrogen use efficiency in *Brassica napus*. *UK Brassica Research Community Annual Meeting*, Warwick HRI, UK, 21 May 2008. P
- Teakle, G., Durnford, J., Stevenson, S., Foulkes, J., White, P.J., Berry, P. & Pink, D.A.C.** 2008. Genetic diversity for nitrogen use efficiency traits in oilseed rape. *5th ISHS International Symposium on Brassicas and 16th Crucifer Genetics Workshop*, Lillehammer, Norway, 8-12 September 2008, 86 (Poster). P
- Terzi, L.C. & Simpson, G.G.** 2009. *Arabidopsis* RNA immunoprecipitation. *Plant Journal* **59**, 163-168. <http://dx.doi.org/10.1111/j.1365-3113X.2009.03859.x> J
- Terzi, L.C. & Simpson, G.G.** 2009. *In vivo* targets of FPA, an RNA binding protein controlling *Arabidopsis* flower development. *20th International Conference on Arabidopsis Research*, Edinburgh, UK, 30 June-4 July 2009 (Poster). P
- Thiel, T., Graner, A., Waugh, R., Grosse, I., Close, T.J. & Stein, N.** 2009. Evidence and evolutionary analysis of ancient whole-genome duplication in barley predating the divergence from rice. *BMC Evolutionary Biology* **9**, 209. <http://dx.doi.org/10.1186/1471-2148-9-209> J
- Thompson, A.J. & White, P.J.** 2008. Identification of genetic markers for water-use-efficiency (WUE) in horticultural crops. *UK Brassica Research Community Annual Meeting*, Warwick HRI, UK, 21 May 2008. P
- Thorsen, M.K., Woodward, S., Hopkins, D.W. & McKenzie, B.M.** 2008. Resilience of Machair soil to amendment with kelp and synthetic fertilizer. *Glasgow Natural History Society. Machair Conservation: Successes and Challenges*, Glasgow, UK, 8 December 2008 (Poster). P
- Tilston, E.L., Szili-Kovács, T. & Hopkins, D.W.** 2009. Contributions of labile and resistant organic materials to the immobilization of inorganic soil N when used in the restoration of abandoned agricultural fields. *Soil Use and Management* **25**, 168-174. <http://dx.doi.org/10.1111/j.1475-2743.2009.00213.x> J
- Torrance, L.** 2009. Extreme resistance to Potyviruses in edible diploid potatoes of the Phureja - Stenotomum group. *Association of Applied Biologists. Advances in Plant Virology*, Harrogate, UK, 1-3 April 2009. P
- Torrance, L., Cowan, G.H., Ziegler, A. & Savenkov, E.I.** 2008. *Potato mop-top virus* RNAs display different requirements for long-distance movement. *Proceedings of 7th Symposium of the International Working Group on Plant Viruses with Fungal Vectors*, Quedlinburg, Germany, 1-4 September 2008. P
- Torrance, L., Lukhovitskaya, N.I., Schepetilnikov, M.V., Cowan, G.H., Ziegler, A. & Savenkov, E.I.** 2009. Unusual long-distance movement strategies of *Potato mop-top virus* RNAs in *Nicotiana benthamiana*. *Molecular Plant-Microbe Interactions* **22**, 381-390. <http://dx.doi.org/10.1094/MPMI-22-4-0381> J
- Uibopuu, A., Moora, M., Saks, U., Daniell, T.J., Zobel, M. & Öpik, M.** 2009. Differential effect of arbuscular mycorrhizal fungal communities from ecosystems along management gradient on the growth of forest understorey plant species. *Soil Biology and Biochemistry* **41**, 2141-2146. <http://dx.doi.org/10.1016/j.soilbio.2009.07.026> J
- Valentine, T.A., Binnie, K., Barnett, C.M., McKenzie, B.M. & Bengough, A.G.** 2009. Root responses to soil physical constraints: Quantitative gene expression analysis. *7th International Symposium of the Society for Root Research: Root Research and Applications*, Vienna, Austria, 2-4 September 2009. P
- Valentine, T.A., Wuyts, N., Roberts, T.J., Du, C.J., McKenna, S.J., Bransby, M.F. & Bengough, A.G.** 2009. Plant Vis: A new software tool for analysis of root growth dynamics. *7th International Symposium of the Society for Root Research: Root Research and Applications*, Vienna, Austria, 2-4 September 2009 (Abstract). P
- van Dijk, J.P., Cankar, K., Scheffer, S.J., Beenen, H.G.J., Shepherd, L.V.T., Stewart, D., Davies, H.V., Wilcockson, S.J., Leifert, C., Gruden, K. & Kok, E.J.**





2009. Transcriptome analysis of potato tubers - effects of different agricultural practices. *Journal of Agricultural and Food Chemistry* **57**, 1612-1623.

<http://dx.doi.org/10.1021/jf802815d> J

**van Poppel, P.M.J.A., Guo, J., van de Vondervoort, P.J.I., Jung, M.W.M., Birch, P.R.J., Whisson, S.C. & Govers, F.** 2008. The *Phytophthora infestans* avirulence gene *Avr4* encodes an RXLR-dEER effector. *Molecular Plant-Microbe Interactions* **21**, 1460-1470.

<http://dx.doi.org/10.1094/MPMI-21-11-1460> J

**Van Toor, R., Malloch, G. & Fenton, B.** 2009. A concept for management of virus vectors and insecticide resistance in *Myzus persicae* on potatoes. *8th International Symposium on Aphids*, Catania, Italy, 8-12 June 2009 (Talk). P

**Vida, G., Gál, M., Uhrin, A., Veisz, O., Syed, N.H., Flavell, A.J., Wang, Z.L. & Bedő, Z.** 2009. Molecular markers for the identification of resistance genes and marker-assisted selection in breeding wheat for leaf rust resistance, *18th General Congress of the European Association for Research on Plant Breeding*, Valencia, Spain, 12 September 2008. *Euphytica* **170**, 67-76. <http://dx.doi.org/10.1007/s10681-009-9945-0> P

**Vink, S., Neilson, R., Robinson, D. & Daniell, T.J.** 2008. Above and below ground responses to the Machair agricultural system. *Glasgow Natural History Society. Machair Conservation Successes and Challenges*, Glasgow, UK, 8 December 2008. P

**Vink, S., Neilson, R., Robinson, D. & Daniell, T.J.** 2009. Belowground microbial communities in a low input agricultural system. *15th Molecular Microbial Ecology Group Meeting*, University of Aberdeen, Aberdeen, UK, 29-30 July 2009. P

**Vinten, A.J.A., Potts, J., Avery, L. & Strachan, N.J.C.** 2009. Microbial pollution of water by livestock: approaches to risk assessment and mitigation. *Animal* **3**, 744-752. <http://dx.doi.org/10.1017/S1751731109004005> J

**Wallington, E., Bentley, S., Rymer, C., Stewart, A., Broadley, M.R., Berry, S., White, P.J. & Greenland, A.** 2009. Development and evaluation of low-phytate wheat germplasm to reduce diffuse phosphate pollution from pig and poultry production units. *MONOGRAM Network Workshop*, Burwalls, Bristol, UK, 29 April-1 May 2009. P

**Wang, M.H., Hu, X.H., Li, G., Leach, L.J., Potokina, E., Druka, A., Waugh, R., Kearsley, M.J. & Luo, Z.W.** 2009. Robust detection and genotyping of single feature polymorphisms from gene expression data. *PLoS Computational Biology* **5**, e1000317. <http://dx.doi.org/10.1371/journal.pcbi.1000317> J

**Waugh, R.** 2008. Genetics of barley quality. *2008 Worldwide Distilled Spirits Conference*, Edinburgh, UK, 8-10 September 2008. P

**Waugh, R., Jannink, J.L., Muehlbauer, G.J. & Ramsay, L.** 2009. The emergence of whole genome association scans in barley. *Current Opinion in Plant Biology* **12**, 218-222. <http://dx.doi.org/10.1016/j.pbi.2008.12.007> J

**Weaver, J., Briscoe, T., Hou, M., Goodman, C., Kata, S., Ross, H.A., McDougall, G.J., Stewart, D. & Riches, A.** 2009. Strawberry polyphenols are equally cytotoxic to tumourigenic and normal human breast and prostate cell lines. *International Journal of Oncology* **34**, 777-786. [http://dx.doi.org/10.3892/ijo\\_00000203](http://dx.doi.org/10.3892/ijo_00000203) J

**Wemelsfelder, F., Nevison, I.M. & Lawrence, A.B.** 2009. The effect of perceived environmental background on qualitative assessments of pig behaviour. *Animal Behaviour* **78**, 477-484. <http://dx.doi.org/10.1016/j.anbehav.2009.06.005> J

**Wheatley, R.E.** 2009. Book Review: Nitrogen-fixing leguminous symbioses. Nitrogen fixation: Origins, applications and research progress. Volume 7. Edited by M.J. Dilworth, E.K. James, J.I. Sprent and W.E. Newton. Springer Science, Dordrecht, The Netherlands (2008). *Experimental Agriculture* **45**, 239-240. <http://dx.doi.org/10.1017/S0014479708007370> O

**Wheatley, R.E.** 2009. Compost and Soils. *WRAP/NFUS Compost in Action Workshop*, Bachelton House, Perth, UK, 28 May 2009. P

**Wheatley, R.E.** 2009. Using quality compost in potato production to increase yields. WRAP, Banbury, UK. [http://www.wrap.org.uk/downloads/Case\\_study\\_-\\_Potatoes\\_in\\_Practice\\_2008.22d824df.6562.pdf](http://www.wrap.org.uk/downloads/Case_study_-_Potatoes_in_Practice_2008.22d824df.6562.pdf) O

**Whisson, S.C., Grenville-Briggs, L.J., van West, P. & Avrova, A.O.** 2009. Mechanism and application of gene silencing in oomycetes. In: K.H. Lamour & S. Kamoun, eds. *Oomycete Genetics and Genomics: Diversity, Interactions and Research Tools*, Wiley-Blackwell, Hoboken, New Jersey, USA, 493-515. R

**White, P.C.L., Böhm, M., Marion, G. & Hutchings, M.R.** 2008. Control of bovine tuberculosis in British livestock: there is no 'silver bullet'. *Trends in Microbiology* **16**, 420-427. <http://dx.doi.org/10.1016/j.tim.2008.06.005> J

**White, P.J.** 2009. Book Review: The Use of Nutrients in Crop Plants. Edited by N.K. Fageria. CRC Press, Boca Raton, Florida (2009). *Experimental Agriculture* **45**, 380. <http://dx.doi.org/10.1017/S0014479709007789> O

**White, P.J.** 2009. Book Review: Efficiency of soil and fertilizer phosphorus use: Reconciling changing concepts of soil phosphorus behaviour with agronomic information. By J.K. Syers, A.E. Johnston and D. Curtis. FAO, Rome (2008). *Experimental Agriculture* **45**, 128. <http://dx.doi.org/10.1017/S0014479709007789> O

**White, P.J.** 2009. Depolarisation-activated calcium channels shape the calcium signatures induced by low-temperature stress. *New Phytologist* **183**, 6-8. <http://dx.doi.org/10.1111/j.1469-8137.2009.02857.x> J

**White, P.J.** 2009. Sustainable approaches to crop mineral nutrition. *Knowledgescotland Science Policy Connections Online*. <http://www.knowledgescotland.org/briefings.php?id=83> O

**White, P.J. & Broadley, M.R.** 2009. Biofortification of crops with seven mineral elements often lacking in human diets - iron, zinc, copper, calcium, magnesium, selenium and iodine (Review). *New Phytologist* **182**, 49-84. <http://dx.doi.org/10.1111/j.1469-8137.2008.02738.x> J

**White, P.J. & Hammond, J.P.** 2009. The sources of phosphorus in the waters of Great Britain. *Journal of Environmental Quality* **38**, 13-26. <http://dx.doi.org/10.2134/jeq2007.0658> J

**White, P.J., Bengough, A.G., Bingham, I.J., George, T.S., Karley, A.J. & Valentine, T.A.** 2009. Induced mutations affecting root architecture and mineral acquisition in barley. In: Q.Y. Shu, ed. *Induced Plant Mutations in the Genomics Era*, Food and Agriculture Organisation of the United Nations, Rome, Italy, 2009., 338-340. R



- White, P.J., Bradshaw, J.E., Dale, M.F.B., Ramsay, G., Hammond, J.P. & Broadley, M.R.** 2009. Relationships between yield and mineral concentrations in potato tubers. *Hortscience* **44**, 6-11. J
- White, P.J., Bradshaw, J.E., George, T.S., Hammond, J.P. & Thompson, A.J.** 2008. The need for improved resource use by potatoes – water and mineral elements. *Improving International Potato Production*, Dundee, UK, 6 August 2008 (Invited Talk). P
- White, P.J., Karley, A.J. & Broadley, M.R.** 2009. Increasing dietary K, Mg and Ca intake through plants. *SciTopics Agricultural and Biological Sciences*, [http://www.scitopics.com/Increasing\\_dietary\\_K\\_Mg\\_and\\_Ca\\_intake\\_through\\_plants.html](http://www.scitopics.com/Increasing_dietary_K_Mg_and_Ca_intake_through_plants.html) O
- White, P.J., Wiesel, L. & Broadley, M.R.** 2008. Caesium uptake by root cells: Revisiting a paradigm. *Society for Experimental Biology. Plant Transport Group Meeting*, Manchester, UK, 3-5 September 2008 (Talk). P
- Wiesel, L. & White, P.J.** 2009. Do arbuscular mycorrhizal fungi influence caesium uptake by *Medicago truncatula*? *International Conference on Mycorrhiza (ICOM 6)*, Belo Horizonte, Brazil, 9-14 August 2009. P
- Wiesel, L. & White, P.J.** 2009. Mycorrhizal colonization does not seem to influence caesium uptake by *Medicago truncatula* grown under potassium deficient conditions. *Society for Experimental Biology Annual Meeting 2009*, Glasgow, UK, 28 June-1 July 2009. P
- Wishart, J., George, T.S., Brown, L.K., Thompson, J.A., Ramsay, G., Bradshaw, J.E., White, P.J. & Gregory, P.J.** 2009. Variation in rooting habit of potatoes: Potential for improving resource capture. *7th International Symposium of the Society for Root Research: Root Research and Applications*, Vienna, Austria, 2-4 September 2009. P
- Wojciechowski, T., Gooding, M.J., Ramsay, L. & Gregory, P.J.** 2009. The effects of dwarfing genes on seedling root growth of wheat. *Journal of Experimental Botany* **60**, 2565-2573. <http://dx.doi.org/10.1093/jxb/erp107> J
- Woodward, F.I., Bardgett, R.D., Raven, J.A. & Hetherington, A.M.** 2009. Biological approaches to global environmental change: mitigation and remediation. *Current Biology* **19**, R615-R623. <http://dx.doi.org/10.1016/j.cub.2009.06.012> J
- Wookey, P.A., Aerts, R., Bardgett, R.D., Baptist, F., Brathan, K.A., Cornelissen, J.H.C., Gough, L., Hartley, I.P., Hopkins, D.W., Laverer, S. & Shaver, G.R.** 2009. Ecosystem feedbacks and cascade processes: understanding their role in the responses of Arctic and alpine ecosystems to environmental change. *Global Change Biology* **15**, 1153-1172. <http://dx.doi.org/10.1111/j.1365-2486.2008.01801.x> J
- Wright, H.W., Bartley, K., Nisbet, A.J., McDevitt, R.M., Sparks, N.H.C., Brocklehurst, S. & Huntley, J.F.** 2009. The testing of antibodies raised against poultry red mite antigens in an *in vitro* feeding assay; preliminary screen for vaccine candidates. *Experimental and Applied Acarology* **48**, 81-91. <http://dx.doi.org/10.1007/s10493-009-9243-5> J
- Wypijewski, K., Hornyik, C., Shaw, J.A., Stephens, J., Goraczniak, R., Gunderson, S.I. & Lacomme, C.** 2009. Ectopic 5' splice sites inhibit gene expression by engaging RNA surveillance and silencing pathways in plants. *Plant Physiology* **151**, 955-965. <http://dx.doi.org/10.1104/pp.109.139733> J
- Xiao, H., McMahon, M., Hayes, J.D. & Stewart, D.** 2009. A mechanistic approach to fruit phytochemicals and cancer. *2nd International Conference: Berries and Human Health. International Berry Health Benefits Symposium*, Monterey, California, USA, 21-24 June 2009 (Abstract). P
- Yang, D., Zhang, T., Zhang, K., Greenwood, D.J., Hammond, J.P. & White, P.J.** 2009. An easily implemented agro-hydrological procedure with dynamic root simulation for water transfer in the crop-soil system: Validation and application. *Journal of Hydrology* **370**, 177-190. <http://dx.doi.org/10.1016/j.jhydrol.2009.03.005> J
- Zazzo, A., Moloney, A.P., Monahan, F.J., Scrimgeour, C.M. & Schmidt, O.** 2008. Effect of age and food intake on dietary carbon turnover recorded in sheep wool. *Rapid Communications in Mass Spectrometry* **22**, 2937-2945. <http://dx.doi.org/10.1002/rcm.3693> J
- Zhang, K., Greenwood, D.J., Hammond, J.P., White, P.J. & Burns, I.G.** 2008. A generic combined model for NPK fertilizer for vegetable and arable crops. *Proceedings of the International Symposium on Plant Nutrition Management in Sustainable Agriculture*, Nanchang, China, 13-16 October 2008, 43-51. P
- Ziegler, A., Cowan, G.H. & Torrance, L.** 2009. Comparative sequence analysis and serological and infectivity studies indicate that cocksfoot mild mosaic virus is a member of the genus *Panicovirus*. *Archives of Virology* **154**, 1545-1549. <http://dx.doi.org/10.1007/s00705-009-0468-8> J

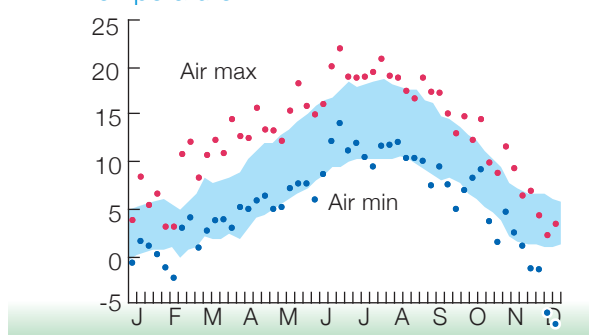
# Meteorological Records

2009

Marion Grassie

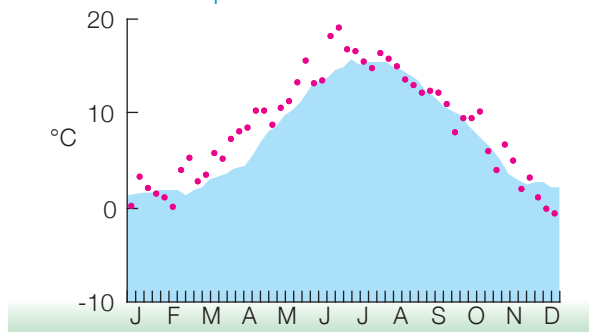
The main features of 2009 were high rainfall, flooding, higher than normal air and soil temperatures and perhaps surprisingly, higher than average sunshine. Outstanding months were March, with record sunshine, warmer temperatures and low rainfall; November with record rainfall; and December which was a particularly cold month breaking a number of our records, held since 1954.

## Air temperature



**Temperature** All months boasted average maximum air temperatures higher than the Long Term Average (LTA), with the exceptions of January, fractionally under the LTA of 5.9°C at 5.8°C, and December, at 4.7°C (LTA 6.7°C). Most significant were March, April and September. The highest maximum air temperature recorded was July's 26.0°C.

## 10cm soil temperature

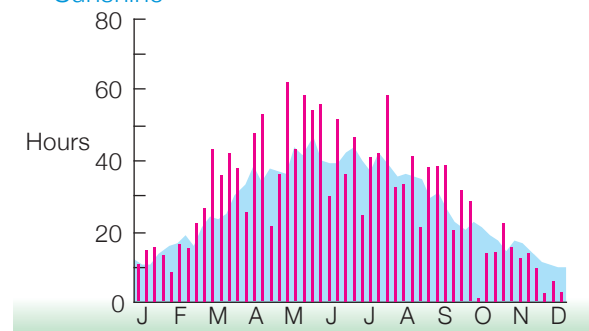


Blue areas on figures indicate long term average 1961–1990

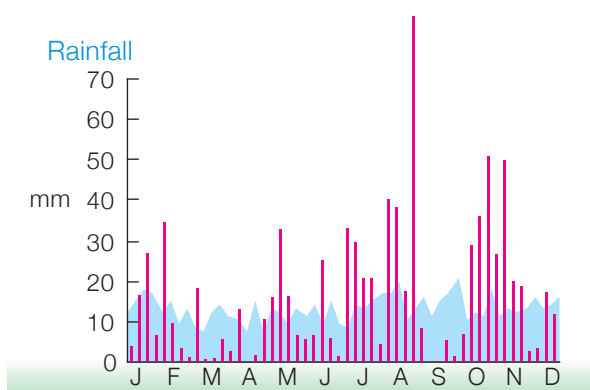
Average minimum air temperatures were all equal to or higher than the LTA with only one exception, December. April's minimum was 5.6°C (LTA 3.7°C). August, with a minimum air temperature of 8.9°C (LTA 5.5°C) was the highest on our records. December's average figure was the lowest on our records at -3.4°C (LTA 1.2°C). The total number of days with air temperature <0°C came to 62, 15 more than the LTA of 47.

Minimum grass temperatures showed mixed results with April, May, and August through to November all displaying higher than expected figures. January to March, June, July and December all showed minimum monthly temperatures below normal. December's LTA figure of -9.2°C was beaten with -11.3°C, but does not compare with the particularly chilly -13.3°C recorded in 2008. The total number of days with grass temperature <0°C (frost) was 125, 9 days fewer than the LTA of 134. Although only August and September escaped frost, most months had fewer nights of frost than expected. December had 31 nights of frost for the first time on our records (LTA 21).

## Sunshine



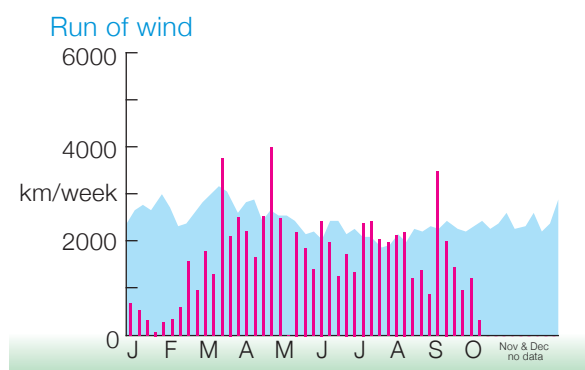
Mean soil temperatures were again higher than the LTA for most months; the exceptions were January and December, where figures were slightly lower than normal.



**Sunshine and solar radiation** Sunshine figures were fairly mixed with a total of 1523.4 hours of bright sunshine, higher than the LTA of 1411.6 hours. January, March through to June, and September all had more hours of sunshine than expected. March logged the highest figures on our records for the second year running. All other months showed lower than normal sunshine figures, the poorest being December. Solar radiation values were again higher than normal with the exception of August.



Dundee Flower & Food Festival, 2009.



**Rainfall** As with 2007 and 2008, 2009 is likely to be remembered for its exceptional rainfall. Once again levels were higher than normal, the annual total reaching 845.9mm as compared to the LTA of 664.5mm. Seven months had greater than normal rainfall – the figures being significantly higher, November produced 147.3mm – three times the LTA of 52.1mm. This was the highest on our records and resulted in major flooding across the country. Also of note is the 65.6mm of rain which fell during a single 24 hour period on the 3rd September. This was more than the LTA for the entire month and again resulted in considerable local flooding and the cancellation of the first day of the Dundee Flower and Food Festival for the first time in its 21 year history.

Thunder was reported three times during the year, twice during July and once on 30th December.

**Wind** On the whole, wind speeds were lower than normal. Worth mentioning are the 10 recorded days of gale (mean wind speeds of 34 knots+) as opposed to 20 in 2008. Also, the unusual occurrence of a funnel cloud was witnessed from SCRI on the 24th August.

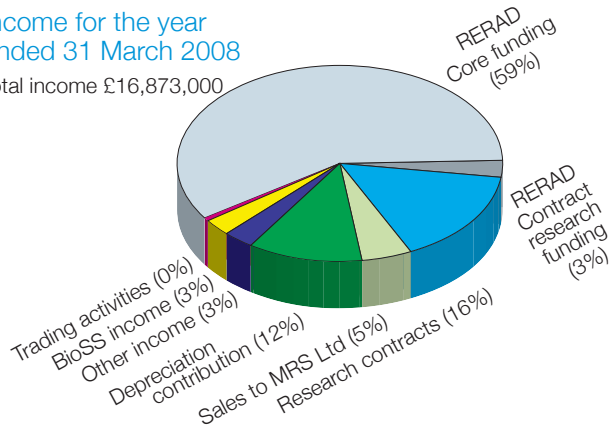
SCRI's weather data is available on our website. You can find a general overview, a data page and a current weather page that displays the present day's data from SCRI's Automatic Weather Station, updated in 15 minute intervals.

Every month, our weather data is featured in the Courier and Dundee Evening Telegraph.

# Accounts

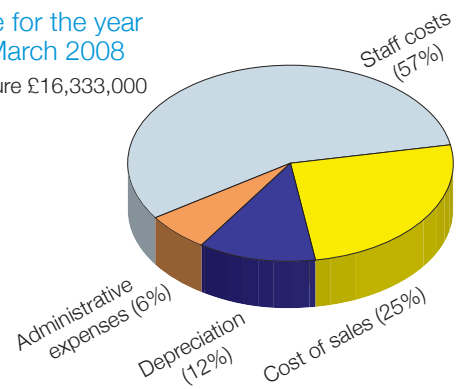
## Income for the year ended 31 March 2008

Total income £16,873,000



## Expenditure for the year ended 31 March 2008

Total expenditure £16,333,000



## Balance sheet at 31 March 2008

Total value £35,204,000

### Assets

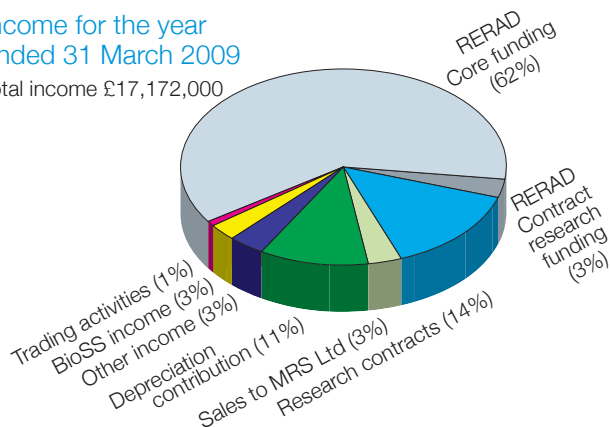
Fixed assets	92 %
Stocks	0 %
Debtors	7 %

### Liabilities

Capital reserve	82 %
Income & expenditure account	7 %
Current liabilities	11 %

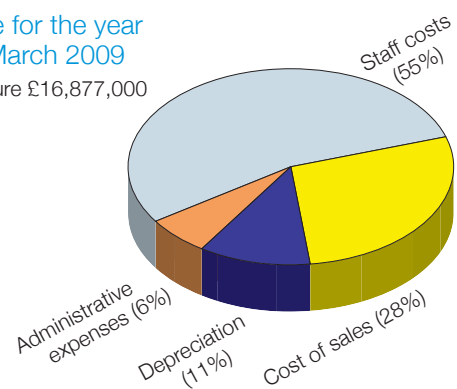
## Income for the year ended 31 March 2009

Total income £17,172,000



## Expenditure for the year ended 31 March 2009

Total expenditure £16,877,000



## Balance sheet at 31 March 2009

Total value £33,655,000

### Assets

Fixed assets	93 %
Stocks	0 %
Debtors	7 %

### Liabilities

Capital reserve	81 %
Income & expenditure account	8 %
Current liabilities	11 %

Copies of the statutory accounts can be obtained from the Director of Finance & Corporate Services, SCRI



# Staff List

as at 1 October 2009

Chief Executive and Institute Director  
Director of Science Co-ordination  
Director of Science Planning  
Director of Finance and Corporate Services

P J Gregory BSc PhD Hon Dr (Debrecen) CBiol FBS FRASE<sup>4,12,13,14,15</sup>  
H V Davies BSc PhD CBiol FBS<sup>1,2,4</sup>  
D W Hopkins BSc PhD CBiol<sup>16,17,20,21,22</sup>  
N G Hattersley BSc PhD ACMA



## Genetics

R Waugh BSc PhD (Programme Leader)<sup>3,4</sup>  
M M Bayer MSc PhD  
N Bonar HNC  
A Booth HNC  
C Booth BSc  
R M Brennan BSc PhD  
G J Bryan BSc PhD  
X Chen BSc PhD  
G P Clark HNC BSc  
J Comadran-Trabal BSc  
D Cullen BSc PhD  
M F B Dale BSc PhD<sup>3</sup>  
D Davidson  
L Donnelly  
A Druka MSc PhD  
I Druka  
J D Fuller  
A Ghatak BSc  
S L Gordon HNC  
J Graham BSc PhD  
B Harrower HND BSc MSc  
P Hedley BSc PhD  
I Hein MSc PhD  
C Hornyik BSc PhD  
K Houston BSc MSc PhD

L Jorgensen HND  
R Keith NEBS  
D Lewandowska BSc PhD  
M S Liney HND  
M E Looseley BSc MSc PhD  
J Lyon  
S McCallum  
S McClatchey  
G McKenzie HND BSc  
K McLean BSc  
M Macaulay HNC BSc  
D F Marshall BSc PhD  
H A Mathews  
J Middlefell-Williams HNC  
I R Milne BSc PhD  
L G Milne BSc PhD  
J Morris HND BSc  
M Myles ONC  
A Prashar BSc MSc PhD  
G Ramsay BSc PhD<sup>3</sup>  
L Ramsay BSc PhD  
J Russell BSc PhD  
S K Sharma BSc PhD  
P D Shaw MSc  
C G Simpson BSc PhD

K Smith DipHE  
P L Smith BSc  
G J Stephen BSc  
J Stephens BSc PhD  
G E L Swan  
J S Swanston BSc PhD CBiol MIBiol  
W T B Thomas BSc PhD  
N Uzrek BSc  
A Weir  
G Wilde  
S L Williamson BSc  
R N Wilson NCH  
M R Woodhead BSc PhD  
J Yildiz HNC BSc  
G R Young HNC

### Mylnfield Research Services

H Bull BSc  
D Coyle  
D Clark  
G Duncan BSc  
L Ferguson  
S N Jennings BSc  
J Robertson  
V Young BSc



## Plant Pathology

L Torrance BSc PhD (Programme Leader)<sup>3</sup>  
A O Avrova BSc PhD  
C J Barker  
V C Blok BSc MSc PhD  
P Boevink BSc PhD  
J L Brierley BSc PhD  
W Burry  
M Burton  
S N Chapman BSc PhD  
P Cock MPhys MSc PhD  
D E L Cooke BSc PhD  
G H Cowan HNC MSc  
A Dolan HNC  
E Douglas BSc  
B Fenton BSc PhD CBiol MIBiol<sup>3</sup>  
G L Fraser  
E Gilroy BSc PhD  
D C Guy HND  
N Holden BSc PhD

A M Holt  
S N Humphris HNC BSc PhD  
J T Jones BSc PhD<sup>3</sup>  
S S Lamond  
A K Lees BSc PhD  
H Liu BSc MSc PhD  
A J Love BSc PhD  
J Lynott BSc  
S A MacFarlane BSc PhD  
W J McGavin BSc  
K D McGeachy HNC  
J McMillan  
G L Malloch DCR BSc PhD  
A C Newton BSc PhD  
P F Palukaitis BSc PhD<sup>1,5,10</sup>  
A J Paterson HND  
Y Pitkin BTec HND  
L Pritchard BSc PhD  
E F O Randall MSc PhD

B Reavy BSc DPhil  
W Ridley  
A G Roberts BSc PhD<sup>3</sup>  
J Shaw BSc  
A Smith BSc  
J N Squires BSc PhD  
J A Stewart HND BSc  
L Sullivan BSc  
M M Swanson BSc PhD  
M Taliany PhD DSc<sup>11</sup>  
J Tilsner BSc PhD  
I K Toth BSc PhD<sup>6</sup>  
R F Van Toor BSc PhD  
E Warden ONC  
S Whisson BSc PhD  
N A Williams HNC  
K M Wright MA PhD  
A Ziegler BSc PhD

<sup>1</sup> Honorary Senior Lecturer in the University of Dundee  
<sup>2</sup> Professor, Universities of Cordoba and Malaga  
<sup>3</sup> Honorary Lecturer, University of Dundee  
<sup>4</sup> Honorary Professor, University of Glasgow  
<sup>5</sup> Adjunct Professor, Cornell University  
<sup>6</sup> Honorary Lecturer, University of Aberdeen  
<sup>7</sup> Honorary Fellow, University of Edinburgh  
<sup>8</sup> Honorary Lecturer, University of Strathclyde

<sup>9</sup> Honorary Professor, Heriot-Watt University, Edinburgh  
<sup>10</sup> Honorary Professor, Seoul Women's University  
<sup>11</sup> Adjunct Professor, Moscow State University  
<sup>12</sup> Visiting Professor, University of Reading  
<sup>13</sup> Honorary Professor, University of Dundee  
<sup>14</sup> Visiting Professor, University of Abertay, Dundee  
<sup>15</sup> Honorary Professor, University of Edinburgh  
<sup>16</sup> Visiting Professor, University of Stirling

<sup>17</sup> Adjunct Professor, University of Canterbury New Zealand  
<sup>18</sup> Special Professor, University of Nottingham  
<sup>19</sup> Visiting Associate Professor, Comenius University Bratislava  
<sup>20</sup> Visiting Professor, Newcastle University  
<sup>21</sup> Honorary Senior Research Fellow, University of Glasgow  
<sup>22</sup> Honorary Fellow, Rothamsted Research



## Plant Products and Food Quality

D Stewart BSc PhD (Programme Leader)  
C Cognat  
S C Conner BSc MSc CChem MRSC  
H V Davies BSc PhD CBiol FBS<sup>1,2,4</sup>  
I N De Abreu BSc PhD  
G Dobson BSc PhD  
P M Dobson  
L J M Ducreux BSc MSc MPhil PhD  
D Grussu  
R D Hancock BSc PhD  
R Hutchison  
E N Kanichukattu  
H Kemp BSc PhD

G J McDougall BSc PhD  
D McRae ONC  
W Meier-Augenstein PhD CChem MRSC  
W L Morris BSc MSc PhD  
S D A Pont BSc  
J Rauscher  
H A Ross HNC PhD CBiol MIBiol  
L V T Shepherd BSc MSc PhD  
T Shepherd BSc PhD  
N Stewart BSc  
J A Sungurtas HND  
M A Taylor BSc PhD  
S R Verrall HNC

P G Walker HND  
J F Wilkie  
[Mylnfield Research Services](#)  
W W Christie MBE BSc PhD DSc FRSE  
K Devlin  
F Gunstone BSc PhD DSc FRSE  
L Hunter BSc  
L Kelly  
S Rowbottom ONC HNC BSc CChem MRSC  
G Sawers BSc AMRSC  
C Scrimgeour BSc MSc  
C Traynor BSc CChem MRSC  
K Wood



## Environment Plant Interactions

P J White BA PhD (Programme Leader)<sup>18,19</sup>  
G Banks BSc MSc  
G S Begg BSc PhD  
A G Bengough BSc PhD<sup>3</sup>  
K Binnie BSc  
A N E Birch BSc PhD CBiol MIBiol FRES<sup>3</sup>  
L K Brown BSc  
S Caul HNC  
T J Daniell BSc PhD<sup>3</sup>  
J Davidson BSc  
L Dupuy MSc PhD<sup>3</sup>  
R Dye  
F Falconer HNC  
L Ford BA

T S George BSc PhD  
D C Gordon HNC  
P D Hallett BSc PhD  
C Hawes BSc PhD  
D W Hopkins BSc PhD CBiol FBS  
R G Hunjan  
P P M Iannetta BSc PGCE(S) PhD  
S N Johnson BSc DPhil  
C Jorna  
A J Karley BA DPhil  
M Krol MSc  
M MacAskill  
B M McKenzie BSc PhD  
C Mitchell BSc

S M Mitchell BSc  
P Neave NC  
R Neilson HNC MSc PhD  
D M Roberts HND BSc PhD  
G R Squire BA PhD  
J A Thompson BSc  
T Valentine BSc PhD  
L Wiesel BSc  
J Wishart BSc PhD  
G M Wright HNC  
M Young HND MSc PG Dip IT  
[Mylnfield Research Services](#)  
S Donn BSc  
R E Wheatley BSc PhD



## University of Dundee, Division of Plant Sciences

J W S Brown BSc PhD<sup>3,4,13</sup> (Head of Division)  
M Alloy-Lleonart  
M Armstrong BSc PhD  
A Barakate BSc PhD  
P R J Birch BSc PhD (Principal Investigator)  
T Bukharova  
P Daly BSc  
A J Flavell BSc PhD (Principal Investigator)  
S Gray BSc  
C Halpin BSc MSc HDip PhD (Principal Investigator)

T Ho  
S F Hubbard PhD (Principal Investigator)  
E Huitema  
H G Jones MA PhD FIHort (Principal Investigator)  
J Kam  
S Y Kim PhD  
C McClellan  
H McLellan  
S Manthri  
J Marshall BSc DPhil  
K Rataj

J A Raven BA PhD HonPhD(Umea) CBiol FBS FRS FRSE  
(Principal Investigator)  
R Shafiei  
G G Simpson BSc PhD (Principal Investigator)  
M Skelly  
M Spensley  
N H Syed BSc PhD  
S Vivera BSc MSc  
Y Wilson BSc  
Y Xiao  
X Xu



## Communication and Information Services

P Taylor MCIPR (Head of CIS)  
P Cassidy  
S E Collier BSc MA MCLIP

U M McKean MA DipLib  
C V Morton BA  
S J Neilson DipBiolSci DipPollCon BSc

I R Pitkethly HND  
E S Ritch  
L Wakefield MA



## Information Technology

S Clark HNC MSc (Head of IT)  
L H Davidson BA  
I Grant HNC

P J R Grimmond BA  
R McCreary BSc  
L A McGregor BSc

V Pandey BSc  
P Smith BSc



## Finance and Corporate Services

N G Hattersley BSc PhD ACMA  
(Director of Finance and Corporate Services)

S Bell

A J Cartwright BA DMS FCIPD

M Dalziel MA PgDPM MCIPD

R G Davidson

P Duncan

F Ferguson BA GradCIPD

S Forsyth

K L Grant BA

B V Gunn

S Inglis

J Keith

C F Kydd FCCA

L Logie HNC MinstAM(AdvDip)

A Pack BA

W Patterson HND

K Robb

F I F Rowe MA DipNEBOSH Grad IOSH EnvDipNEBOSH  
(Safety Quality Environment Manager)

A Sandilands

S Sinclair

M Soutar

E L Stewart

L Young



## Engineering and Maintenance

S Petrie BSc (Head of Engineering and Maintenance)

J Anderson

D Byrne

C Conejo

L A Crichton

G J E Ewart

A G Fox

D Gray HNC

K A Henry

R D McLean

I C McNaughton HNC

G Pugh

D J Redford

G C Roberts

J Rowe

W Scott

B Semple



## Glasshouse and Field Services

P A Gill HND NEBOSH  
(Glasshouse and Field Services Manager)

J Abernethy

P Baird

J R K Bennett

J T Bennett

E Caldwell

M Cook

A G Dobson HNC HND

I Fleming

J M Ford

A C Fuller

M A Grassie HNC BEI

P Heffell ONC

R Keith

M Lipska

J Mason

T A Mason NEBSM

D I Matthew BSc

A W Mills

A D Munro HND

R Ogg

G R Pitkin HND

A M Thain HNC

J K Wilde

D Young



## Mylnfield Research Services

N W Kerby BSc PhD CBiol FBS (Managing Director)

L Beaton HNC DMS MBA

A Ross HNC CPP

J B Snape MA MSc PhD CBiol MIBiol MBA

H Wilson HNC



## Biomathematics and Statistics Scotland

D A Elston BA MSc PhD (Director of BioSS)

King's Buildings, University of Edinburgh

C A Glasbey MA DipMathStats PhD DSc MScI  
(Head of Group)<sup>7,8,9</sup>

A Butler BSc PhD

S Catterall BA MSc PhD

J M Dickson BSc

D Glancy

E M Heyburn MA

D Husmeier BSc PhD

G T Innocent

H R Kettle BSc MSc PhD

M A M Kirkwood DA

A D Mann BSc

G R Marion BSc MSc PhD

I J McKendrick BSc PhD

M Nath BSc MSc PhD

I M Nevison MA

D Nutter BSc

J Polechova BSc MSc PhD

A M I Roberts BSc MSc

Y Song BSc MSc PhD

L Spezia BSc PhD

West of Scotland Unit, Hannah

S Brocklehurst BSc PhD

Environmental Modelling Unit, Macaulay

D A Elston BA MSc PhD (Head of Group)

M J Brewer BSc PhD

E I Duff BSc

J M Potts BSc MSc PhD

Aberdeen Unit, Rowett

G W Horgan BA MSc PhD (Head of Group)

G Holtrop MSc PhD

C D Mayer MSc PhD

Dundee Unit, SCRI

C A Hackett BA DipMathStats PhD (Acting Head of Group)

C Alexander BSc PhD

K M MacKenzie BSc MSc PhD

F G Wright BSc MSc PhD



## Visiting workers

Name	Country of origin	Programme	Month/yr of arrival	Length of stay	Name	Country of origin	Programme	Month/yr of arrival	Length of stay
N Aberdein	UK	PPFQ	Sep-09	5 mth	R Martin	UK	PP	Jul-09	1 mth
A Ahmad	Pakistan	EPI	Apr-09	6 mth	I Martinussen	Norway	PPFQ	Apr-09	2 wks
B Alhusen		GEN	Sep-09	2 wks	L McCormick	UK	UoD	Sep-09	6 mth
M Al-Khairulla	UK	PPFQ	Sep-09	5 mth	F McLeod	UK	EPI	Jun-09	4 mth
A Alliaume	France	EPI	May-09	3 mth	J Molmann	Norway	PPFQ	Apr-09	2 wks
S Apparow	Malaysia	GEN	May-09	3 mth	M Mucci	Italy	PP	Feb-09	4 mth
A Artaux	France	EPI	Aug-09	4 mth	S Mucyo	Rwanda	PPFQ	May-09	4 mth
M Ashfaq	Pakistan	PP	May-09	8 mth	S Nunsavathu	India	PPFQ	May-09	5 mth
A Augustine	India	GEN	May-09	1 yr	M Opik	Estonia	EPI	Feb-09	3 wks
I Badjakov	Bulgaria	PPFQ	Apr-09	3 wks	J Papuchon	France	EPI	Jun-09	3 mth
P Baranowski	Poland	UoD	Feb-09	2 wks	L S Pardo	Spain	PPFQ	Jan-09	4 mth
K Becker	UK	EPI	May-09	3 mth	A Paul	India	EPI	May-09	4 mth
E Blixt	Sweden	PP	Feb-09	4 mth	S Perez Gomez	Spain	GEN	Aug-09	4 mth
A Block	USA	UoD	May-09	3 mth	H Pietrykonska	Poland	GEN	Sep-09	2 wks
J Bruce	UK	EPI	Sep-09	9 mth	B Pindelski	Poland	IT	Feb-09	4 mth
M A R Cervera	Spain	PPFQ/MRS	May-09	3 mth	J Ponchart	France	PP	Jan-09	7 mth
B N Chung	Korea	PP	Apr-09	6 mth	C Primeau	Denmark	PPFQ	Jul-09	2 mth
P Connelly	UK	PPFQ	Sep-09	5 mth	A Przetakiewicz	Poland	PP	Jun-09	2 mth
I Dawson	UK	GEN	Dec-08	7 mth	T Pulickal	India	PPFQ	Apr-09	4 mth
M De Sain	Netherlands	PP	Apr-09	1 mth	L Pylypenko	Ukraine	PP	Jan-09	3 mth
V Delcroix	France	EPI	Jun-09	3 mth	N Raboutdin	France	EPI	Jun-09	3 mth
P Dennis	UK	EPI	Apr-07	3 yrs	D Raczyńska	Poland	GEN	Sep-09	2 wks
A K Dev	India	EPI	Oct-08	5 mth	R Radhakrishnan	India	PP	Apr-09	5 mth
P Dyer	UK	PP	Sep-09	8 mth	M B Ramos	Spain	PP	Oct-08	7 mth
P Dyer	UK	UoD	Jun-09	2 mth	S Reid	UK	UoD	Jun-09	9 mth
J Elder	UK	CIS	Sep-09	2 mth	E Repplinger	France	EPI	Apr-09	2 mth
S Engelhardt	Germany	PP	Feb-09	2 yr	C Rizza	Italy	PP	Oct-08	2 mth
S Ewert	France	GEN	Sep-09	6 mth	L Robertson	UK	PP	May-09	3 wks
H Fankem	Cameroon	EPI	Feb-09	10 mth	K Ross	UK	EPI	Sep-09	7 mth
J Fikowicz-Krosko	Poland	PP	Jul-09	1 mth	R Rothenberger	Germany	PP	Mar-09	6 mth
L Forster	UK	EPI	Jul-09	1 mth	J Russell	UK	PP	Sep-09	9 mth
A Galbraith	UK	UoD	Apr-09	5 mth	B Sable	Canada	EPI	Jan-09	3 wks
P Gandham	India	PP	Aug-09	5 mth	R Sallepalli	India	PPFQ	Apr-09	5 mth
E Gil	Spain	EPI	Jul-09	1 mth	J Samson	UK	CIS	Jan-09	3 mth
A Gill	India	EPI	Dec-08	5 mth	A S Samuel	India	UoD	Sep-09	1 yr
D Gokhale	India	PPFQ	Aug-09	3 mth	M C Scandura	Italy	PPFQ	Jun-09	4 mth
K Goszcz	Poland	PPFQ	May-09	5 mth	V Schnepf	Germany	PP	Sep-09	4 mth
K Graham	UK	UoD	Feb-09	3 wks	S Scibetta	Italy	PP	Oct-08	2 mth
A Gras	France	EPI	Jun-09	3 mth	C Sharma	India	PPFQ	Apr-09	5 mth
D Grussu	UK	PPFQ	Jan-09	5 mth	A Snadden	UK	UoD	Jul-09	9 mth
J Guzy-Wroblewska	Poland	GEN	Feb-09	3 mth	P K Sreekumari	India	PP	May-09	5 mth
C Hale	UK	EPI	Jul-09	2 mth	O Stasik	Ukraine	UoD	May-09	3 mth
M Haq	Bangladesh	UoD	Jul-09	2 mth	C Stushnoff	USA	PPFQ	Oct-08	2 mth
A Hartmann	Germany	PPFQ	Jul-09	3 wks	B Sun	China	EPI	Aug-08	1 yr
J Helps	UK	EPI	May-09	4 mth	J Sun	China	EPI	Jan-09	2 mth
A Hernandez	USA	UoD	Mar-09	9 mth	A Tahir	Pakistan	UoD	Sep-09	1 yr
E K James	UK	EPI	Jan-09	1 yr	H A F A Thahar	India	PP	Apr-09	5 mth
A John	India	PP	May-09	4 mth	Z Tian	China	PP	Nov-08	11 mth
F John	India	EPI	May-09	4 mth	A Tite	France	EPI	Jun-09	3 mth
M Johnston	UK	EPI	Jul-09	2 mth	A Tondelli	Italy	UoD	Apr-09	2 wks
M Kaczmarek	Poland	UoD	Sep-09	1 yr	M Tyagi	Portugal	PPFQ	Sep-09	3 wks
E N Kanichukattu	India	PPFQ	Jun-09	4 mth	E Udugbezi		UoD	Apr-09	5 mth
J Kevrele	UK	PPFQ	Oct-08	6 mth	C Vagne	France	GEN	Jan-09	5 mth
T Kikuchi	Japan	PP	Oct-08	4 mth	Y Valdes	Cuba	PP	Jan-09	1 mth
P Kongprakhon	Thailand	GEN	Mar-09	4 mth	V Vasukuttan	India	PPFQ	Dec-08	4 mth
A Kumar	India	PPFQ	Apr-09	5 mth	A Vaughan	UK	EPI	Jun-09	1 mth
S Kurian	India	PP	May-09	6 mth	V Vendramin	Italy	GEN	Apr-09	1 mth
B Kusenda	Slovakia	GEN	Oct-08	2 wks	A Visiani	Spain	GEN	Feb-09	3 mth
S Labibida	Syria	GEN	Jul-09	3 mth	U Vrhovsek	Italy	PPFQ	Feb-09	3 wks
R Lamarque	France	PP	Jun-09	2 mth	N Walker	UK	EPI	Sep-09	6 mth
C Lambert	France	EPI	Jun-09	2 mth	R Wenchel	Germany	PP	Feb-09	3 mth
J Lamont	UK	UoD	Sep-09	1 yr	L Whiteside	UK	UoD	Sep-09	6 mth
A Landau	Argentina	GEN	May-09	2 mth	J Wilson	UK	PP	Sep-09	9 mth
T Lemmermeyer	Germany	PP	Oct-08	1 mth	X Wu	China	EPI	Nov-08	10 mth
M Lisowska	Poland	PP	Jun-09	2 mth	C Y Yang	Korea (South)	PP	Jul-09	3 mth
M Liu	China	EPI	Apr-09	2 wks	W Yang	China	PPFQ	Apr-09	5 mth
V Lund	UK	PPFQ	Dec-08	2 mth	C Yule	UK	CIS	Sep-09	3 wks
M Maronova	Slovakia	GEN	Oct-08	2 wks	Z Zhang	China	GEN	Oct-08	9 mth
P Martin	France	EPI	Mar-09	5 mth					





## Postgraduate students

Name	Programme	Project Title
Aqueel Al-Abedy	PP	Studies on <i>Potyvirus</i> resistance in <i>Solanum Phureja</i> .
Ahmad Al-Mrabeh	PP	<i>Potato Virus Y</i> and aphid transmission.
Ananthi Anandanadesan	UoD/EPI	Mathematical modelling of the spatio-temporal dynamics of aphid-parasitoid-plant virus interactions.
Sandra Bacon	PPFQ	Potential of the action of novel nutrient regulators of the longevity factor FOXO1a by the trace metal zinc.
Gillian Banks	EPI	Dynamics for feral oilseed rape populations and the impacts on associated insect communities.
Nathalie Bernaert	PPFQ	Metabolite profiles of leek in function of genetic variation, breeding and processing techniques.
Rachel Berry	PPFQ	The real uncertainty of stable isotope data for provenancing of humans and drugs.
Susan Breen	PP	Translocation of effector proteins into host plants.
Hazel Bull	GEN	Development and characterisation of a meta-population of <i>Hordeum spontaneum</i> introgression lines into cultivated barley.
Andrew Burgess	UoD/EPI	Mathematical modelling of disease dynamics in host-parasitoid systems.
Raymond Campbell	PPFQ	Genetics of carotenoid levels in potato tubers.
Alison Chapman	PP	The changing <i>Phytophthora infestans</i> population: implications for late blight epidemics and control.
Xiaoyun Chen	EPI	Biodiversity and soil food web activity in different managed grasslands.
Emily Clark	EPI	Multitrophic factors influencing aphid vector competence in a spatially heterogeneous environment.
Katy Clark	EPI	Does mother know best? Is host plant selection by above ground insects influenced by below ground herbivores?
Hannah Clarke	UoD/EPI	The role of bacterial secondary symbionts of <i>Macrosiphum euphorbiae</i> in the dynamics of multitrophic interactions.
Emma Coates	PPFQ	Anti-cancer effects of soft fruit phytochemicals.
Nicky Cook	UoD/EPI	Population genetics of fernland sawflies.
Marion Cubitt	PP	A systems approach to virulence regulation in <i>Pectobacterium</i> .
Paul Daly	UoD/GEN	Manipulation of lignin biosynthesis in barley.
Jayne Davis	EPI	Molecular interactions of potassium deficiency and pathogen resistance in barley.
Sherine Deakin	PPFQ	Antioxidant effects of phytochemicals in conditions of oxidative stress: impact on endothelial cell survival and function.
Wenni Deng	EPI	Engineering novel geotextiles from an understanding of the dynamic properties of seed coat mucus.
Ralf Dietrich	UoD/EPI	Root soil dynamics: New ways of studying roots in a changing climate.
Helen Downie	EPI	3D imaging of root particle interactions using optical projection tomography and x-ray microtomography.
Natasha Duckett	EPI	Soil reinforcement using plant roots: A sustainable alternative to traditional slope stabilisation.
Daniel Dzidzienyo	GEN	Manipulation of self-incompatibility in diploid potato species.
Alexandre Foito	PPFQ	Development of metabolomics based methods to benefit marker assisted breeding in perennial ryegrass.
Fiona Fraser	EPI	Temperature responses of soil nitrogen transformations.
Joanna Fyans	UoD/PP	The role of protein transport in the pathogenicity of <i>Streptomyces spp.</i>
Madeline Giles	EPI	Where does denitrification occur in the rhizosphere?
Varun Gopalakrishnan	PP	Acquisition and transmission of Potato mop top virus by <i>Spongopora subteranea</i> .
Clement Gravouil	PP	Multiple disease interactions on cereals and interactions with other crop environment organisms.
Severine Grouffaud	PP	Translocation of <i>Phytophthora</i> effectors and their manipulation of host plant disease resistance.
Miriam Herold	EPI	Plant roots as drivers of denitrification.
Gerald Hochshartner	GEN	Application of a novel snoRNA marker system in plant evolution and systematics.
Lydia Hunter	PP	Role of potato RNA-dependent RNA Polymerase 1 in virus resistance.
Ning Jiang	GEN	Linkage disequilibrium based mapping of complex traits in crops.
Florian Jupe	GEN	Utilising next generation sequencing to clone and characterise durable potato resistance genes.
Angzzas Kassim	GEN	Genetic and environmental control of anthocyanin and volatiles in red raspberry.
Pavel Kerchev	PPFQ	Influence of ascorbic acid on plant aphid interactions.
David Lloyd	GEN	Mapping genes and QTLs linked to flavour and texture in potatoes.
Kenneth Loades	EPI	Quantifying the role of fibrous roots on soil reinforcement.
Marta Maluk	UoD/GEN	Improving barley for biofuel production – investigating the roles of CCR and 4CL in lignin biosynthesis.
Susan McCallum	GEN	Linking phenotype to genotype for fruit quality traits in raspberry.
Lindsay McMenemy	EPI	Cooperation between plant enemies – do raspberry viruses attract more aphid vectors?
Stefania Pasare	PPFQ	The regulation of isoprenoid metabolism in <i>Solanaceae</i> crop plants.
Katarzyna Rataj	UoD/GEN	Functional characterisation of the protein interaction domain of FPA.
Namrata Reetoo	UoD/GEN	Controlling <i>Arabidopsis</i> RNA binding proteins with phosphorylation.
Christelle Robert	BioSS	Prediction of promoter binding sites and transcriptional regulatory networks in two gamma-proteobacterial pathogen genomes.
Sonja Schmidt	EPI	Microtomography in soil root interactions.
Zulkifli Ahmad Bin Seman	GEN	Functional and evolutionary characterisation of host genes targeted by <i>Phytophthora infestans</i> effectors and defence associated candidate genes.
Jane Shaw	PP	The role of Cajal bodies in the nucleolus in plant virus infection.
Marcin Skiba	EPI	Biological nitrification inhibition in the rhizosphere of temperate arable plants.
Nithya Subramanian	GEN	Increasing mineral delivery through potatoes.
Hui Tan	PP	Dissection of the <i>Pectobacterium</i> virulence quorum sensing regulation.
Rosalind Mary Taylor	PP	Ubiquitin-proteasome directed proteomic approach to dissect biotic stress signalling in plants.
Amar Thirugnanasambandam	PP	Role of seed-borne infection in <i>Rhynchosporium</i> and <i>Ramularia</i> epidemics in barley.
Maja Thorsen	EPI	Biological mechanisms involved in stabilising sandy soils of the machair.
Stephanie Vink	EPI	Functional soil ecology and conservation in the machair in relation to changing land management.
Lea Wiesel	EPI	Caesium uptake by roots of <i>Medicago</i> plants.
Tobias Wojciechowski	GEN	Root development in semi dwarfing lines of wheat and barley.
Han Xiao	PPFQ	Function of polyphenols in chemoprevention.
Vanessa Young	MRS	Genetic analysis in a commercially relevant cross and application of markers in a tetraploid breeding programme.
Dzeti Zait	GEN	Sensory and compositional analysis of flavour in red raspberry.
Monica Zwirek	UoD/GEN	Improving barley for biofuel production – efficient transformation for lignin manipulation.



## Honorary Research Fellows

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Professor Richard Abbott BSc PhD  
Dr Pamela Anderson PhD  
Dr Hugh Barker BSc PhD  
Professor Geoffrey J Barton BSc PhD  
Professor Jim Beynon BSc PhD  
Professor Stephen Blackmore BSc PhD FLS CBiol FBiol FRSE  
Professor Mike Blatt BSc PhD FRSE  
Dr Meredith Bonierbale PhD  
Dr Fraser Bransby BA MA PhD  
Dr William W Christie MBE BSc PhD DSc FRSE  
Professor Sir Philip Cohen FRS FRSE  
Professor Pierre de Wit PhD  
Professor Pete Downes OBE FRSE MIBiol PhD  
Dr Jim M Duncan MBE BSc PhD  
Dr Roger Ellis BSc PhD  
Dr Andy Flavell BSc PhD  
Professor Geoff Gadd BSc PhD DSc FBS FLS  
Professor Mary Gibby BSc PhD FRSE FRSA  
Professor Frank Gunstone BSc PhD DSc FRSC FRSE  
Professor Claire Halpin BSc MSc H Dip PhD  
Professor Bryan Harrison CBE BSc PhD DAgFor FRS FRSE  
Dr John Hayes BSc PhD  
Professor John Hillman BSc PhD DSc FLS CBiol FBS FIHort FCMi SHM FRAGS FRSE  
Dr Pete Hollingsworth BSc PhD

Dr Steve Hubbard MSc DIC(London) DPhil (Oxon)  
Professor Lindsey Innes OBE BSc PhD DSc FRSE  
Professor Hamlyn Jones MA PhD FIHort  
Professor Peter Langridge BSc PhD  
Dr William H Macfarlane Smith BSc PhD CBiol MIBiol FIMgt  
Professor Gordon Machray BSc PhD  
Mr George R Mackay MBE BSc MSc CBiol FBS  
Dr Donald K L MacKerron MBE BSc PhD  
Professor Tom Meagher BA PhD  
Professor Andrew J Millar BA PhD  
Professor Hugh Nimmo MA PhD FRSA  
Professor Karl Oparka BSc PhD  
Dr Michel Pérombelon MBE PhD MSc BSc Dip Agric  
Professor Wayne Powell BSc MSc PhD DSc  
Professor John Raven BA PhD HonPhD(Umea) CBiol FBS FRS FRSE  
Dr David J Robinson MA PhD  
Professor David J Robinson BSc PhD  
Professor Karel Schubert PhD  
Professor Janet Sprent OBE BSc DSc PhD ARCS FRSE  
Dr Alyson Tobin BSc PhD  
Dr Pieter van West MSc PhD  
Professor Roberto Viola DipAgrSci PhD  
Mr David Walker OBE  
Dr Brian Williamson BSc MSc PhD DSc

# SCRI Research Programme

ongoing as at 1 October 2009

The research programme is commissioned by RERAD (Scottish Government: Rural and Environment Research and Analysis Directorate) and a variety of other funders. The list contains the body that awarded the grant and the title of the project and, in the case of RERAD, the commissioning number.

## RERAD Core – Programme 1

Workpackage 1.1	Barley Genetics
Workpackage 1.2	Potato Genetics
Workpackage 1.3	Soft Fruit Genetics & Pathology
Workpackage 1.4	Barley Pathology
Workpackage 1.5	Potato Pathology
Workpackage 1.7	Sustainable Crop Systems

## RERAD Contract Research Funding

SCR/849/08	Support for additional knowledge exchange activities.
SCR/918/07	Genomics-assisted dissection of barley morphology and development.
SCR/919/07	<i>Dickeya dianthicola</i> – a threat to Scottish seed potatoes.
SCR/921/09	Cell wall lignin programme: manipulating lignin to improve biofuel conversion of plant biomass.
SCR/927/09	Emerging threat to Scottish potato production posed by <i>Dickeya solani</i> .
BSS/037/08	Scottish badger survey.
BSS/845/06	Epidemiology, population, health and infectious disease control.

## External research contracts

Biotechnology and Biological Sciences Research Council (CSI)	Optimising wheat grain shape for improved processing quality.
BBSRC (CSI)	Exploiting the <i>Phytophthora infestans</i> genome to identify gene targets for sustainable potato protection.
BBSRC (CSI)	The establishment and application of a forward genetic resource for the development of efficient breeding strategies in grass and cereals.
BBSRC LINK	Association genetics of UK elite barley.
BBSRC LINK	Role of inoculum sources in <i>Rhynchosporium</i> population dynamics and epidemics on barley.
BBSRC (via University of Dundee)	LoLa: What are the roles of oomycete RXLR effectors in the establishment of plant disease?
BBSRC (via University of Birmingham)	LoLa: Meiosis in barley: manipulating crossover frequency and distribution.
Bioforsk	Systems experiments.



Bioforsk	Plant metabolites for healthy plants and healthy people.
British Council	Analysis of the interaction between the pine wood nematode <i>Bursaphelenchus xylophilus</i> and the maritime pine <i>Pinus pinaster</i> .
British Ecological Society	Climate change in the underworld: how elevated CO <sub>2</sub> affects root–herbivore interactions.
Carnegie Trust	Trophic cascades in a changing climate: the impact of elevated CO <sub>2</sub> on above– and below–ground interactions.
Carnegie Trust	The role of bacterial secondary symbionts of cabbage aphids on aphid-parasitoid interactions in Scotland.
Centre for Ecology & Hydrology (via Department for Environment Food and Rural Affairs (Defra))	Use of 'UKCIP08 scenarios' to determine the potential impact of climate change on the pressures/threats to soils in England and Wales.
Commercial	Brassica breeding.
Commercial	Soft fruit analysis.
Commercial	Potato breeding.
Commercial	Blackcurrant breeding.
Commercial	Lipid analysis.
Commercial	Barley analysis.
Commercial	Soil and water conservation.
Defra/Home-Grown Cereals Authority (HGCA)	GREENGRAIN: Genetic reduction of energy use and emissions of nitrogen in cereal production (LINK).
Defra LINK (via ADAS)	TRAMLINES: Practical cost-effective techniques to reduce pollution from tramlines in combinable crops: a field and catchment scale evaluation.
Defra LINK (via SAC)	CORACLE: Control of Ramularia leaf spot in a changing climate.
Defra/RERAD	Integrated pest and disease management for high quality protected raspberry production.
Estacion Experimental de Aula Dei, CSIC, Spain	Immunogold localisation of antioxidant proteins in legume nodules.
European Union (EU)	EURASNET: European alternative splicing network.
EU	EU-SOL: High quality solanaceous crops for consumers, processors and producers by exploration of natural biodiversity.
EU	BIOEXPLOIT: Exploitation of natural plant biodiversity for the pesticide-free production of food.
EU	DEVELONUTRI: Development of high throughput approaches to optimise the nutritional value of crops and crop-based foods.
EU	GENBERRY: European small berries.
EU	SPICY: Smart tools for prediction and improvement of crop yield.
EU	<i>Triticeae</i> Genome: Genomics for <i>Triticeae</i> improvement.
EU	PLAPROVA: Plant production of vaccines.
EU	BrainHealthFood: Bioactive compounds from blackcurrant processing waste for brain health.
EU	Meiosys: Systematic analysis of factors controlling meiotic recombination in higher plants.
EU Marie Curie	Role of arbuscular mycorrhizal fungi on the accumulation of radiocaesium by plants.
EU Marie Curie	Managing insecticide resistance in aphids in Scotland.



European Science Foundation	Exploiting genomics to understand plant-nematode interactions.
Food and Environment Research Agency	Identification and reduction of risks posed by Potato Ring Rot disease to the Scottish potato industry.
French National Institute for Agricultural Research (INRA)	Global change at the microbial scale – effects of climate change on microbial decomposition and modulation of physical structure of soil.
HGCA	Fungicide performance information for barley growers.
Horticultural Development Council (HDC)	Developing techniques to manage raspberry leaf and bud mite in tunnel produced raspberry.
International Maize and Wheat Improvement Center CIMMYT	Genomic dissection of tolerance to drought stress in wild barley.
Medical Research Council (via University of Dundee)	New geotechnical approaches to soil biological processes.
Ministerio de Educacion y Cienca (CICYT) in Spain	Study of the effect of the treatment with methyl jasmonate on the bioformation of chiral volatile compounds in vegetal foods.
Nairns	Knowledge transfer partnership.
Natural Environment Research Council (via University of Stirling)	Microbial diversity in Antarctic soils.
Norwegian Research Council	Metabolic profiling of <i>Rubus</i> (cloudberry and raspberry): effect of inheritance and environment on phytochemicals beneficial to human health and the identification of viable targets for nutritional enhancement.
Potato Council Ltd	Independent variety trials.
Potato Council Ltd	GB late blight populations: monitoring and implications of population changes.
Potato Council Ltd	Informing management of potato diseases through epidemiology and diagnostics.
Potato Council Ltd	Aphids and virus transmission in seed potato crops.
RERAD	Whole genome and interaction transcriptome sequencing of the barley pathogen <i>Rhynchosporium secalis</i> . (SCR/922/08)
RERAD	Study of the epidemiology of <i>Phytophthora ramorum</i> and <i>Phytophthora kernoviae</i> in managed gardens and heathland in Scotland.
RERAD/Defra/Potato Council Ltd	Sequencing the ‘gene space’ of potato chromosome IV, comparative analysis with tomato and development of a gene-based mapping platform. (SCR/920/08)
RERAD/GSK (HortLink)	Development of high profile germplasm for UK production of blueberries.
RERAD/HDC (HortLink)	Developing breeding and selection tools to reduce spoilage of soft fruits and wastage in the supply chain.
RERAD/Rothamsted International LINK	Producing low acrylamide risk potatoes.
Rothamsted International	Selecting indigenous phosphate solubilising fungi ( <i>Aspergillus</i> , <i>Penicillium</i> ) for crop improvement in nutrient deficient soils of Cameroon. (African Fellowship)
Royal Society	Genetic control of lateral root development stimulated by rhizosphere nematodes.
Royal Society	Transcript profiling of meristem activation in potato using the POCI microarray.
Royal Society	A novel caspase-like protease for plant programmed cell death (PCD).
Royal Society of Edinburgh	RSE/Scottish Executive Personal Fellowship.



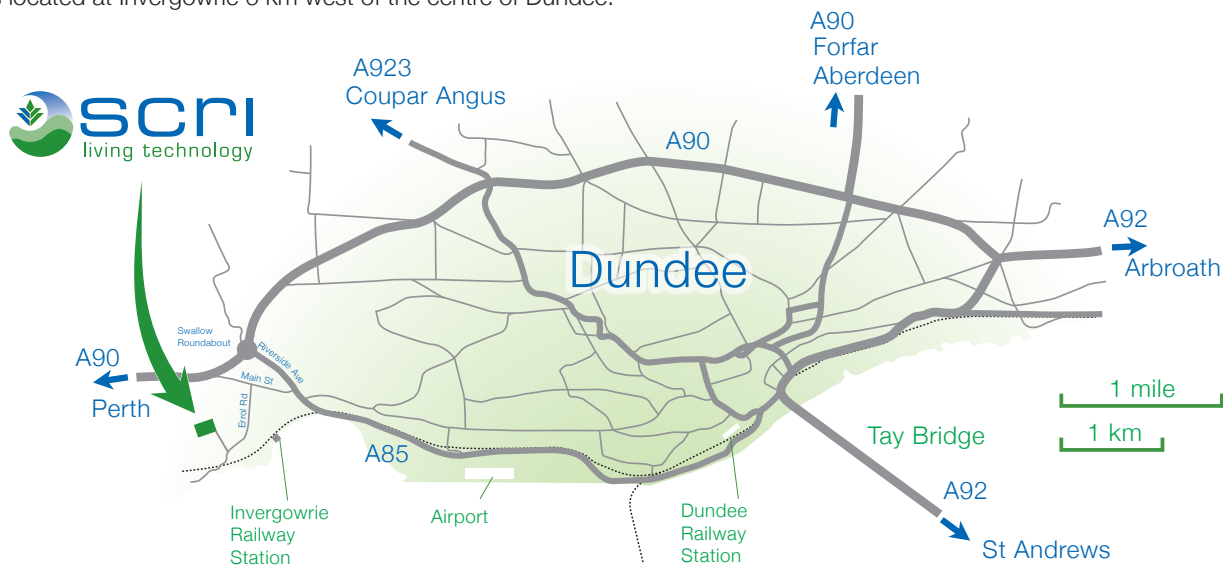
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Rural Development Administration of Korea	Development of manipulation techniques of plant viruses for the development of multiple virus resistant horticultural crops.
Scottish Enterprise	Additive-free treatments for shelf-life extension of minimally processed foods.
Scottish Enterprise	Improving transformation efficiency in barley using a novel method that protects <i>Agrobacterium</i> -mediated gene transfer from attack by caspases.
Scottish Enterprise	A platform enabling technology to characterise plants for their therapeutic potential: anti-inflammatories from <i>Capsella</i> as a model.
Scottish Funding Council (SFC)	Scottish Bioinformatics Research Network (SBRN) maximising bioinformatics infrastructure for Scottish Health, Agriculture and Industry.
Swedish University of Agricultural Science	Characterisation of efficient symbionts via microbial isolation and root nodule structure: a comparison of N <sub>2</sub> -fixing legumes from an island (Scotland) with those from a continental ecosystem (Sweden).
Teagasc	Development of metabolomics based methods to benefit marker assisted breeding in perennial ryegrass.

## Directions to SCRI

SCRI is on the east coast of Scotland, midway between Edinburgh and Aberdeen.

It is located at Invergowrie 6 km west of the centre of Dundee.



### By road

**From Dundee:** Leave the city in a westerly direction along Riverside Drive and Riverside Avenue, towards Perth (A85). Take the left hand turn into Invergowrie, continue past the shops and the Post Office then turn left into Errol Road, which is signposted for SCRI. Follow the road round a sharp right hand bend. The entrance to SCRI is marked with a sign at the foot of the drive on your right.

**From Aberdeen:** Take the A90 south to Dundee, following the Kingsway around the city in the direction of Perth to the Swallow/Landmark Hotel roundabout. Turn left into Riverside Avenue (signposted for the city centre A85) and take the next on the right signposted for Invergowrie. Follow Main Street past the shops and the Post Office taking the next left into Errol Road, signposted for SCRI. Follow the road round a sharp right hand bend. The entrance to SCRI is marked with a sign at the foot of the drive on your right.

**From Perth:** Take the A90 in an easterly direction to Dundee to the roundabout at the junction with the A85. Turn right into Riverside Avenue (signposted for the city centre A85) and take the next on the right signposted for Invergowrie. Follow Main Street past the shops and the Post Office taking the next left into Errol Road,

signposted for SCRI. Follow the road round a sharp right hand bend. The entrance to SCRI is marked with a sign at the foot of the drive on your right.

### By bus

Invergowrie is served by the 16 and 77 bus routes. There are bus stops on Main Street, Station Road and Errol Road and there is a footpath to SCRI between the houses on Errol Road. For information on travelling in Dundee visit the Dundee Travel Information website. For bus timetables and information see the Stagecoach Bus website.  
[www.dundee-travelinfo.com](http://www.dundee-travelinfo.com)  
[www.stagecoachbus.com](http://www.stagecoachbus.com)

### By train

Regular trains from Glasgow, Edinburgh, London and other UK cities run to Dundee. Infrequent trains stop at Invergowrie  
[www.nationalrail.co.uk](http://www.nationalrail.co.uk)

### By air

Air France runs regular flights between London City and Dundee airports as part of its CityJet service. Flybe flies to Belfast City and Birmingham International airports from Dundee.  
[www.airfrance.co.uk](http://www.airfrance.co.uk)  
[www.flybe.com](http://www.flybe.com)





